

**THE CLINICAL EFFECTIVENESS OF PROBIOTICS AND
AUTOPROBIOTICS IN TREATMENT OF *HELICOBACTER PYLORI*-
ASSOCIATED DYSPEPSIA**

Ermolenko E.I.^{a,b},

Molostova A.S.^{a,g},

Baryshnikova N.V.^{a,c,d},

Svarval A.V.^e,

Gladyshev N.S.^{e,f},

Kashchenko V.A.^{f,g},

Suvorov A.N.^{a,f}

^aThe Federal State Budgetary Scientific Institution 'Institute of Experimental Medicine' 197376, Akademika Pavlova street, 12A, St. Petersburg, Russia

^bNorth-Western State Medical University named after I.I.Mechnikov, 191015, 41, Kirochnaya street, St. Petersburg, Russia

^cPavlov First Saint Petersburg State Medical University, 6-8 Lva Tolstogo street, 197022 St. Petersburg, Russia.

^dSt.Petersburg State Pediatric Medical University, 2 Litovskaya street, 194100, St. Petersburg, Russia

^eSt. Petersburg Pasteur Institute, 197101, 14 Mira street, St. Petersburg, Russia

^fSaint Petersburg State University, 199034, Universitetskaya nab., 7-9, St. Petersburg, Russia

^gNorth-Western district scientific and clinical center named after L.G.Sokolov Federal Medical and Biological Agency, 194291, 4 Kultury Pr., St. Petersburg, Russia

ЭФФЕКТИВНОСТЬ ПРОБИОТИКОВ И АУТОПРОБИОТИКОВ В МОНОТЕРАПИИ ДИСПЕПСИИ, АССОЦИИРОВАННОЙ С ИНФЕКЦИЕЙ *HELICOBACTER PYLORI*

Ермоленко Е.И.^{1,2},

Молостова А.С.^{1,7},

Барышникова Н.В.^{1,3,4},

Сварваль А.В.⁵,

Гладышев Н.С.^{5,6},

Кащенко В.А.^{6,7},

Суворов А.Н.^{1,6}

¹ФГБНУ «Институт экспериментальной медицины», 197376, Санкт-Петербург, ул.Академика Павлова, 12А

²ФГБОУ ВОСЗГМУ им И.И.Мечникова, 191015, Санкт-Петербург, ул.Кирочная, д.41

³ФГБОУ ВО ПСПбГМУ им. акад. И.П.Павлова, 197022, Санкт-Петербург, ул.Льва Толстого, 6-8

⁴ФГБОУ ВО СПбГПМУ, 194100, Санкт-Петербург, ул.Литовская, 2

⁵ФБУН Научно-исследовательский институт эпидемиологии и микробиологии имени Пастера, 197101, Санкт-Петербург, ул.Мира, д. 14

⁶ФГБОУ ВПО Санкт-Петербургский государственный университет, 199034, Санкт-Петербург, Университетская наб., д. 7–9

⁷ФГБУ «Северо-Западный окружной научно-клинический центр имени Л.Г.Соколова», 194291, Санкт-Петербург, Проспект культуры, 4

Abstract.

The aim of our study was to evaluate the clinical performance of a monotherapy by *Enterococcus faecium*-based probiotics and indigenous autoprobiotics against *H. pylori* associated dyspepsia.

Materials and Methods. There were examined 95 patients with dyspepsia. The entire patient cohort underwent clinical evaluation including filling out the questionnaire to assess dyspepsia symptoms before and after treatment, gastric endoscopy as well as gastric multi-focal biopsy (gastric body and gastric antrum) and verification of *H. pylori* infection with the three clinical laboratory methods (biochemical, bacteriological and molecular detection). An antagonistic in vitro activity of probiotics against *H. pylori* was detected by drop plate method for probiotic strains *Enterococcus faecium* SF68 and *Bifidobacterium bifidum* (Bifiform), *Enterococcus faecium* L3 (Laminolact), and autoprobiotic strains combined with indigenous *Enterococcus faecium*. To examine an antagonistic activity of probiotics and autoprobiotics in clinical trials, we used a starter culture based on the *Enterococcus faecium* L3 strain and an autoprobiotic based on indigenous *Enterococcus faecium*. The probiotic or autoprobiotic were administered orally to patients with gastritis twice a day at dose of 50 ml (8.0 lgCFU/ml) for 20 days. *H. pylori* eradication was assessed by stool antigen test 1.5-2 months after the end of treatment.

Results. Initially the *H. pylori* infection was confirmed with 49.4% of patients. The sensitivity of *H. pylori* to the probiotics was detected in 81% of individuals for indigenous *Enterococci* (the autoprobiotic), 76% - for Laminolact, and in 62% - for Bifiform. 22 patients with previous history of allergic reactions to antibiotics used in routine *H. pylori* eradication regimens were divided in two cohorts. One cohort (10 patients) received the autoprobiotic only, another cohort (12 patients) received only probiotic. Monotherapy with autoprobiotic resulted in 100% *H. pylori* eradication, single-agent therapy with probiotic led to 60%

eradication of *H. pylori*. Dyspepsia symptoms were completely resolved in both groups of patients.

Conclusion. Our research demonstrated the sensitivity of examined *H. pylori* strains to be similar for traditional eradication treatment agents (antibiotics) and the proposed intervention agents (probiotics and autoprobiotics). An autoprobiotic monotherapy with indigenous enterococci led to higher levels of *H. pylori* eradication than with *E. faecium* L3-based probiotic agent. Our work demonstrated advantage for application of probiotics in patients with antibiotic allergies or other obstacles for the standard eradication therapy. Nonetheless, further investigation to better understand underlying mechanisms of action, as well as larger observational and randomized studies, are necessary to determine the scope of therapeutic application for probiotics and autoprobiotics to eradicate *H. pylori* infection.

Key words: *Helicobacter pylori*, eradication, probiotics, autoprobiotics, enterococci, *Enterococcus faecium*

Резюме.

Цель исследования: оценка эффективности пробиотиков на основе энтерококков и индигенных энтерококков (аутопробиотиков) в монотерапии диспепсии, ассоциированной с *Helicobacter pylori*.

Материалы и методы. Мы провели обследование 95 пациентов, страдающих диспепсией. Обследование включало в себя опрос для оценки жалоб до и после лечения, фиброгастродуоденоскопию (ФГДС) с взятием биоптатов из тела и антрального отдела желудка для верификации инфекции *H. pylori* (биохимический, бактериологический и молекулярно-генетический метод). Для исследования антагонистической активности капельным методом в системе *in vitro* использовали пробиотики бифидоформ (*Enterococcus faecium* SF68 и *Bifidobacterium bifidum*) и ламинолакт

(*Enterococcus faecium* L3), аутопробиотик на основе индигенного *Enterococcus faecium*. Для исследования антагонистической активности *in vivo* использовали пробиотическую закваску на основе штамма *Enterococcus faecium* L3 и аутопробиотик на основе индигенного *Enterococcus faecium* (патент РФ №2546253). Препараты назначали per os дважды в день по 50 мл (8,0 IgКОЕ/мл) на 20 дней. Контроль эрадикации проводился с использованием определения антигена микроорганизма в кале через 1,5-2 месяца после окончания лечения.

Результаты. Инфекция *H. pylori* была выявлена у 49,4 % пациентов. Определена чувствительность изолятов микроорганизма к индигенным энтерококкам (аутопробиотику) в 81%, ламинолакту - 76% и бифиформу - 62% случаев. Часть обследованных получала в качестве монотерапии пробиотик или аутопробиотик (пациенты с указанием в анамнезе на аллергические реакции на прием антибиотиков, используемых в схемах стандартной эрадикационной терапии). При использовании аутопробиотика элиминация возбудителя составила 100%, при использовании пробиотика – 60%. Купирование симптомов диспепсии было полным как при приеме пробиотика, так и аутопробиотика.

Заключение. Чувствительность исследуемых штаммов *H. pylori* к аутопробиотику и пробиотикам сравнима с чувствительностью микроорганизма к часто используемым в схемах эрадикации антибиотикам. Монотерапия аутопробиотиком на основе индигенных энтерококков показала более высокий процент элиминации возбудителя, чем применение закваски на основе штамма *E. faecium* L3. В случае невозможности использования стандартной антихеликобактерной терапии назначение как пробиотиков, так и аутопробиотиков является обоснованным. Однако необходимы дальнейшие исследования для расширения доказательной базы оценки эффективности препаратов на основе энтерококков в эрадикации *H. pylori*.

Ключевые слова: *Helicobacter pylori*, эрадикация, пробиотики, аутопробиотики, энтерококки, *Enterococcus faecium*

1 **Introduction**

2 Since the discovery of the role of *Helicobacter pylori* infection in the
3 development of various diseases, particularly peptic ulcer and chronic gastritis, there
4 has been a continuous search for improved methods of eradication of this
5 microorganism. One potential way to improve anti-*H. pylori* treatment regimens is
6 to include probiotics — medications (live microorganisms) that are used to improve
7 the gut microbiota. An emerging need for new treatment agents for *H. pylori*
8 eradication is growing in importance on the grounds of: 1) a decrease in the
9 effectiveness of standard anti-*H. pylori* therapy due to an increase in *H. pylori*
10 resistance to antibiotics, 2) side effects of proton pump inhibitors and antibacterial
11 drugs, 3) reluctance of patients to take antibiotics [7]. Both international and Russian
12 treatment guidelines allow for the use of probiotics. Both the fourth and fifth editions
13 of the Maastricht Consensus Report state that some probiotics and prebiotics may be
14 an effective supplement to standard eradication therapy [16, 17]. The clinical
15 guidelines of the Russian Gastroenterological Association on the treatment of *H.*
16 *pylori* infection in adults state that including probiotics in anti-*H. pylori* therapy
17 improves therapy success and reduces the incidence of adverse events, namely
18 remove the risk of *C. difficile*-associated diarrhea [2]. The VI Moscow Consensus
19 of the Gastroenterological Scientific Society of Russia on the management of
20 patients infected with *H. pylori* also emphasized that anti-*H. pylori* treatment is most
21 effective and safe when supplemented with prebiotics or probiotics [3].

22 A number of meta-analyses demonstrated that the use of probiotics in addition
23 to standard anti- *H. pylori* therapy improves both the effectiveness of eradication
24 and reduces the frequency of side effects [15, 18, 20, 22, 24].

25 In addition, reduction of the side effects incidence of standard eradication
26 therapy, some probiotics may have an antagonistic effect on *H. pylori* by inhibiting
27 the growth of the microorganism. The underlying mechanism of described inhibition
28 might be driven by producing antimicrobial products (bacteriocins, lactic acid,
29 hydrogen peroxide and other) or by competing for survival (through colonization

30 resistance) [6]. This prompted studies to evaluate the effectiveness of probiotic
31 monotherapy in the treatment of *H. pylori* infection. This kind of therapy can be
32 recommended for people who have allergic reactions to antibiotics, who are non-
33 compliant to antibiotic therapies, as well as for family members of patients infected
34 with *H. pylori*.

35 There are many of both Russian and foreign studies confirming the promising
36 positive results of using probiotics monotherapy to eradicate *H. pylori*, with efficacy
37 varying from 6 to 48% [6, 9, 10, 11, 13, 14 ,19]. Probiotics are an emerging
38 promising solution not only due to their ability to inhibit the growth of pathogenic
39 microorganisms, but also because they are effective in restoring the composition of
40 the gastrointestinal tract microbiota, as well as have a positive effect on the human
41 immune system, mucus formation, and motility of the gastrointestinal tract [6].

42 However, the use of probiotics monotherapy, despite their high safety, also
43 has its disadvantages: a relatively low eradication rate and a long course of treatment
44 (1 month or more). The use of probiotic strains may not have a sufficiently
45 significant antagonistic effect on *H. pylori* and a pronounced positive effect on the
46 gastrointestinal microbiota, because they transit through the small intestine and
47 colon. Moreover, it remains unclear how to choose a suitable probiotic for each
48 individual.

49 Autoprobiotics, strains of normal microbiota isolated from a particular
50 individual and designed to correct human microecology, are an innovative way to
51 increase the effectiveness of eradication without producing negative effects on the
52 microbiota. Autoprobiotics stay in the colon longer, which allows to reduce the time
53 of treatment. Autoprobiotics prepared from native (indigenous) lactobacilli,
54 bifidobacteria, or enterococci may become the drugs of choice, since immunological
55 tolerance to them is formed from the first years of life, and they do not come into
56 conflict with other the resident microbiota of the human body [21]. There already
57 are studies showing the effectiveness of autoprobiotics based on indigenous strains
58 of *Lactobacillus spp.* in the restoration and stabilization of the content of the main

59 representatives of the normal gut microbiota (*Bifidobacterium spp.*, *Lactobacillus*
60 *spp.* and autoprobiotics based on *E. coli*) in treating dysbiotic disorders caused by
61 the use of antibacterial drugs [1, 8], as well as indigenous strains of *Enterococcus*
62 *spp.* in the treatment of intestinal pathology and neurological diseases [12].

63 **The aim** of our study was to evaluate the clinical performance of a
64 monotherapy by probiotics and autoprobiotic *Enterococcus faecium* for *H. pylori*
65 associated dyspepsia. We also evaluated gastric microbiota characteristics in the
66 absence and in the presence of this microorganism.

67 **Materials and Methods**

68 We examined 95 patients suffering from dyspepsia. Prior to commencing the
69 study, all patients signed an informed consent to a comprehensive medical
70 examination. The following groups were not included in the study: people who had
71 received a course of eradication therapy within the previous two years, people who
72 had taken antibiotics, proton pump inhibitors (PPIs), antacids, or bismuth containing
73 drugs within the previous two weeks, as well as people with severe physical illnesses
74 (including oncologic ailments) and/or infectious pathologies, pregnant and
75 breastfeeding women.

76 The comprehensive examination prior to treatment included: survey to
77 evaluate complaints (epigastric pain and signs of dyspepsia), gastroendoscopy,
78 which included biopsies from gastric antrum and body to confirm *H. pylori*
79 infection, and gastric microbiota analysis. The closing examination following the
80 full treatment included an survey to evaluate complaints and collection of fecal
81 samples to perform immunochromatographic stool tests for the detection of *H.*
82 *pylori*.

83 **Confirmation of *Helicobacter pylori* infection**

84 Biochemical, bacteriological, immunological and genetic methods were used
85 to confirm the presence of a pathogenic microorganism in the gastric mucosa. The
86 result was considered positive when the infection was detected by all methods or by

87 any one of the methods. The effectiveness of eradication was evaluated by
88 determining the *H. pylori* antigen in feces.

89 ***Rapid urease test***

90 We used the AMA RUT Expert test system to evaluate the urease activity of
91 bacteria in the biopsy specimen and the AMA RUT Reader (AMA, Russia) for
92 detection and record keeping. The AMA RUT Expert indicator is a test-slide with a
93 well containing a reactive element sealed with a film. The slide has special marking
94 on it, ensuring that the test results can be processed automatically.

95 ***Bacteriological method***

96 Pure culture of the pathogen was isolated from biopsy specimens of gastric
97 mucosa for each participant individually. Incubation protocol for *H. pylori* isolation
98 microaerophilic conditions at 37°C for 5 days on the surface of a special culture
99 medium (Columbia agar with 10% horse serum and 1% IsoVitalex, bio Merieux,
100 France). The number of viable bacteria (CFU/g) was determined by plating
101 corresponding 10-fold serial dilutions of biopsy specimens. Antimicrobial
102 susceptibility testing performed with the disc-diffusion method, sensitivity to
103 probiotics was determined by the drop plate method and the two-layer agar method.
104 The bacteriological method is the gold standard in the diagnosis of helicobacteriosis,
105 as it does not give false positive results, is specific and informative. Application of
106 bacteriological method allowed our team to confirm that *H. pylori* was present in the
107 sample, as well as to determine its sensitivity to antibiotics, probiotics, and
108 autoprobiotics.

109 In addition to detecting *H. pylori* infection, we also performed a comparative
110 analysis of gastric microbiota in the presence and in the absence of *H. pylori*. The
111 viable bacteria count (CFU/g) in gastric biopsy specimens was determined by plating
112 corresponding tenfold serial dilutions of suspensions on a number of selective dense
113 culture media in Petri dishes and counting the bacterial colonies after incubation (24
114 hours) at 37 °C. To determine the count of several genera of microorganisms such
115 as *E. coli*, *Enterococcus spp.*, *Klebsiella spp.*, *Proteus spp.*, *Enterobacter spp.* at the

116 same time, we used the following chromogenic selective media: Pronadisa 1424
117 (Spain), HiCrome Coliform Agar (India). The lactobacilli count was determined by
118 plating the culture on the Pronadisa 1043 Agar MRS medium (Spain) and culturing
119 in anaerobic jars with gas generating sachets (Thermo Scientific AN0025A (USA))
120 at 37°C for 48 hours.

121 ***Polymerase chain reaction***

122 PCR was used to detect the *cagA* and the *vacA* genes and thereby detect *H.*
123 *pylori* in the biopsy specimens. This method was chosen because it is highly precise
124 and informative. Moreover, features of the gastric microbiota were determined by
125 molecular genetic study (real-time PCR) using the Colonoflor test system and 16 S
126 rRNA metagenomic analysis.

127 ***Quantitative polymerase chain reaction***

128 Quantitative polymerase chain reaction (qPCR) was performed using the kit
129 Colonoflor 16 («AlphaLab», Russia) corresponding to the set of marker colonic
130 bacteria on the qPCR unit Mini-Opticon, BioRad. qPCR data on certain bacterial
131 species were confirmed by classical bacteriology study.

132 ***Immune chromatographic test***

133 The effect of probiotics and autoprobiotics used alone against *H. pylori* was
134 evaluated by a non-invasive stool antigen test 1.5–2 months after treatment
135 completion. Antigen determination in feces was carried out using the H&R *H. pylori*
136 Vegal Farmaceutica S.L. test system, Spain.

137 ***Probiotic medication used for intervention***

138 We used the probiotic autoprobiotic strains: *Enterococcus faecium* SF68 and
139 *Bifidobacterium bifidum* (Bifiform, Ferrosan, Denmark) and (*Enterococcus faecium*
140 L3 (Laminolact, «Avena», Russia) to study antagonistic activity *in vitro*.
141 Antagonistic activity was determined using the drop plate method. The investigated
142 probiotics were diluted in distilled water at a ratio of 1:100 and then added to a dish
143 with agar on which the *H. pylori* strain was plated. Growth was assessed on day 6-
144 7.

145 We used a starter culture based on the *Enterococcus faecium* L3 strain to study
146 the antagonistic activity *in vivo*. This strain was isolated from fermented milk,
147 deposited in *GenBank* (No SUB167269, 2 629 318 base pairs, contains 2717 genes)
148 and in the collection of the All-Russia Research Institute for Agricultural
149 Microbiology, ND-79, patent in Russia No 2220199. Genes encoding the synthesis
150 of several bacteriocins (including enterocins A, B, Enx α , and Enx β) were found in
151 the genome of this strain. The probiotics were administered for 20 days. The
152 probiotic was administered per os twice a day at doses of 50 ml (8.0 lgCFU/ml).

153 ***Autoprotobiotics making***

154 Autoprotobiotics were obtained as described in Russian patent No. 2546253 [5]:
155 at least 1 ml fecal samples were collected from patients who had not taken antibiotics
156 and/or probiotics for at least 10 days prior to collection; clones of indigenous strains
157 of *Enterococcus faecium* were isolated from the samples using a culture medium
158 containing sodium azide and crystal violet dye; then, colonies were selected based
159 on the coloring; pure cultures were obtained by plating three pink-colored colonies
160 with a burgundy center onto three sectors of Petri dishes with the same medium and
161 incubated in a thermostat under aerobic conditions at $t=37^{\circ}\text{C}$, and tested by PCR for
162 absence of genes of pathogenicity; then, non-pathogenic clones were selected and
163 cultured in a soy hydrolysate at no less than 10 ml per liter.

164 The 5% culture medium was prepared by diluting a lactose-free dry protein-
165 vitamin mixture "Super LF" (SLF) in a small amount of distilled water heated to 40
166 $^{\circ}\text{C}$ in a ratio of 1:1 until a homogeneous suspension was obtained. The resulting
167 suspension was filtered through 4 layers of medical gauze and diluted with the
168 remaining amount of distilled water (DW). The ratio of components by weight in
169 the final suspension should be: DW:SLF=95.5. The resulting suspension was
170 dispensed into 1-2 L plastic bottles and autoclaved at 120 $^{\circ}\text{C}$ and 1.2 atm for 15-30
171 minutes, then cooled to a temperature of 40 $^{\circ}\text{C}$.

172 Seed doses were prepared by aseptically taking 50 mg of lyophilized starter
173 culture and inoculating it with 2 ml of 5% culture medium cooled to 40 $^{\circ}\text{C}$. They

174 were then cultivated in an aerobical condition for 14-16 hours at 37°C. The grown
175 cell culture was transferred into 300 ml of sterile 5% culture medium cooled to 40°C
176 and incubated in a dry-air thermostat for 14-16 hours at 37°C. The resulting biomass
177 was used as a seeding dose for 1-2 L of culture medium. The starter culture which
178 changed the structure of hydrolysate earlier than others was selected and used to
179 prepare two liters of individual autoprobiotic product containing at least 10⁸ CFU
180 per 1 ml, which was administered to the patient orally at a dose of 50 ml 2 times a
181 day for at least 20 days.

182 *Methods of statistical analysis of study results*

183 Statistical processing of the results was carried out using Statistica 10 for
184 Windows (StatSoft, USA). Nonparametric pairwise multiple-comparison was used
185 to evaluate the effectiveness of diagnostic methods and treatments. A p-value <0.05
186 was considered statistically significant.

187 **Results and discussion**

188 Using various diagnostic methods, *H. pylori* infection was detected in 47 out
189 of the 95 patients, or in 49.4% of the patients. The bacteriological method produced
190 21 positive results.

191 *Evaluation of sensitivity of clinical isolates of Helicobacter pylori to* 192 *antibiotics*

193 We analyzed the sensitivity of the 21 isolated strains of *H. pylori* to the four
194 antibacterial drugs most commonly used in the eradication therapy of *H. pylori*-
195 associated diseases (Fig. 1).

196 The chart shows that the sensitivity to amoxicillin is the highest and reaches
197 100%, while sensitivity to metronidazole is half as high, with sensitivity to
198 levofloxacin and clarithromycin falling between these two values. The data obtained
199 are similar to the results of previous studies also conducted in St. Petersburg [4],
200 which indicates a stable level of resistance of the pathogen to the antibacterial agents
201 traditionally used in this region.

202 ***Evaluation of isolates of Helicobacter pylori isolates sensitivity to probiotics***
203 ***and autoprobiotics***

204 The bacteriological (cultural) method also allowed to determine the sensitivity
205 to probiotic and indigenous (autoprobiotic) strains of enterococci isolated from the
206 fecal samples of patients prior to eradication therapy. According to the chart (Fig.
207 2), the highest number of clinical isolates were sensitive to indigenous enterococci
208 (the autoprobiotic).

209 *H. pylori* sensitivity to antibiotics and probiotics allows for personalized
210 treatment of *H. pylori*-associated dyspepsia. Such an individualized approach makes
211 it possible to select the most effective means for both adjuvant therapy and
212 monotherapy (if necessary).

213 ***Gastric microbiocenosis assessment in the presence or absence of***
214 ***Helicobacter pylori***

215 We performed a comparative analysis of the gastric microbiota from 22
216 patients, 10 with positive *H. pylori*-status and 12 with negative *H. pylori*-status. The
217 gastric microbiota of the patients from these two groups differed significantly (Fig.
218 3).

219 The chart demonstrates that bacteria from the genera *Proteus*, *Klebsiella* and
220 *Enterobacter* were found only in samples collected from patients infected with *H.*
221 *pylori*. We found no statistically significant correlation between the presence of *H.*
222 *pylori* and *Fusobacterium spp.*, *Faecalibacterium prausnitzii* and *Bacteroides*
223 *fragilis*, *B. thetaiotaomicron*, *Bifidobacterium spp.*

224 It should be noted that when lactobacilli and enterococci were detected in the
225 gastric samples at a concentration greater than 3 lgCFU/mL, the probability of
226 detecting *H. pylori* was lower (Fig. 4).

227 Consequently, as demonstrated on Figure. 3 and Figure 4, we observe an
228 increased presence of opportunistic pathogen belonging to the Enterobacteriaceae
229 family combined with concurrent regress in numbers of colonies of enterococci and

230 lactobacilli (non-pathogenic microorganism) in *H. pylori*-positive patients. We
231 suggest that observed imbalance in gastric microbiota can be attributed as an
232 underlying cause for development of symptoms of dyspepsia and following *H.*
233 *pylori*-associated diseases.

234 ***Gut microbiome study by qPCR***

235 The study was performed by comparing the following microorganisms (the
236 quantitative content of representatives of the intestinal microbiota): the total number
237 of bacteria, *Acinetobacter spp.*, *Citrobacter spp.*, *Escherichia coli* and
238 *enteropathogenic E. coli*, *Proteus spp.*, *Lactobacillus spp.*, *Bifidobacterium spp.*,
239 *Bacteroides thetaiotaomicron*, *Bacteroides fragilis* group, *Clostridium difficile*,
240 *Clostridium perfringens*, *Enterococcus spp.*, *Faecalibacterium prausnitzii*,
241 *Fusobacterium nucleatum* and *Parvimonas micra*.

242 Changes in the microbiota before and after therapy had no significant
243 differences in patients receiving probiotics and autoprobiotics. When considering
244 the composition of the microbiota before and after therapy of all patients, it was
245 shown that the quantitative content of Ruminococcus Metanobrevibacterium.
246 Roseburia, Eubacterium Blautia Enterococcus increased. The populations of
247 *Prevotella*, *Streptococcus*, *Salmonella*, *Parvimonas Fusobacterium*, *Citrobacter*,
248 *Klebsiella*, *Enterobacter* *Bacteroides thetaiotaomicron* on the contrary decreased (fig.
249 5).

250 ***Assessment of the clinical impact in treatment of H. pylori infection***

251 Within the main group of patients, we distinguished a separate cohort of 11
252 patients who previously had recorded allergic reactions to antibiotics that are used
253 in standard eradication treatment regimens. This cohort was divided into two
254 subgroups: one received probiotic alone (5 patients) and the other received solely
255 autoprobiotic therapy (6 patients). The summarized results for clinical efficacy in
256 relieving the symptoms of dyspepsia and the anti-*Helicobacter* activity of these
257 drugs is demonstrated in Tables 1 and 2.

258 According to questionnaire assessment symptoms of dyspepsia were

259 completely eliminated after treatment with autoprobiotic and probiotic. The use of
260 autoprobiotics based on indigenous enterococci alone is more effective in
261 eradicating *H. pylori* than the use of a starter culture based on the *E. faecium* L3
262 strain.

263 **Discussion**

264 The prerequisite for this study were problems with the use of antibiotics, such
265 as insufficient efficacy and side effects (diarrhea, nausea, bloating, allergic reactions
266 etc.). In this study, for the first time, the possibility of using autoprobiotics in
267 monotherapy of *H. pylori*-associated dyspepsia is considered.

268 The choice of the type of autoprobiotic was associated with the high efficiency
269 of autoprobiotic enterococci in the correction of gut dysbiosis, therapy of irritable
270 bowel syndrome and metabolic syndrome. In addition, this study has already
271 revealed an inverse correlation between the presence of enterococci in stomach
272 biopsies and enterococcus and lactobacilli.

273 It is not surprising that when correcting the microbiota of the gastrointestinal
274 tract with the help of indigenous enterococci isolated from the patient's feces, the
275 elimination of the pathogen and the disappearance of dyspeptic symptoms were
276 observed. Previously, such effects were described with the introduction of several
277 probiotics, among which some of the most effective were based on *Enterococcus*
278 *faecium* [21].

279 In vitro studies have demonstrated a high sensitivity of *H. pylori* to probiotics
280 based on enterococci, including autoprobiotic, comparable to sensitivity to
281 antibiotics. As it was shown earlier, the effect of probiotics is associated with the
282 production of enterocins [6].

283 The intake of the functional food product containing *E. faecium* L3 and the
284 autoprobiotic starter culture containing *E. faecium* have many positive effects: the
285 disappearance of pain syndrome, heartburn, belching, flatulence, apparently due to
286 the normalization of the composition of the gut microbiota and *H. pylori* eradication.

287 The use of autoprobiotics did not reveal significant differences in the

288 composition of the gut microbiota after administration of probiotic *E. faecium* L3.
289 The advantage of autoprobiotic can be the duration of the effect of autoprobiotics,
290 established earlier [12].

291 **Conclusion**

292 For the investigated *H. pylori* strains the sensitivity is similar to both
293 antibiotics used in standard eradication protocols and probiotics. The sensitivity of
294 *H. pylori* to autoprobiotics based on indigenous enterococci is slightly higher than
295 to probiotics. Treatment regimen with an autoprobiotic based on indigenous
296 enterococci alone showed a higher eradication rate compared to a starter culture
297 based on the *E. faecium* L3 strain. It is reasonable to include both probiotics and
298 autoprobiotics in comprehensive eradication regimens due to dysbiotic changes of
299 gastric microbiota in patients with dyspepsia and persisting *H. pylori* infection.
300 When standard anti-helicobacter therapy cannot be used, autoprobiotics should be
301 used as the preferred treatment. Enterococci-based drugs are the most promising for
302 further research into the anti-Helicobacter effect of probiotics and autoprobiotics.

FIGURES

Figure 1. Prevalence of detected of *H. pylori* strains sensitive to antibacterial drugs

X axis: types of antibiotics

Y axis: **Prevalence** of detected *H. pylori* strains, %

Рисунок 1. Распространенность выявленных штаммов *H. pylori*, чувствительных к антибактериальным препаратам

Ось X: типы антибиотиков

Ось Y: Распространенность выявленных штаммов *H. pylori*, %

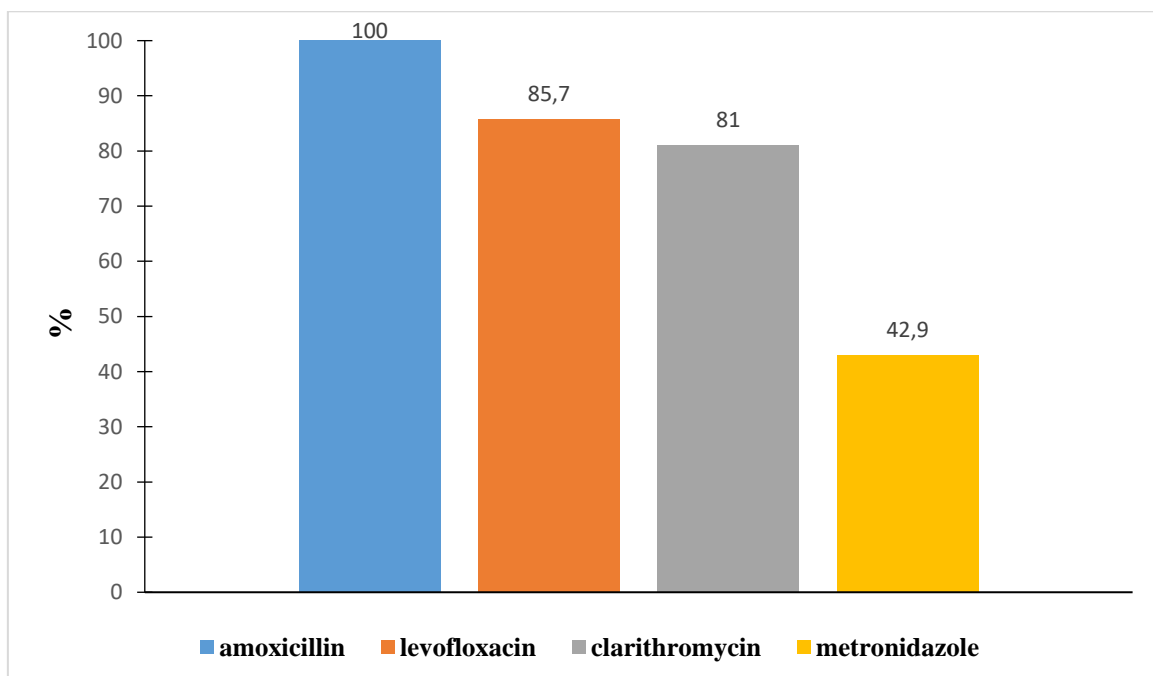


Figure 2. Prevalence of detected *H. pylori* strains sensitive to probiotics and autoprobiotics

X axis: Types of probiotics and autoprobiotics

Y axis: **Prevalence** of detected sensitive *H. pylori* strains, %

Рисунок 2. Распространенность выявленных штаммов *H. pylori*, чувствительных к пробиотикам и аутопробиотикам.

Ось X: Типы пробиотиков и аутопробиотиков.

Ось Y: Распространенность выявленных чувствительных штаммов *H. pylori*, %

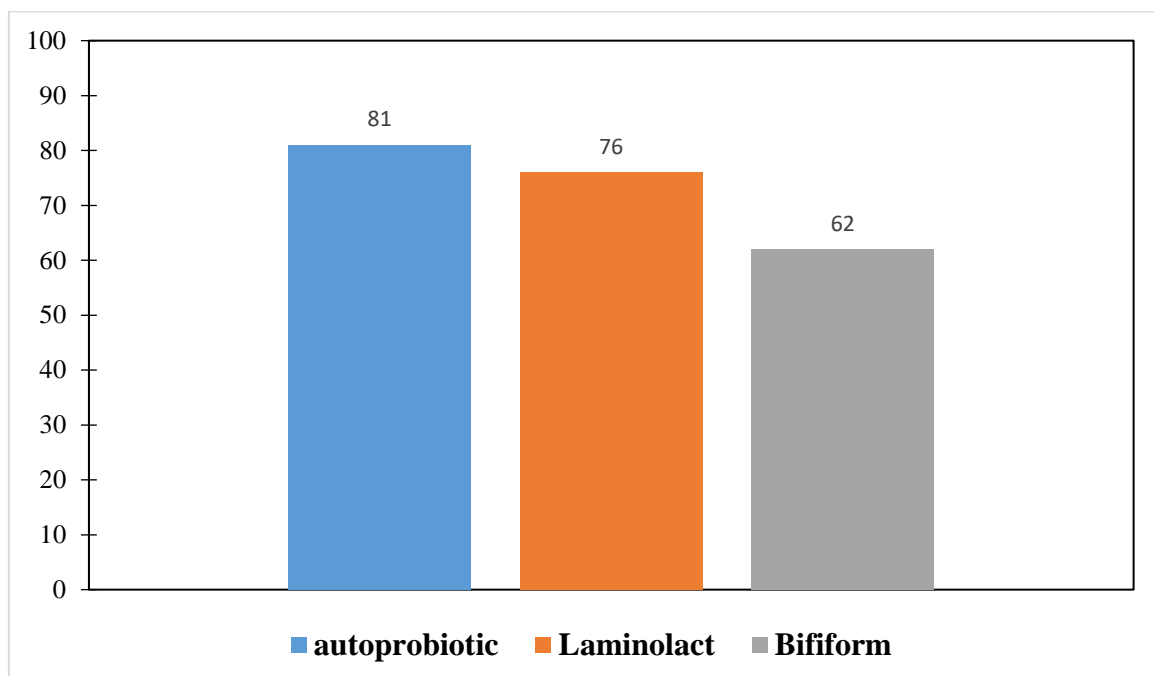


Figure 3. Quantitative level of various opportunistic bacteria in gastric biopsy specimens from patients with positive and negative *H. pylori*-status.X axis: *H. pylori*-status of patients

Y axis: level of gastric microorganisms, lgCFU/g

Рисунок 3. Количественный уровень различных условно-патогенных бактерий в образцах биопсии желудка от пациентов с положительным и отрицательным статусом *H. pylori*.Ось X: *H. pylori*-статус пациентов

Ось Y: уровень желудочных микроорганизмов, lgКОЕ/г

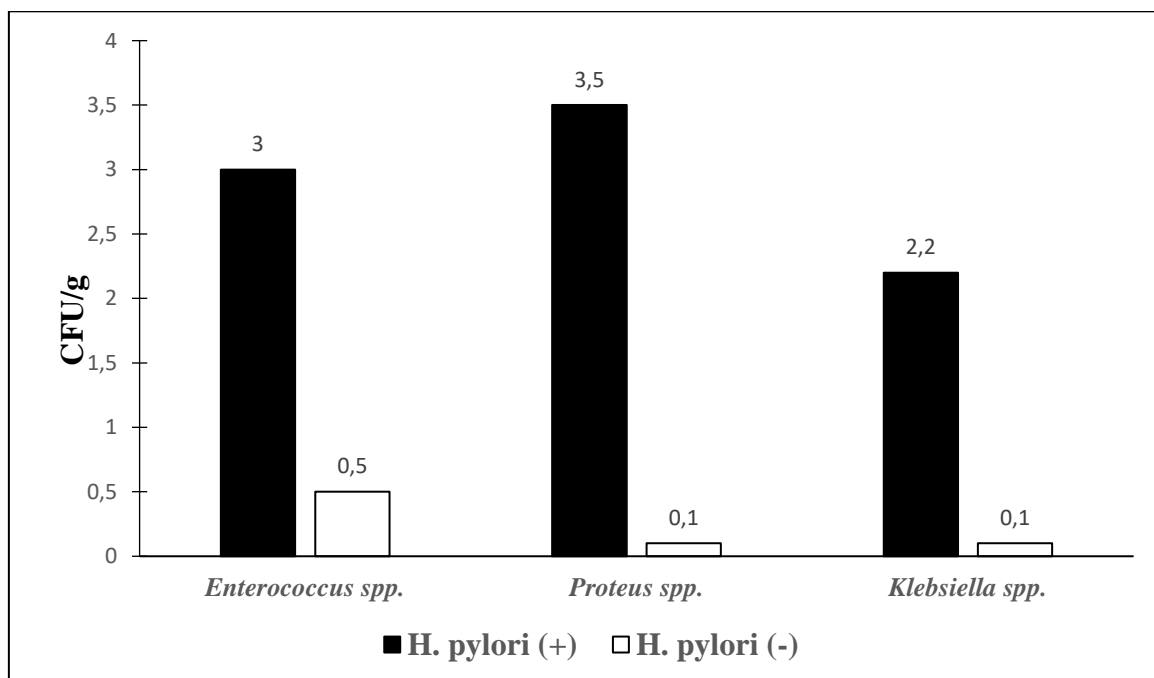


Figure 4. Prevalence of detected gastric *H. pylori* in patients with dyspepsia with isolated lactobacilli and enterococci.X axis: *H. pylori*-status of patientsY axis: **Prevalence** of isolated gastric lactobacilli and enterococci, %**Рисунок 4. Распространенность выявленной желудочной *H. pylori* у больных диспепсией с изолированными лактобациллами и энтерококками.**Ось X: *H. pylori*-статус пациентов

Ось Y: Распространенность изолированных желудочных лактобацилл и энтерококков, %

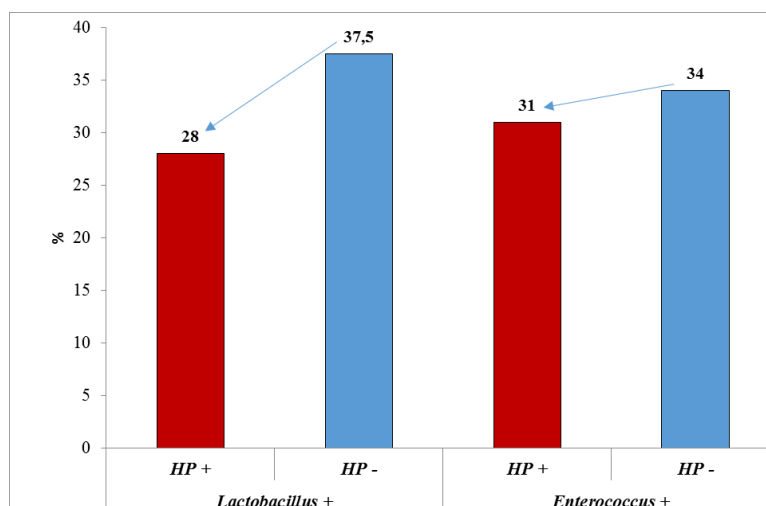
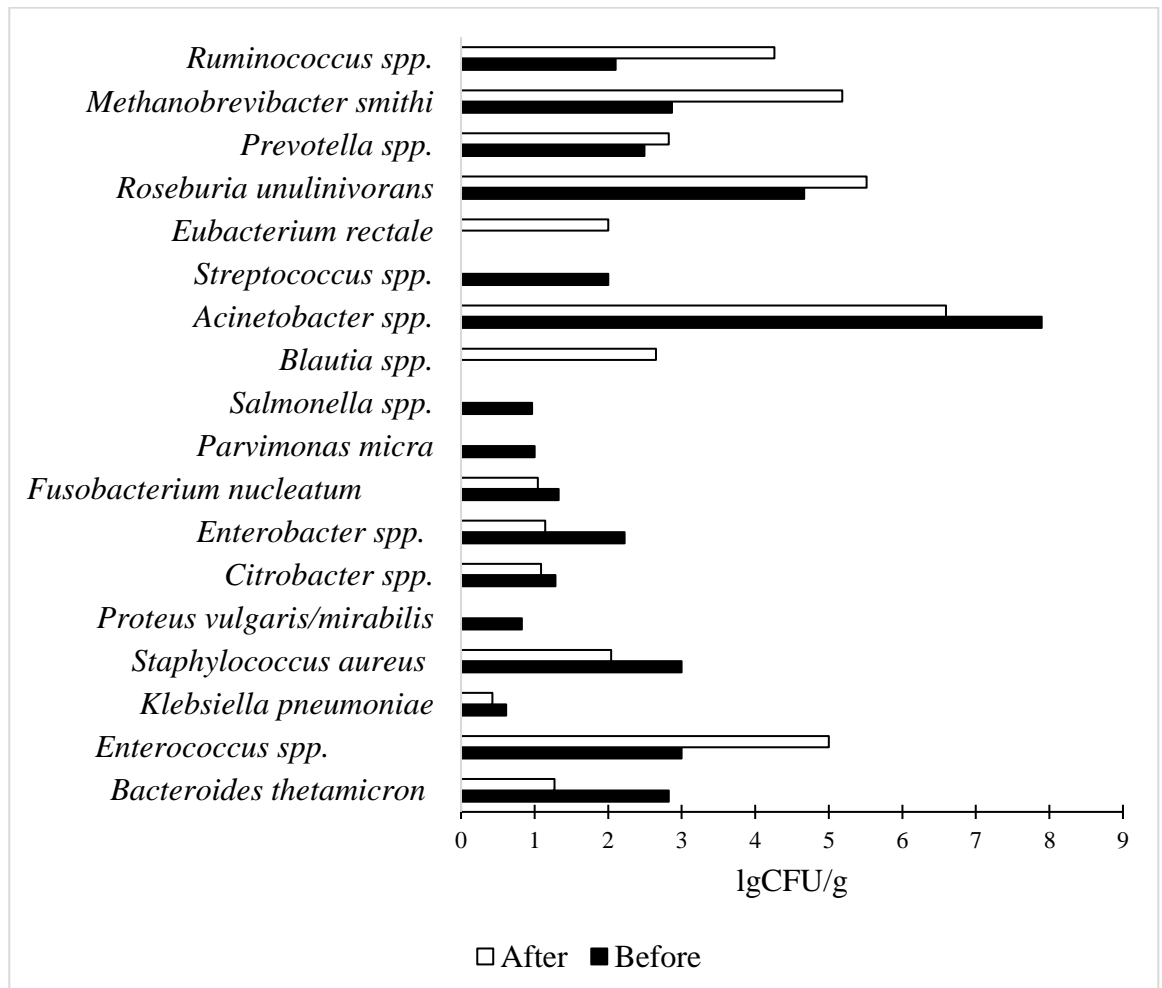


Figure 5. Gut microbiota profile before and after autoprobiotic- and probiotic-therapy of *H. pylori*+ gastritis

Рисунок 5. Профиль микробиоты кишечника до и после аутопробиотической и пробиотической терапии *H. pylori*+ гастрита.



TABLES

Table 1 Evaluation of the clinical effectiveness for autoprobiotics and probiotics in reversing symptoms in patients with *Helicobacter pylori*-associated dyspepsia**Таблица 1 Оценка клинической эффективности аутопробиотиков и пробиотиков в купировании симптомов у пациентов с *Helicobacter pylori*-ассоциированной диспепсией**

Symptom, frequency in % Симптом, частота в %	Autoprobiotic Аутопробиотик		Probiotic Пробиотик	
	Before treatment До лечения	After treatment После лечения	Before treatment До лечения	After treatment После лечения
Eructation Отрыжка	33	0	60	0
Heartburn Изжога	50	0	60	0
Epigastric pain Боли в эпигастральной области	100	0	100	0
Bloating Вздутие живота	67	0	80	0
Nausea Тошнота	17	0	40	0

Table 2 Evaluation of the clinical effectiveness of autoprobiotics and probiotics in *Helicobacter pylori* eradication

Таблица 2 Оценка клинической эффективности аутопробиотиков и пробиотиков при эрадикации *Helicobacter pylori*

Parameters Параметры	Autoprobiotic (n=10) Аутопробиотик (n=10)	Probiotic (n=12) Пробиотик (n=12)
Effectiveness of anti- <i>Helicobacter</i> action: number of <i>H. pylori</i> -negative samples based on stool antigen test (immunochromatographic method), % (n) Эффективность антихеликобактерного действия: количество <i>H. pylori</i> -негативных образцов по результатам анализа кала на антиген (иммунохроматографический метод), % (n)	83(5)	60(2)

TITLE PAGE_METADATA

Блок 1. Информация об авторе ответственном за переписку

Сварваль Алена Владимировна - кандидат медицинских наук, старший научный сотрудник лаборатории идентификации патогенов, Федеральное бюджетное учреждение науки «Санкт-Петербургский научно-исследовательский институт эпидемиологии и микробиологии имени Пастера» 197101, 14, Mira street, St. Petersburg, Russia, 197101, Санкт-Петербург, ул. Мира, д. 14, Телефон: +7 812 232 84 76, E-mail: alenasvar@rambler.ru

Svarval Alena Vladimirovna - PhD, Senior Researcher, Saint-Petersburg Pasteur Institute, 197101, 14, Mira street, St. Petersburg, Russia, 197101, Санкт-Петербург, ул. Мира, д. 14, Tel. +7 812 232 84 76, E-mail: alenasvar@rambler.ru

Блок 2. Информация об авторах

Elena I. Ermolenko - Doctor of Medical Sciences, Head, Laboratory of Biomedical Microecology; Professor, Faculty of Medical Microbiology, Department of Medical Microbiology; of Federal State Budgetary Educational Institution 'Institute of Experimental Medicine'

Ермоленко Елена Игоревна – доктор медицинских наук, заведующий лабораторией биомедицинской микроэкологии, отдел молекулярной микробиологии Федерального государственного бюджетного научного учреждения «Институт экспериментальной медицины»

Anastasiia S. Molostova – Gastroenterologist at Federal State Budgetary Educational Institution 'Institute of Experimental Medicine', Junior Researcher at North-Western district scientific and clinical center named after L.G. Sokolov Federal Medical and Biological Agency

Молостова Анастасия Сергеевна - врач-гастроэнтеролог, Федерального государственного бюджетного научного учреждения «Институт экспериментальной медицины»

Natalia V. Baryshnikova – PhD, MD, Junior Researcher at Laboratory of Medical and Social Pediatric Problems at Federal State budgetary Educational Institution of Higher Education “St. Petersburg State Pediatric Medical University” of the Ministry of Health of the Russian Federation; Associate Professor of Internal Diseases Department of Dental Faculty of ³Pavlov First Saint Petersburg State Medical University of the Ministry of Health of the Russian Federation; Researcher at Molecular Microbiology Laboratory of Federal State Budgetary Educational Institution ‘Institute of Experimental Medicine’

Барышникова Наталья Владимировна – кандидат медицинских наук, доцент, младший научный сотрудник лаборатории медико-социальных проблем педиатрии Федерального государственного бюджетного образовательного учреждения высшего образования «Санкт-Петербургский государственный педиатрический медицинский университет» Министерства здравоохранения Российской Федерации; доцент кафедры внутренних болезней стоматологического факультета Федерального государственного бюджетного образовательного учреждения высшего образования «Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова» Министерства здравоохранения Российской Федерации; научный сотрудник лаборатории молекулярной микробиологии Федерального государственного бюджетного научного учреждения «Институт экспериментальной медицины»

Nikita S. Gladyshev - Researcher, Pathogen Identification Laboratory at Saint-Petersburg Pasteur Institute; student of the educational program "Medical Business", Faculty of Medicine Saint Petersburg State University

Гладышев Никита Сергеевич – студент образовательной программы «Лечебное дело», медицинского факультета ФГБОУ ВПО Санкт-Петербургский государственный университет; лаборант лаборатории идентификации патогенов, Федеральное бюджетное учреждение науки «Санкт-Петербургский научно-исследовательский институт эпидемиологии и микробиологии имени Пастера»

Victor A. Kashchenko - Doctor of Medical Sciences, Professor, MD, Head of the Department of faculty surgery Saint Petersburg State University; Deputy Director for scientific and education work, Chief Surgeon at North-Western district scientific and clinical center named after L.G. Sokolov Federal Medical and Biological Agency

Кащенко Виктор Анатольевич - доктор медицинских наук, профессор, заведующий кафедрой факультетской хирургии ФГБОУ ВПО «Санкт-Петербургский государственный университет», заместитель генерального директора по научно-образовательной работе, главный хирург Федерального государственного бюджетного учреждения «Северо-Западный окружной научно-клинический центр имени Л.Г. Соколова Федерального медико-биологического агентства».

Alexander N. Suvorov - Doctor of Medical Sciences, Professor, MD, Corresponding Member of the Russian Academy of Sciences, Head of the Department of Molecular Microbiology of Federal State Budgetary Educational Institution 'Institute of Experimental Medicine', ORCID iD 0000-0003-2312-5589, Email: alexander_suvorov1@hotmail.com

Суворов Александр Николаевич - доктор медицинских наук, профессор, чл.-корр. РАН, руководитель отдела молекулярной микробиологии Федерального государственного бюджетного научного учреждения «Институт

экспериментальной медицины», ORCID iD 0000-0003-2312-5589, Email:
alexander_suvorov1@hotmail.com

Блок 3. Метаданные статьи

**The clinical effectiveness of probiotics and autoprobiotics in the treatment
Helicobacter pylori-associated dyspepsia**

**Эффективность пробиотиков и аутопробиотиков в монотерапии
диспепсии, ассоциированной с инфекцией *Helicobacter pylori***

Сокращенное название статьи:

The treatment of *H. pylori*-associated dyspepsia

Лечение диспепсии, ассоциированной с инфекцией *H. pylori*

Key words: *Helicobacter pylori*, eradication, probiotics, autoprobiotics,
enterococci, *Enterococcus faecium*

Ключевые слова: *Helicobacter pylori*, эрадикация, пробиотики,
аутопробиотики, энтерококки, *Enterococcus faecium*

11 страниц, 1 таблица, 5 рисунков

Оригинальная статья

13.04.2022 г.

REFERENCES

Reference sequence number	Authors, title of a publication and source where it was published, publisher's imprint	Full name, title of a publication and source in English	Reference's URL
1	Боровкова Е.А., Алиева Е.В. Микробиологическое исследование микрофлоры толстого кишечника на дисбактериоз в оценке эффективности аутопробиотикотерапии. Естественные и технические науки.2020; 8 (146):24-33.	Borovkova E.A., Alieva E.V. Mikrobiologicheskoe issledovanie mikroflory tolstogo kishechnika na disbakterioz v otsenke effektivnosti autoprotiotikoterapii. Estestvennye i tekhnicheskie nauki.2020; 8 (146):24-33.	https://www.elibrary.ru/item.asp?id=44148083 [eLIBRARY ID: 44148083]
2	Ивашкин В.Т., Маев И.В., Лапина Т.Л. и соавт. Клинические рекомендации Российской гастроэнтерологической ассоциации по диагностике и лечению инфекции <i>Helicobacter pylori</i> у взрослых. Российский журнал гастроэнтерологии, гепатологии, колопроктологии. 2018; 28(1):55-70.	Ivashkin V.T., Mayev I.V., Lapina T.L et al. Diagnostics and treatment of <i>Helicobacter pylori</i> infection in adults: Clinical guidelines of the Russian gastroenterological association. Ross z gastroenterolgepatolokoloproktol. 2018; 28(1):55-70.	https://www.gastro-j.ru/jour/article/view/218 [doi.org/10.22416/1382-4376-2018-28-1-55-70].
3	Лазебник Л. Б., Ткаченко Е. И., Абдулганиева Д. И. и соавт. VI	Lazebnik L. B., Tkachenko E. I., Abdulganieva D. I. et al. VI national	https://www.elibrary.ru/item.asp?id=28870080

	национальные рекомендации по диагностике и лечению кислотозависимых и ассоциированных с <i>Helicobacter pylori</i> заболеваний (VI Московские соглашения). Экспериментальная и клиническая гастроэнтерология 2017; 138 (2): 3–21.	guidelines for the diagnosis and treatment of acid-related and <i>Helicobacter pylori</i> -associated diseases (VI Moscow agreement). Eksperimental'naya i Klinicheskaya Gastroenterologiya. 2017; 138 (2): 3–21.	[eLIBRARY ID: 28870080]
4	Симаненков В.И., Захарова Н.В., Жебрун А.Б., Сварваль А.В., Савилова И.В., Ферман Р.С. Резистентность <i>Helicobacter pylori</i> к антимикробным препаратам по результатам бактериологического тестирования. Лечащий врач. 2015.;4:91.	Simanenkova V.I., Zakharova N.V., Zhebrun A.B., Svarval' A.V., Savilova I.V., Ferman R.S. Rezistentnost' <i>Helicobacter pylori</i> k antimikrobnym preparatam po rezul'tatam bakteriologicheskogo testirovaniya. Lechashchii vrach. 2015.;4:91	https://www.elibrary.ru/item.asp?id=23280121 [eLIBRARYID: 23280121]
5	Симаненков В.И., Суворов А.Н., Соловьева О.И. Способ получения персонифицированного аутопробиотического продукта и способ лечения синдрома раздраженного кишечника с использованием этого продукта. Патент РФ на изобретение № 2546253 / 02.03.2015. Бюл. № 10.	Simanenkova V.I., Suvorov A.N., Solov'eva O.I. Sposob polucheniya personifitsirovannogo autoprotioticheskogo produkta i sposob lecheniya sindroma razdrzhennogo kishchnika s ispol'zovaniem etogo produkta. Patent RF na izobretenie № 2546253 / 02.03.2015. Byul. № 10.	http://www.findpatent.ru/patent/254/2546253.html

6	Суворов А.Н., Барышникова Н. В., Сварваль А. В., Ниязов Р. М. Возможности некоторых пробиотических штаммов в эрадикации <i>Helicobacter pylori</i> in vitro и in vivo. Фарматека. 2018;(2):74–8	Suvorov A.N., Baryshnikova N. V., Svarval A. V., Niyazov R. M. Possibilities of some probiotic strains in the eradication of <i>Helicobacter pylori</i> in vitro and in vivo. Pharmateca. 2018; (2): 74–8.	https://www.elibrary.ru/item.asp?id=32530273 [eLIBRARY ID: 32530273]
7	Ткаченко Е.И., Успенский Ю.П., Барышникова Н.В. Оптимизация лечения заболеваний, ассоциированных с <i>Helicobacter pylori</i> . Врач. 2012; 1:36-38.	Tkachenko E., Uspenskiy Yu., Baryshnikova N. Optimization of treatment for <i>Helicobacter pylori</i> -associated diseases. Vrach. 2012; 1:36-38.	https://www.elibrary.ru/item.asp?id=17632284 [eLIBRARY ID: 17632284]
8	Цапиева А.Н., Боровкова Е.А., Карасёва А.Б., Алиева Е.В., Суворов А.Н. Разработка метода идентификации индигенных лактобацилл кишечника при создании аутопробиотиков. // Вопросы детской диетологии. 2019; 17 (3): 52-59.	Tsapieva A.N., Borovkova E.A., Karaseva A.B., Alieva E.V., Suvorov A.N. Razrabotka metoda identifikatsii indigennykh laktobatsill kishechnika pri sozdanii autoprobiotikov. // Voprosydetskoidietologii. 2019; 17 (3): 52-59.	https://www.elibrary.ru/item.asp?id=41375709 [doi:10.20953/1727-5784-2019-3-52-59]
9	Boonyaritchaikij S, Kuwabara K, Nagano J, Kobayashi K, Koga Y. Long-term administration of probiotics to asymptomatic pre-school children for either the eradication or the prevention	-	https://pubmed.ncbi.nlm.nih.gov/19702850/ [doi:10.1111/j.1523-5378.2009.00675.x.]

	of <i>Helicobacter pylori</i> infection. <i>Helicobacter</i> . 2009 Jun;14(3):202-7.		
10	Canducci F, Cremonini F, Armuzzi A, Di Caro S, Gabrielli M, Santarelli L, Nista E, Lupascu A, De Martini D, Gasbarrini A. Probiotics and <i>Helicobacter pylori</i> eradication. <i>Dig Liver Dis</i> . 2002 Sep; 34 Suppl2:S81-3.	-	https://pubmed.ncbi.nlm.nih.gov/12408448/ [doi:10.1016/s1590-8658(02)80172-4.]
11	Dore MP, Cuccu M, Pes GM, Manca A, Graham DY. <i>Lactobacillus reuteri</i> in the treatment of <i>Helicobacter pylori</i> infection. <i>Intern Emerg Med</i> . 2014 Sep;9(6):649-54.	-	https://pubmed.ncbi.nlm.nih.gov/24178436/ [doi: 10.1007/s11739-013-1013-z]
12	Ermolenko E.I., Abdurasulova I.N., Kotyleva M.P., Svirido D.A., Matsulevich A.V., Karaseva A.B. Effects of Indigenous Enterococci on the Intestinal Microbiota and the Behavior of Rats // <i>Neuroscience and Behavioral Physiology</i> , 2018, 48(4):496–505.	-	https://www.elibrary.ru/item.asp?id=35499976& [doi: 10.1007/s11055-018-0591-7]
13	Gotteland M, Poliak L, Cruchet S, Brunser O. Effect of regular ingestion of <i>Saccharomyces boulardii</i> plus inulin or <i>Lactobacillus acidophilus</i> LB in children colonized by <i>Helicobacter pylori</i> .	-	https://pubmed.ncbi.nlm.nih.gov/16421034/ [doi: 10.1111/j.1651-2227.2005.tb01848.x.]

	ActaPaediatr. 2005 Dec;94(12):1747-51.		
14	Losurdo G, Cubisino R, Barone M, Principi M, Leandro G, Ierardi E, Di Leo A. Probiotic monotherapy and Helicobacter pylori eradication: A systematic review with pooled-data analysis. World J Gastroenterol. 2018 Jan 7;24(1):139-149.	-	https://pubmed.ncbi.nlm.nih.gov/29358890/ [doi: 10.3748/wjg.v24.i1.139.]
15	Lü M, Yu S, Deng J, Yan Q, Yang C, Xia G, Zhou X. Efficacy of Probiotic Supplementation Therapy for Helicobacter pylori Eradication: A Meta-Analysis of Randomized Controlled Trials. PLoS One. 2016 Oct 10;11(10):e0163743.	-	https://pubmed.ncbi.nlm.nih.gov/27723762/ [doi: 10.1371/journal.pone.0163743.]
16	Malfertheiner P, Megraud F, O'Morain CA et al. Management of Helicobacter pylori infection--the Maastricht IV/ Florence Consensus Report. Gut. 2012 May;61(5):646-64.	-	https://pubmed.ncbi.nlm.nih.gov/22491499/ [doi: 10.1136/gutjnl-2012-302084.]
17	Malfertheiner P, Megraud F, O'Morain CA et al. Management of Helicobacter pylori infection-the Maastricht	-	https://pubmed.ncbi.nlm.nih.gov/27707777/ [doi: 10.1136/gutjnl-2016-312288.]

	V/Florence Consensus Report. Gut. 2017 Jan;66(1):6-30.		
18	Molina-Infante J, Gisbert JP. Probiotics for <i>Helicobacter pylori</i> eradication therapy: not ready for prime time. Rev Esp Enferm Dig. 2013 Sep;105(8):441-4.	-	https://pubmed.ncbi.nlm.nih.gov/24274440/ [doi: 10.4321/s1130-01082013000800001.]
19	Rosania R, Minenna MF, Giorgio F, Facciorusso A, De Francesco V, Hassan C, Panella C, Ierardi E. Probiotic multistrain treatment may eradicate <i>Helicobacter pylori</i> from the stomach of dyspeptics: a placebo-controlled pilot study. Inflamm Allergy Drug Targets. 2012 Jun;11(3):244-9.	-	https://pubmed.ncbi.nlm.nih.gov/22452604/ [doi: 10.2174/187152812800392698.]
20	Shi X, Zhang J, Mo L, Shi J, Qin M, Huang X. Efficacy and safety of probiotics in eradicating <i>Helicobacter pylori</i> : A network meta-analysis. Medicine (Baltimore). 2019 Apr;98(15):e15180		https://pubmed.ncbi.nlm.nih.gov/30985706/ [doi: 10.1097/MD.00000000000015180]
21	Suvorov A, Karaseva A, Kotyleva M et al. Autoprobiotics as an approach for restoration of personalised microbiota. Front Microbiol. 2018; Sep 12; 9:1869.		https://pubmed.ncbi.nlm.nih.gov/30258408/ [doi: 10.3389/fmicb.2018.01869]

22	Szajewska H, Horvath A, Piwowarczyk A. Meta-analysis: the effects of <i>Saccharomyces boulardii</i> supplementation on <i>Helicobacter pylori</i> eradication rates and side effects during treatment. <i>Aliment Pharmacol Ther.</i> 2010 Nov;32(9):1069-79.	-	https://pubmed.ncbi.nlm.nih.gov/21039671/ [doi: 10.1111/j.1365-2036.2010.04457.x.]
23	Wang F, Feng J, Chen P, Liu X, Ma M, Zhou R, Chang Y, Liu J, Li J, Zhao Q. Probiotics in <i>Helicobacter pylori</i> eradication therapy: Systematic review and network meta-analysis. <i>Clin Res Hepatol Gastroenterol.</i> 2017 Sep;41(4):466-475.	-	https://pubmed.ncbi.nlm.nih.gov/28552432/ [doi: 10.1016/j.clinre.2017.04.004.]
24	Zhang MM, Qian W, Qin YY, He J, Zhou YH. Probiotics in <i>Helicobacter pylori</i> eradication therapy: a systematic review and meta-analysis. <i>World J Gastroenterol.</i> 2015 Apr 14;21(14):4345-57.	-	https://pubmed.ncbi.nlm.nih.gov/25892886/ [doi: 10.3748/wjg.v21.i14.4345.]