

**ASSESSED CORRELATION BETWEEN BIOLOGICAL DIVERSITY OF
OROPHARYNGEAL MICROBIOTA AND ATOPIC DERMATITIS
SEVERITY AND EXACERBATIONS**

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**ОЦЕНКА КОРРЕЛЯЦИИ БИОЛОГИЧЕСКОГО РАЗНООБРАЗИЯ МИКРОБИОТЫ
РОТОГЛОТКИ СО СТЕПЕНЬЮ ТЯЖЕСТИ И ЧАСТОТОЙ ОБОСТРЕНИЙ У
ПАЦИЕНТОВ С АТОПИЧЕСКИМ ДЕРМАТИТОМ**

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Abstract

Atopic dermatitis (AtD) is a multifactorial inflammatory skin disease characterized by itching, chronic recurrent course and age-related features of lesions. AtD pathogenesis has not been fully elucidated yet. An important factor for AtD emergence and progression is the imbalance in symbiotic microbiota. The research publications provide a few studies about a role for oropharyngeal microorganisms in AtD immunopathogenesis. The aim of the study is to analyze biological diversity of oropharyngeal microbial communities in varying AtD severity. 97 male patients, aged from 16 to 19 years, with different AtD severity were included in the study. Culture study of oropharyngeal discharge was also performed. Biological material was seeded on the expanded list of growth media and incubated for 5 days at the 37°C. To assess the biological diversity of the oropharyngeal microbiota, the coefficient of constancy (C) was used, in order to classify individual microorganisms as permanent, additional or transient. Statistical data processing was performed using the Stat Tech software (version 4.0.0, Stattech LLC, Russia). While examining biological diversity of the oropharyngeal microbiota in AtD patients, 58 microbial species were isolated and identified. After statistical analysis the significant differences in frequency of isolation, depending on different AtD severity were observed for microbes such as *Streptococcus vestibularis* and *Rothia dentocariosa*. When *R. dentocariosa* is isolated from the oropharynx, the chances of AtD exacerbation emergence decreased by 6 times, whereas in case of *S. vestibularis*, on the contrary, it increased by 5 times. Therefore, identification of transitions of individual microbes from transient to additional and permanent microbiota and vice versa, depending on the AtD stage and severity, allows to analyze an influence of specific microorganisms in AtD pathological processes and to establish definite new microbiological predictors of AtD exacerbation and remission.

МИКРОБИОТА РОТОГЛОТКИ У ПАЦИЕНТОВ С АТОПИЧЕСКИМ ДЕРМАТИТОМ
OROPHARYNGEAL MICROBIOTA IN PATIENTS WITH ATOPIC DERMATITIS
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Keywords: atopic dermatitis, oropharyngeal microbiota, biological diversity, skin diseases, immunological disorders, microbiome.

Резюме

Атопический дерматит (АтД) - многофакторное воспалительное заболевание кожи, характеризующееся зудом, хроническим рецидивирующим течением и различными возрастными особенностями. Патогенез АтД еще полностью не выяснен. Важным фактором возникновения и прогрессирования АтД является дисбаланс симбиотической микробиоты. Научная литература содержит небольшое количество информации об участии микроорганизмов ротоглотки в иммунопатогенезе АтД. Цель исследования - провести анализ биологического разнообразия микробных сообществ ротоглотки в группах пациентов с различной степенью тяжести АтД и на разных стадиях АтД (ремиссия/обострение). В исследование были включены 97 пациентов с различной степенью тяжести АтД. Также было проведено культуральное исследование выделений из ротоглотки. Посев материала проводили на расширенный перечень питательных сред и инкубировали при температуре 37°C в течение 5 суток. Для оценки биологического разнообразия микробиоты ротоглотки использовался коэффициент постоянства (С), позволяющий классифицировать отдельные микроорганизмы как постоянные, дополнительные или временные. Статистические расчёты проводились с использованием программного обеспечения StatTech (версия 4.0.0, разработчик ООО "Статтех", Россия). В ходе исследования у включенных пациентов было выделено и идентифицировано 58 видов микроорганизмов. В ходе статистического анализа были получены значимые различия в частоте выделения, в зависимости от различной степени тяжести АтД, для таких микроорганизмов, как *Streptococcus vestibularis* и *Rothia dentocariosa*. При выделении *R. dentocariosa* из ротоглотки вероятность возникновения обострения АтД снижалась в 6 раз. При выделении *S. vestibularis* вероятность возникновения обострения АтД, в отличие от *R. dentocariosa*, увеличивалась в 5 раз. Таким образом, выявление переходов отдельных микроорганизмов от транзиторной к дополнительной и постоянной микробиоте и наоборот, в

зависимости от стадии и тяжести АтД, позволяет нам проанализировать влияние определенных микроорганизмов на патологические процессы при АтД и установить предпосылки для открытия новых микробиологические предикторов обострения и ремиссии АтД.

Ключевые слова: атопический дерматит, микробиота ротоглотки, биологическое разнообразие, кожные болезни, иммунологические нарушения, микробиом.

1 Introduction

2 Atopic dermatitis (AtD) is a multifactorial inflammatory skin disease
3 characterized by itching, chronic recurrent course and age-related features of skin
4 lesions. This is one of the most common skin diseases (from 20% to 40% in the
5 structure of this group), emerging in all countries, mainly in young people of both
6 genders. The prevalence of AtD in Europe has amounted to 15.6%, in the USA –
7 17.2%, in Japan – 24%, in Russia – 30-35%, reflecting the steady increase in the
8 frequency of AtD detection over the past three decades [20, 28, 29].

9 The pathogenesis of AtD has not been fully elucidated yet. Presumably,
10 several factors might be the initiators of the inflammatory processes. One of such
11 factors is hereditary determinism, leading to a violation of the skin barrier, defects
12 in the immune system, hypersensitivity to allergens and non-specific stimuli,
13 colonization by pathogenic microorganisms, and also to an imbalance of the
14 autonomic nervous system with increased production of inflammatory mediators [3,
15 13].

16 According to the clinical recommendations of the American Academy of
17 Allergy, Asthma, and Immunology (AAAAI), AtD is a chronic inflammatory
18 process of the skin that emerges due to a genetic malfunction, under the influence of
19 external factors, a violation of the skin barrier and a defect in the immune defense
20 [19].

21 Recent scientific data increasingly indicate that cytokines play one of the main
22 roles in the AtD pathogenesis. In particular, much attention is paid to the
23 proinflammatory cytokines of the IL-36 subfamily: IL-36 α , IL-36 β and IL-36 γ and
24 their receptor antagonist IL-36Ra. These molecules are produced by keratinocytes,
25 Langerhans cells and macrophages and they serve as activators of the innate and
26 acquired defense systems [22, 32].

27 Another important factor contributing to the emergence and progression of
28 AtD is the imbalance of the symbiotic microbiota of the human body [2, 27].

29 As a rule, most researches, dedicated to relations between microorganisms
30 and allergic or autoimmune diseases, mainly concentrate on the microbiota of the
31 skin or intestines. At the same time, microbial communities of the upper respiratory
32 tract remain ignored. These anatomical structures are the most diverse and plastic in
33 the composition of the microbiome. The qualitative characteristics of the microbiota
34 in them varies depending on the biotope (nasal cavity, nasopharynx and oropharynx)
35 [5, 10].

36 Microbiota of the oropharynx is especially interesting in the context of our
37 topic, as it is the most abundant and diverse biotope. The surface of the tonsils is
38 characterized by a particularly high microbial diversity.

39 The scientific literature provides a certain amount of information about the
40 participation of microorganisms in the immunopathological processes. However,
41 only a small part of data on the involvement of individual oropharyngeal species in
42 the immunopathogenesis of AtD. This fact requires a more detailed further study of
43 the microbial diversity of the upper respiratory tract in patients with varying AtD
44 severity degrees.

45 Although much is known about the diagnosis and severity assessment of AtD,
46 there are too many ambiguous points about this disease. Additional knowledge about
47 the pathogenesis is needed in order to introduce different biomarkers into practical
48 work for more accurate diagnosis and monitoring of patients' condition. This
49 requires an integrated approach to the problem of AtD, which will take into account
50 various aspects of the pathogenesis of this disease.

51 Aim of the study is to analyze the biological diversity of oropharyngeal
52 microbial communities in groups of patients at different stages of AtD in order to
53 identify certain microbiological predictors.

54 **Materials and methods**

55 The study included 97 male AtD patients, aged from 16 to 19. 15 of them had
56 remission, 82 had an exacerbation of varying severity (22 – mild, 53 – moderate and

57 7 - severe). Only patients with no exacerbations of other chronic diseases were
58 included.

59 The semi-quantitative SCORAD (Scoring of Atopic Dermatitis) scale was
60 used to assess the severity of the skin pathological process. The SCORAD scale
61 provides a score assessment of six objective symptoms: erythema, edema/papular
62 elements, crusts/weeping, excoriation, peeling, dry skin.

63 Atopic dermatitis of mild severity corresponds to a SCORAD value <25,
64 moderate severity - 25-50, and severe atopic dermatitis corresponds to a SCORAD
65 value >50.

66 A smear was taken from the walls of the oropharynx for cultural examination
67 with a sterile cotton swab. In a tube with a liquid transport medium, the material was
68 delivered to a bacteriological laboratory. The material was seeded on the following
69 growth media: universal chromogenic agar (HiMedia, India), 5% blood agar with
70 mutton blood (HiMedia, India), chocolate agar (HiMedia, India), selective media for
71 the isolation of lactobacilli (HiMedia, India), bifidobacteria (HiMedia, India),
72 clostridium (HiMedia, India), obligate anaerobes (HiMedia, India), veilonella
73 (HiMedia, India), non-fermenting Gram-negative bacteria (HiMedia, India),
74 enterobacteria (HiMedia, India), Saburo agar (HiMedia, India).

75 The preparation of the material for seeding was carried out by homogenizing
76 it in a liquid Ames growth medium (GEM LLC, Russia), followed by spreading 100
77 µl of the final suspension on the surface of each growth medium.

78 Media were incubated in aerobic, microaerophilic (using a CO₂ incubator
79 (Sanyo, Japan) and anaerobic conditions (using gas-generating packages
80 (Anaerogaz, Russia), at a temperature of 37 ° C for 5 days.

81 Colonies of all grown microorganisms were identified using the MALDI-ToF
82 mass spectrometry on the Microflex LT device (Bruker, Germany) by direct
83 application and extended application with the use of formic acid. During
84 identification, the obtained spectra of microorganisms were compared with the
85 database of the Bruker Daltonik GmbH standard library. The accuracy of

86 identification was assessed automatically using the MALDI Biotyper RTC software
87 according to the level of the coincidence coefficient (Score) from 0 to 3. The level
88 of 0.000-1.699 was regarded as the result of low-confidence identification, the level
89 of Score from 1.700 to 1.999 was considered as identification on the level of genus;
90 highly reliable identification to the species level was accepted at Score values of
91 2,000-2.999.

92 To assess the biological diversity of the oropharyngeal microbiota, the
93 coefficient of constancy (C) was used. According to this assessment,
94 microorganisms were considered as participants of permanent, additional or
95 transient microbiota.

96 In the case of isolation of individual microorganisms from more than 50% of
97 patients, this microorganism was regarded as permanent. The isolation from patients
98 in the range of 25-50% corresponded to an additional microbiota, isolation less than
99 in 25% of cases corresponded to transient microbiota. Coefficient was calculated
100 using the following formula:

$$101 \quad C=(p*100)/P$$

102 In which p – number of isolations of individual microorganisms, P – total
103 number of isolations.

104 Accumulation, correction, systematization of the obtained data and
105 visualization of the results were carried out in Microsoft Office Excel 2016
106 spreadsheets. Statistical calculations were performed using the Stat Tech software
107 (version 4.0.0, Stattech LLC, Russia).

108 A predictive model, reflecting the dependence of a quantitative variable on
109 factors, was created using the linear regression method. The construction of a
110 predictive model of the possibility of a certain outcome was performed using the
111 logistic regression method. The Nigekirk coefficient R^2 served as a measure of
112 certainty, indicating the part of the variance that can be explained using logistic
113 regression.

114 For assessment of the diagnostic significance of quantitative signs, during
115 predicting a certain outcome, the ROC curve analysis method was used. The dividing
116 value of the quantitative feature at the cut-off point was determined by the highest
117 value of the Yuden index.

118 **Results**

119 During examination of the biological diversity of the oropharyngeal
120 microbiota in AtD patients, 58 species of microorganisms were isolated and
121 identified.

122 To assess the contribution of different species to biological diversity, for each
123 microbe the coefficient of constancy was calculated in three groups of patients with
124 different severity and in patients with remission.

125 Microbes of additional and permanent oropharyngeal microbiota are shown in
126 Figure 1.

127 The only species that can be classified as permanent for AtD patients with all
128 severity degrees was *Neisseria subflava*, which was isolated in 50.9-73.3% of the
129 examined individuals. For the rest of the microbes, three types of patterns were
130 identified: an increase of the coefficient of constancy with the transition from
131 transient to additional and permanent microbiota along with increase of AtD severity
132 (*Streptococcus vestibularis*, *Streptococcus mitis*, *Actinomyces oris*, *Rothia*
133 *mucilaginoso*); a decrease of the coefficient of constancy with transition to transient
134 species along with increase of AtD severity (*Streptococcus salivarius*, *Streptococcus*
135 *parasanguinis*, *Streptococcus oralis*, *Staphylococcus aureus*, *Rothia dentocariosa*);
136 the absence of significant changes in the coefficient of constancy depending on the
137 AtD severity (*Neisseria flavescens*).

138 For individual microorganisms of the first and second groups, significant
139 differences in frequency of isolation were obtained depending on the stage of AtD
140 (remission or exacerbation) (Table 1).

141 The isolation of *S. vestibularis* was significantly more often in the group of
142 patients with exacerbations of AtD, whereas *S. oralis* and *R. dentocariosa* were more
143 often isolated in patients with remission.

144 Above written data, on the one hand, shows the possibilities of the culture
145 method in assessing the biological diversity of the oropharyngeal microbiota. On the
146 other hand, it opens up opportunities for searching for new potential microbial
147 markers that determine the severity of AtD.

148 The species of microorganisms, which were isolated from the oropharynx
149 with statistically significant differences between patients with different stages and
150 severity degrees of AtD, were also considered as potential microbiological
151 predictors.

152 To predict the emergence of AtD exacerbation, mathematical models were
153 created, which were characterized by a higher quality of the prognostic test. These
154 models included such microbiological criteria as the isolation of *R. dentocariosa* and
155 *S. vestibularis* from the oropharynx.

156 For *R. dentocariosa*, the dependence with AtD stage is described by the
157 equation:

$$158 \quad P = 1 / (1 + e^{-z}) \times 100\%$$

$$159 \quad z = 1,83 - 1,83 \times R.d.$$

160 in which P – possibility of AtD exacerbation, *R.d.*– isolation of *R.*
161 *dentocariosa* from oropharynx (0 – not isolated, 1 – isolated).

162 Created regression model is statistically significant ($p=0.013$). When *R.*
163 *dentocariosa* is isolated from the oropharynx, the chances of AtD exacerbation
164 emergence decreased by 6 times, which is shown in Figure 2.

165 For *S. vestibularis*, the observed dependence is described by the equation:

$$166 \quad P = 1 / (1 + e^{-z}) \times 100\%$$

$$167 \quad z = 1,07 + 1,6 \times S.v.$$

168 in which P – possibility of AtD exacerbation, *S.v.* – isolation of *S. vestibularis*
169 from oropharynx (0 – not isolated, 1 – isolated).

170 Created regression model is statistically significant ($p=0.021$). When isolating
171 *S. vestibularis*, the chances of AtD exacerbation emergence, in opposite to *R.*
172 *dentocariosa*, increased by 5 times, which is shown in Figure 3.

173 Depending on the isolation of *S. vestibularis* from the oropharynx, a
174 prognostic model has also been developed to determine the possibility of emergence
175 of moderate AtD by binary logistic regression. The observed dependence is
176 described by the equation:

$$177 \quad P = 1 / (1 + e^{-z}) \times 100\%$$
$$178 \quad z = 0,39 + 1,73 \times S.v.$$

179 in which P – possibility of emergence of AtD with moderate severity, *S.v.* –
180 isolation of *S. vestibularis* from oropharynx (0 – not isolated, 1 – isolated).

181 Created regression model is statistically significant ($p = 0.004$). In the
182 presence of *S. vestibularis* in the oropharynx, the chances of emergence of AtD of
183 moderate severity increased in 5.7 times, which is shown in Table 2.

184 Discussion

185 The microbiota of the oropharynx is divided into permanent, additional, and
186 transient groups. As an example, *Streptococcus* spp. is a part of the permanent
187 microbiota; coagulase-negative staphylococci, *Corynebacterium* spp., *Haemophilus*
188 *influenzae* - additional microbiota (25-50% of people); *Enterobacteriaceae*,
189 *Pseudomonas* spp., *Moraxella* spp., *Micrococcus* spp. represent a transient group (5-
190 20% of people). The main flora of the tonsils consists of such microorganisms as:
191 *Staphylococcus* spp. (44.3%) and *Streptococcus* spp. (40.2%) [9].

192 The structure of oropharyngeal microbiota depends on the factors of the
193 pathogenicity of the commensals and on the nature of the interaction between
194 microorganisms and biotope, colonized by them. The published studies show that
195 *Staphylococcus* spp. and *Aerococcus* spp. are most likely to increase virulent
196 properties, and that an indifferent process is detected in the microbiota of a healthy
197 human body, when pathogenic commensals are stabilized by the eubiosis of the
198 tonsils of a healthy person [16].

199 In our study, we identified significant variations in the species composition of
200 the oropharyngeal microbiota. This fact allows us to think about its possible
201 functional connection with the emergence of atopic dermatitis. In addition, the
202 question arises whether changes in the microbiota and pathological processes in AtD
203 are interdependent, or is modified microbiocenosis a consequence of AtD?

204 Most of the studies, dedicated to relations between microorganisms and AtD,
205 focus on the skin or intestinal microbiota. The most common theory about the
206 influence of the human microbiome on the emergence of AtD is associated with the
207 dysbiosis in these loci, which in turn leads to the emergence of inflammatory
208 processes. Defects of the skin and intestinal barrier occur, which leads to the leakage
209 of different bacterial toxins and metabolites into the systemic bloodstream. Among
210 these harmful factors there are such as lipopolysaccharides, metabolic products of
211 tryptophan and serotonin, which can cause immunological dysfunctions [23, 31].

212 Significant number of researches concentrate on *S. aureus*, the number of
213 which increases significantly in the areas of skin lesions during AtD. As a rule, this
214 is associated with a reduced amount of an important structural skin protein -
215 phyllagrin [12, 17]. On the contrary, some authors consider the predominance of *S.*
216 *aureus* not only as a consequence of AtD. It is believed that its presence may have a
217 direct connection with the emergence of immunological disorders, especially
218 through the induction of synthesis of such factors as IL-31 and IL-33 [11]. Moreover,
219 increased expression of these molecules is not always associated with the death of
220 epithelial cells – it is associated with the direct presence of *S. aureus* at the locus [6].
221 It is noteworthy that in our study, with an increase in the severity of AtD, the
222 coefficient of constancy of *S. aureus* in the oropharynx decreased on the contrary.

223 The oropharyngeal locus is often not paid with enough attention from
224 researchers when studying AtD. And this is despite the fact that, on the one hand,
225 the oropharynx is one of the most microbial-populated biotopes in the human body,
226 and on the other hand, individual oropharyngeal microorganisms are very often
227 associated with other immunological (including allergic ones) disorders [14, 21, 30].

228 Perhaps this is due to the close relationship between the microbiota and the immune
229 system, mediated by colonization of the tonsils. Especially interesting is that tonsils
230 are normally colonized by such clinically important microbes in the context of AtD
231 as *Staphylococcus* spp. and *Streptococcus* spp.

232 Only individual attempts have been made to assess the biological diversity of
233 oropharyngeal microorganisms in relation to the AtD. In some early studies, there
234 was information about a direct correlation between the cutaneous and oropharyngeal
235 microbiota in patients with AtD [4]. Later, targeted studies were conducted to
236 compare the colonization of tonsils, affected and unaffected skin by *S. aureus*, but
237 the difference in results was almost not analyzed [7]. Beheshti et al. [8] conducted a
238 molecular genetic examination of patients with AtD. However, saliva was used as
239 the material, not smears or scrapings, and in addition, the study was linked only to
240 the total load of microbial RNAs and a correlation was found only with a large group
241 of bacteria – *Proteobacteria*.

242 The most interesting correlations with the stages of AtD in our study were
243 found for *Streptococcus* spp. Scientific works, which investigate the skin microbiota,
244 provided information about a decrease in the number of these microorganisms in the
245 species structure in AtD patients [17]. On the contrary, studies of the correlation
246 between the intestinal microbiota and AtD associate the *Streptococcus* spp. with the
247 onset and progression of this disease [18, 26]. In our study, controversial correlations
248 were established for individual microbes from this genus. Summarizing all the
249 information provided, it is possible to indicate a heterogeneous immunological effect
250 of *Streptococcus* spp.

251 Regarding the effect of *R. dentocariosa* (which in our study was correlated
252 with a more favorable course of the disease) on the emergence of AtD, no scientific
253 publications were found.

254 Many allergic diseases are characterized by a pathogenetic relation with the
255 microbiota and the presence of certain microbiological predictors. In the context of
256 AtD, this is the above mentioned increase of *S. aureus* in the composition of the skin

257 microbiota. The intestinal microbiota in AtD, in addition to the already described
258 role of *Streptococcus* spp., is characterized by an increased contribution to the
259 species structure from various opportunistic flora (*Parabacteroides* spp.,
260 *Clostridium difficile*, *Escherichia coli*) and a decrease in the number of
261 *Lactobacillus* spp. and *Bifidobacterium* spp. [1, 15].

262 The oropharyngeal microbiota so far lacks microorganisms that could
263 definitely be called predictors of AtD, or predictors of exacerbation or remission.
264 Therefore, the patterns identified in our study should be further analyzed, and the
265 search for such correlations should be continued.

266 **Conclusion**

267 Therefore, we have identified significant correlations between the various
268 stages of AtD and the frequency of isolation of individual oropharyngeal
269 microorganisms. Taking into account our data, as well as data on the possible
270 influence of oropharyngeal flora on various immunological processes, we believe
271 that it is necessary to continue work in this direction in order to identify additional
272 microbiological predictors of AtD and its exacerbations. Moreover, studies with a
273 larger studied groups are needed, including a comparison of the microbial diversity
274 between different loci and a deeper analysis of their differences, as well as taking
275 into account various clinical and laboratory parameters, including the level of
276 interleukin expression.

ТАБЛИЦЫ

Table 1. Analyzed frequency of AtD stage-related isolation for individual oropharyngeal microorganisms.

Species	Result of culture study	AtD stage		p
		Remission abs. (%)	Exacerbation abs. (%)	
<i>S. vestibularis</i>	Isolated	2 (13,3)	36 (43,9)	0,039*
	Not isolated	13 (86,7)	46 (56,1)	
<i>S. oralis</i>	Isolated	6 (40,0)	13 (15,9)	0,042*
	Not isolated	9 (60,0)	69 (84,1)	
<i>R. dentocariosa</i>	Isolated	5 (33,3)	6 (7,4)	0,016*
	Not isolated	10 (66,7)	76 (92,6)	
abs. – absolute number; * – significant differences at $p < 0,05$				

Table 2. Features of correlation between *S. vestibularis* isolation and odds for AtD moderate emergence.

Predictors	Unadjusted		Adjusted	
	COR; 95% CI	p	AOR; 95% CI	p
Isolation of <i>Streptococcus vestibularis</i> from oropharynx	5,655; 1,493 – 21,413	0,011*	5,655; 1,493 – 21,413	0,011*

* – significant predictor’s influence ($p \leq 0,05$); CI – confidence interval; Unadjusted – odds ratio is unadjusted; Adjusted – odds ratio is adjusted; COR – crude odds ratio (rough odds ratio), i.e. the odds ratio calculated for one of the factors without taking into account the influence of other factors; AOR – adjusted odds ratio (corrected odds ratio), i.e. odds ratio calculated for one of the factors, taking into account the influence of other factors.

РИСУНКИ

Figure 1. AtD severity-driven Species diversity for permanent and additional oropharyngeal microbiota.

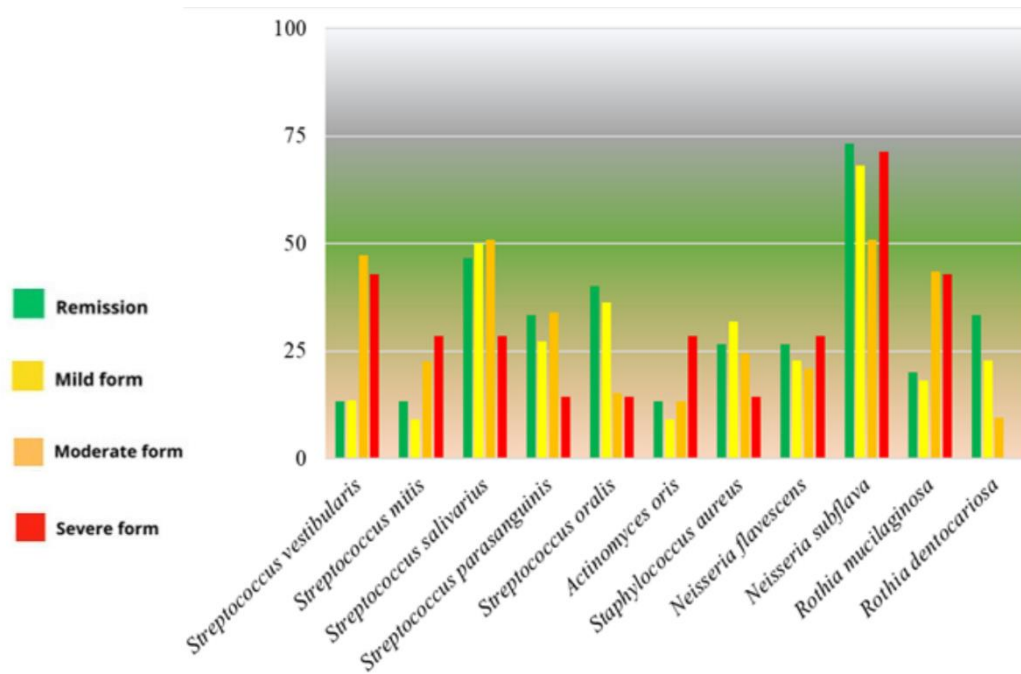


Figure 2. Assessed odds ratio with 95% confidence intervals for isolation of oropharyngeal *R. dentocariosa* as a predictor of emerging AtD exacerbation.

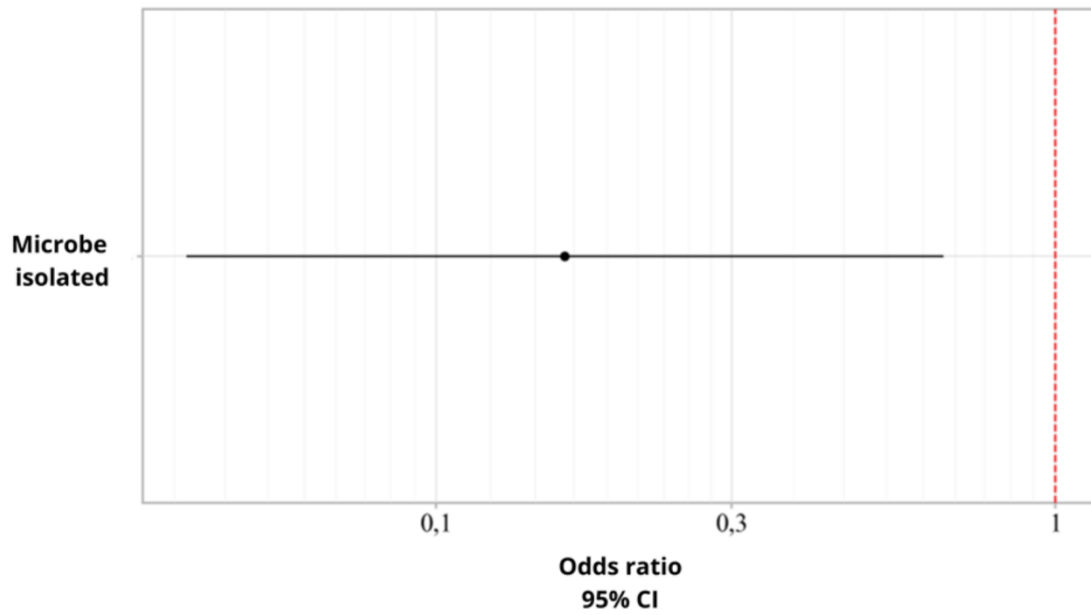
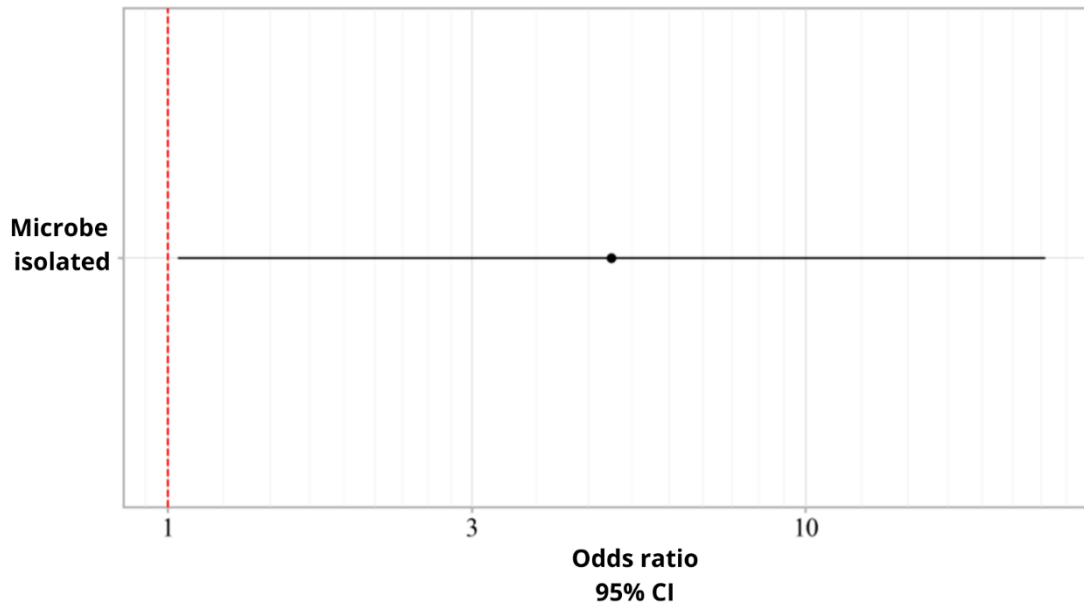


Figure 3. Assessed odds ratio with 95% confidence intervals for isolation of oropharyngeal *S. vestibularis* as a predictor of emerging AtD exacerbation.



ТИТУЛЬНЫЙ ЛИСТ_МЕТАДААННЫЕ

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Блок 3. Метаданные статьи

ASSESSMENT OF CORRELATION OF BIOLOGICAL DIVERSITY OF
OROPHARYNGEAL MICROBIOTA WITH SEVERITY AND
EXACERBATIONS IN PATIENTS WITH ATOPIC DERMATITIS

ОЦЕНКА КОРРЕЛЯЦИИ БИОЛОГИЧЕСКОГО РАЗНООБРАЗИЯ МИКРОБИОТЫ
РОТОГЛОТКИ СО СТЕПЕНЬЮ ТЯЖЕСТИ И ЧАСТОТОЙ ОБОСТРЕНИЙ У
ПАЦИЕНТОВ С АТОПИЧЕСКИМ ДЕРМАТИТОМ

**ASSESSED CORRELATION BETWEEN BIOLOGICAL DIVERSITY OF
OROPHARYNGEAL MICROBIOTA AND ATOPIC DERMATITIS
SEVERITY AND EXACERBATIONS**

Сокращенное название статьи для верхнего колонтитула:

МИКРОБИОТА РОТОГЛОТКИ У ПАЦИЕНТОВ С АТОПИЧЕСКИМ
ДЕРМАТИТОМ

OROPHARYNGEAL MICROBIOTA IN PATIENTS WITH ATOPIC
DERMATITIS

Ключевые слова: атопический дерматит, микробиота ротоглотки,
биологическое разнообразие, кожные болезни, иммунологические нарушения,
микробиом.

Keywords: atopic dermatitis, oropharyngeal microbiota, biological diversity, skin
diseases, immunological disorders, microbiome.

Оригинальные статьи.

Количество страниц текста – 11, количество таблиц – 2, количество рисунков
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