PREDICTION OF INFLAMMATION IN HEMODIALYSIS PATIENTS USING NEURAL NETWORK ANALYSIS

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НЕЙРОСЕТЕВОЙ АНАЛИЗ В ПРОГНОЗИРОВАНИИ ВОСПАЛЕНИЯ У ГЕМОДИАЛИЗНЫХ ПАЦИЕНТОВ

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Abstract

Background. Numerous hemodialysis patients (HD) suffer from severe, lifethreatening inflammation that must be treated to prevent further complications. Early diagnosis of inflammation in HD is highly needed. The present study used matrix metalloproteinase-1 (MMP3) and tissue inhibitor of metalloproteinases-1 (TIMP1) to differentiate between patients with/without inflammation using the neural network analysis (NN).

Methods. The positive results of C-reactive protein were used as a criterion for the presence of inflammation in the patients (HD+CRP) versus the negative group (HD-CRP). The NN analysis was used to discriminate between groups using the measured biomarkers.

Results. HD+CRP patients have a higher duration of disease, MMP3 and lower calcium than the HD-CRP level is significantly higher, while vitamin D is significantly lower in the HD+CRP group compared with both other groups (all p<0.05). TIMP1 is significantly correlated with inorganic phosphate and CRP. In NN#1, the model for the prediction of HD+CRP from HD-CRP has an area under the curve (AUC) of the receiver operating characteristic (ROC) of 0.907 with a sensitivity and specificity 89.2% and a specificity of 100.0%. The top predicting variable for the prediction of HD+CRP is MMP3 (100%), followed by creatinine (87.1%). MMP3 is linked to the pathophysiology of HD, at least through their correlation with the inflammation in HD. In NN#2, the AUC of the ROC for predicting the kidney disease and subsequent HD was 98.9%, with a sensitivity of 100.0% and a specificity of 97.1%. The top four predicting variables for the prediction of high risk of inflammation in HD patients are urea (100%), creatinine (100%), MMP3 (59.7%), and vitamin D (57.1%).

Conclusion. The NN analysis may differentiate between HD patients with inflammation from the HD without inflammation. Also, the measured parameters,

especially MMP3, TIMP1, and vitamin D are useful as a diagnostic tools for the kidney diseases and inflammation linked with the disease.

Keywords: Hemodialysis patients (HD), Tissue inhibitor of metalloproteinases-1 (TIMP1), matrix metalloproteinase-1 (MMP3), vitamin D, neural network, and inflammation.

Резюме.

Многие пациенты, находящиеся на гемодиализе (ГД), страдают от тяжелого, опасного для жизни воспаления, которое необходимо лечить для предотвращения дальнейших осложнений. Крайне необходимо проведение ранней диагностика воспаления при ГД. Для разделения пациентов с воспалением и без него в настоящем исследовании использовалась показатели металлопротеиназы-1 (MMP3) И тканевого металлопротеиназ-1 (TIMP1) с использованием анализа нейронных сетей (HC).

Методы. Положительные результаты оценки уровня С-реактивного белка использовали в качестве критерия наличия воспаления у пациентов (ГД+СРБ) по сравнению с отрицательной группой (ГД-СРБ). Анализ НС использовался для разделения групп на основании применяемых биомаркеров.

Результаты. Пациенты HD+CRP имеют более высокую продолжительность заболевания, ММРЗ и более низкий уровень кальция, по сравнению с группой HD-CRP, уровень витамина D значительно ниже в группе HD+CRP по сравнению с обеими другими группами (все p<0,05). ТІМР1 достоверно коррелирует с уровнем неорганического фосфата и СРБ. В HC#1 модель прогнозирования HD+CRP на основе HD-CRP имеет площадь под кривой (AUC) рабочей характеристики приемника (ROC) 0,907 с чувствительностью и специфичностью 89,2% и специфичностью 100,0%

соответственно. Главной прогностической переменной для прогнозирования HD+CRP является уровень MMP3 (100%), а также и уровень креатинина (87,1%). ММРЗ связана с патофизиологией ГБ, по крайней мере, через их корреляцию с воспалением при ГБ. В HC#2 AUC ROC для прогнозирования заболевания почек и последующей ГБ составила 98,9% при чувствительности 100,0% и специфичности 97,1%. Четырьмя ведущими прогностическими параметрами для прогнозирования высокого риска воспаления у пациентов с Γ Б являются уровень мочевины (100%), креатинина (100%), MMP3 (59,7%) и витамина D (57,1%).

Заключение. Анализ НС может разграничивать пациентов с ГБ с воспалением и без него. Кроме того, измеряемые параметры, особенно ММРЗ, TIMP1 и витамин D, полезны в качестве диагностических инструментов заболеваний почек и сопутствующего воспаления.

Ключевые слова: пациенты, находящиеся на гемодиализе (ГД), тканевой ингибитор металлопротеиназы-1 (TIMP1), матриксная металлопротеиназа-3 (MMP3), витамин D, нейронная сеть и воспаление.

Introduction

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There is a growing increase in patients receiving long-term hemodialysis (HD) for end-stage renal disease (ESRD) [45]. Patients with ESRD have a higher risk of cardiovascular disease and other coexisting diseases [11, 33] and an adjusted allcause mortality rate at least 10-fold higher than that of the non-ESRD population [45]. As such, perioperative management of patients with HD requires special regarding considerations disease pathophysiology, including cardiovascular volume disturbances, electrolyte disorders, dysfunction, anemia, and pharmacokinetics/pharmacodynamics alterations [21].

Several types of cellular injury occur in AKI, including necrosis, apoptosis, or necroptosis combined. This latter type of cellular injury is a highly immunogenic form of programmed cell death that normally represents a defense against viruses expressing caspase-8 inhibitors but may also be triggered by cytokine imbalance [8]. HD remains the most specific and clinically relevant endpoint for patients with chronic kidney disease (CKD) [2]. Poor nutritional status is frequently observed in HD patients and is associated with adverse clinical outcomes and increased mortality. Loss of amino acids during HD may contribute to protein malnutrition in these patients [18].

Matrix metalloproteinases (MMPs) represent a family of dependent metal ion capable endopeptidases of degrading all extracellular matrix (ECM) components. MMPs are classified by substrate specificity into collagenases, gelatinases, stromelysins, and membrane-bound types. MMP expression is regulated by cytokines [28].

Matrix metalloproteinase 3 (MMP3) is well-known as a secretory endopeptidase that degrades extracellular matrices [14]. MMP3 is an important member of a large family of MMPs containing zinc-dependent endopeptidases. Matrix degradation and remodeling have been recognized as the main function of MMPs. However, subsequent studies revealed that MMPs might participate in 29

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diverse pathophysiological processes, such as the regulation of inflammatory and immune responses as well as cell-cell communication, among others [47]. MMP3 is an important member of a large family of MMPs containing zinc-dependent endopeptidases. Matrix degradation and remodeling have been recognized as the main function of MMPs. However, subsequent studies revealed that MMPs might participate in diverse pathophysiological processes, such as the regulation of inflammatory and immune responses as well as cell-cell communication, among others [22, 29, 54]. MMPs participate in many physiological and pathological processes associated with the inflammatory process [47].

Tissue inhibitor of metalloproteinases-1 (TIMP1) is a founding member of the TIMP family that comprises four members, TIMP1 to TIMP4, which as a whole act as major inhibitors of metalloproteinases including the matrix metalloproteinases (MMPs) and members of a disintegrin and metalloproteinase domain (ADAM) family of proteases [53]. The results of this research indicate that increased TIMP1 level is an independent predictor of an increase in hospitalization and mortality of patients with congestive heart failure (CHF) [57]. The significant correlation between TIMP1 expression and the presence of lymph node metastases, as well as that between TIMP1 plasma concentration and stage of cancer histological differentiation, might indicate the importance of this molecule as a prognostic factor during carcinogenesis [30]. MMPs and TIMPs are considered important mediators of the periapical immune response to infection [51]. Hypertension is a leading risk factor for cardiovascular disease. MMPs and their tissue inhibitors are thought to be actively involved in remodeling the CV extracellular matrix during hypertensive damage [24]. The present study aims to use neural network analysis for the prediction of overt inflammation (positive serum CRP test) in hemodialysis patients by entering the clinical and biochemical biomarkers in the analysis set.

Subjects and Methods

Patients

The present study involved a total of sixty patients diagnosed with chronic 57 HD, as well as thirty healthy controls. The patients group was divided into two 58 categories based on the results of C-reactive protein (CRP) levels. Thirty 59 hemodialysis (HD) patients with evident inflammation were categorized as 60 HD+CRP, while thirty HD patients without inflammation were categorized as HD-61 CRP. The specimens were collected from Al-Sader medical city in Najaf 62 governorate-Iraq from November 2021 to March 2022. Patients were under 63 hemodialysis and previously diagnosed by a specialist following the International 64 Statistical Classification of Diseases and Related Health Problems, 10th Revision, 65 criteria (2021 ICD-10-CM Diagnosis Code N18.6). The Urologist and Internists 66 performed patients' diagnoses according to clinical signs and laboratory tests. 67 According to the used definition, the patients were having ESRD requiring chronic 68 dialysis. All patients have elevated urea and creatinine, electrolyte disturbances, with 69 eGFR less than 15 ml/minute. A full medical history and examination to explore the 70 presence of any systemic diseases that might affect the studied parameters; diabetes, 71 liver, and heart diseases were excluded from the study. All patients were given 72 calcium carbonate, epoetin alpha (Eprex®), heparin, and either continuous folic acid 73 or iron and folate formula (Fefol®). Thirty apparently healthy subjects were 74 classified as a control group. Their age and sex ratios were comparable to both 75 patient groups. Subjects were selected to be free of kidney disease or other systemic 76 or inflammatory disorders. Approval for the study was obtained from the IRB of the 77 University of Kufa (T1375/2020), which complies with the International Guidelines 78 for Human Research Protection as required by the Declaration of Helsinki. 79

Measurements

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Following overnight fasting between 7:00-10:00 a.m., five milliliters of venous blood were withdrawn utilizing a disposable syringe and transferred directly to a serum gel tube. All samples were incubated for 10 minutes at room temperature before centrifugation for 5 minutes at 3500 rpm. Then, we distributed the serum into

a small Eppendorf and stored it at -80 °C until the measurement time. Melsin 85 Medical Co., Ltd., Jilin, China, provided ELISA kits to assess the sera's MMP3, 86 TIMP1, and vitamin D levels. Serum creatinine, uric acid, urea, phosphorus, 87 glucose, calcium, magnesium, and albumin were determined spectrophotometrically 88 using kits supplied by Agappe Diagnostics Ltd., Cham, Switzerland. Serum CRP 89 was measured semi-quantitatively by a kit supplied by Spinreact[®], Spain, utilizing 90 an agglutination test that produced a positive result when the CRP level in serum 91 was higher than 6 mg/L. The following equation was used to calculate the estimated 92 93 glomerular filtration rate (eGFR):

eGFR = $175 \times (S_{c}Cr)^{-1.154} \times (Age)^{-0.203} \times 0.742$ [if female] x 1.212 [if Black] which is derived from the Modification of Diet in Renal Disease (MDRD) study equation [26]. To get the body mass index, we multiplied each individual's weight in kilos by their height in meters squared (BMI).

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Statistical analysis

We used analysis of variance (ANOVA) to assess differences in continuous variables between categories and analysis of contingency tables (χ^2 -test) to check associations between categorical variables. Fisher's Least Significant Difference (LSD) Post Hoc Test analysis was done to compare the levels of the measured parameters among the three study groups. Kruskal–Wallis test was used to compare the not normally distributed variables among the three groups measured by Kolmogorov-Smirnov for normality testing. Multiple comparisons were examined using a p-correction for false discovery rate (FDR) [5]. Spearman's correlation coefficients were calculated for the correlation study of MM3, TIMP1, and vitamin D with other measured parameters. Multilayer perceptron Neural Network (NN) models (IBM SPSS Windows version 25, 2017) were used to delineate the more complex relationships between biomarkers (entered as input variables) in predicting the diagnostic classes (HD with inflammation (HD+CRP) versus HD without inflammation (HD-CRP)) as well as HD versus healthy controls). The same input variables were entered as input variables in predicting the presence of overt inflammation (HD+CRP) versus patients with no inflammation (HD-CRP). The models were trained using an automated feed-forward architecture with two hidden layers with up to 8 nodes in each layer, employing minibatch training with gradient descent, 250 epochs, and one consecutive step with no further decrease in the error term as a stopping rule. For NN#1, we considered three samples, i.e., "a training sample to estimate the network parameters (50.5% of all participants), testing set to prevent overtraining (36.7%) and a holdout set to evaluate the final network (13.3%). For NN#2, we considered three samples, i.e., a training sample to estimate the network parameters (68.9% of all participants), a testing set to prevent overtraining (20.0%), and a holdout set to evaluate the final network (11.1%). Error, relative error, and importance and relative importance of all input variables were computed.

Results

Demographic and Clinical data

Table 1 presents the demographic and clinical data of the HD+CRP, HD-CRP, and the healthy controls group. The results showed no significant difference in the demographic characteristics (age, sex ratio, TUD, family history, albumin, T.Mg, ionized Mg, T.Ca/Mg, TIMP1, and IonizedCa/Mg, and tobacco use disorder (TUD)) among the three groups. HD+CRP patients have a higher duration of disease than HD-CRP. Total and ionized calcium are significantly lower in HD+CRP than in the HD-CRP group. MMP3 level is significantly higher, while vitamin D is significantly lower in the HD+CRP group compared with both groups. BMI is significantly lower in patient groups than in the control group. Serum urea, creatinine, inorganic phosphate (Pi), uric acid, and glucose are significantly higher in HD groups compared with the control groups.

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Correlation between Stromelysin-1, TIMP1, and TIMP1/Stromelysin-1 with all parameters

The correlations of vitamin D, MMP3, and TIMP1 with other biomarkers are presented in Table 2. TIMP1 is significantly correlated with Pi (ρ =0.222, p<0.05) and CRP (p=0.279, p<0.01). Vitamin D is significantly correlated with BMI $(\rho=0.216, p<0.05)$, total calcium $(\rho=0.215, p<0.05)$, and ionized calcium $(\rho=0.222, p<0.05)$ p<0.05). While vitamin D is inversely correlated with duration of HD (ρ =-0.603, p<0.001), urea (ρ =-0.482, p<0.01), creatinine (ρ =-0.518, p<0.001), Pi (ρ =-0.552, p<0.001), CRP ($\rho=-0.507$, p<0.001), and MMP3 ($\rho=-0.221$, p<0.05). MMP3 showed significant correlations with urea (ρ =0.273, p<0.01), creatinine (ρ =0.238, p<0.05), Pi (ρ =0.324, p<0.01), and CRP (ρ =0.425, p<0.01).

Neural Network study

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The results of two neural network information of the model on HD patients for predicting HD patients with inflammation (HD+CRP) versus HD-CRP patients are presented in Table 3. The NN analysis used feed-forward architecture because the network connections flow from the input layer to the output layer without any feedback loops. In this analysis, the input layer contains the predictors. The hidden layer contains unobservable nodes or units. The value of each hidden unit is some function of the predictors; the exact form of the function depends in part upon the network type and in part upon user-controllable specifications. The last layer is the output layer contains the responses. Since the history of default is a categorical variable with two categories, it is recorded as two indicator variables. Each output unit is some function of the hidden units. Again, the exact form of the function depends partly on the network type and controllable specifications. There are 11 units (measured parameters) in the input layer (layer containing factors for predicting HD from control and patients with inflammation).

In NN#1, the hyperbolic tangent and identity were used as activation functions in the hidden layers, and identity was used in the output layer to train this model, which has two hidden layers with two units in layer 1 and two units in layer 2. The area under the curve (AUC) of the receiver operating characteristic (ROC) was 0.907, with a sensitivity of 89.2% and a specificity of 100.0%, in each of the three sets of data. These results showed the model's poor sensitivity in predicting HD+CRP without entering CRP as an input factor. However, Figure 1 shows the significance of each model's input variable in terms of the model's predictive ability. In terms of predictive capability, the top four predicting variables (effect >50%) for the prediction of high risk of inflammation in HD patients are MMP3 (100%) followed by creatinine (87.1%), duration of disease (73.0%), and total calcium (70.7%).

In NN#2, two hidden layers with four units in layer 1 and three in layer 2 were used. The AUC of the ROC was 98.9%, with a sensitivity of 100.0% and a specificity of 97.1%, in each of the three sets of data. These results showed a great sensitivity of the model in predicting HD patients from the control group. The top four predicting variables for the prediction of high risk of inflammation in HD patients are urea (100%), creatinine (100%), MMP3 (59.7%), and vitamin D (57.1%), as presented in Figure 2.

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Discussion

Comparison study

Beyond the routinely increased parameters in HD, Table 1 shows that patients with higher disease duration have more inflammation. The longer duration of the disease is associated with inflammation [39]. It is suggested that inflammatory status and duration of dialysis treatment are the most important factors relating to oxidative stress in HD patients [36]. The greater serum creatinine levels and a longer 194

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duration of illness were associated with larger tubulointerstitial inflammatory cell infiltrates in CKD and diabetic nephropathy in human kidney biopsy specimens [7].

Total and ionized calcium are significantly lower in HD+CRP than in the HD-CRP group. Serum urea, creatinine, uric acid, potassium and phosphate levels, and urine proteins were significantly higher, while serum albumin and calcium were significantly lower in CKD patients [10]. Abnormal calcium and phosphate metabolism have been proposed to explain this greater risk of CVD [46]. low PTH and calcium levels are associated with mortality [4]. Vascular calcification was considered an imbalance between the inhibitors and promoters of osteogenesis initiated in vessels by uremic factors of CKD patients [55]. Consistently, the risk of cardiovascular death associated with hyperphosphatemia is attenuated among hemodialysis patients with high serum magnesium levels, whereas this risk is exacerbated among low serum magnesium levels [44].

Due to low serum calcium, CKD patients begin dialysis with vitamin D supplementation, calcium-based phosphate binders, and dialysate calcium. Dialysis increases serum calcium levels [31]. However, serum phosphate levels rose throughout this time, and comorbidity was related to higher calcium and phosphate levels [31]. In a common population, long-term dialysis users had increased phosphate levels [6]. Vitamin D drugs like calcitriol improve intestine absorption of serum phosphate, which rises the following dialysis. Loss of residual renal function may increase phosphate levels [13].

Another important finding of the present study is the increase in MMP3 in HD patients with inflammation compared to the controls. Albumin increases TIMP1 production [40]. Therefore, the lack of significant difference between study groups may be due to the compensation of the possible increase in TIMP1 by the decrease in albumin level in HD patients. Previous work showed that increased TIMP1 level is an independent predictor of increased hospitalization and mortality of patients with CHF regardless of renal function [24].

Therefore it is not dependent on renal function and not increased in HD patients as seen in our research. However, an increase in TIMP1 level is associated with the development of endothelial dysfunction in both groups [34]. Evidence suggests that MMP3 plays an inductive role in acute kidney injury induced by ischemia and reperfusion [27]. MMP3 level is significantly higher, while vitamin D is significantly lower in the HD+CRP group compared with both groups. BMI is significantly lower in patient groups than in the control group. Serum urea, creatinine, Pi, uric acid, and glucose are significantly higher in HD groups compared with the control groups. Serum urea, creatinine, uric acid, potassium and phosphate levels, and urine proteins were significantly higher, while serum albumin and calcium were significantly lower in CKD patients [10].

MMP9 and TIMP1 were elevated in renal patients compared to controls. Logistic regression analyses disclosed galectin-3, MMP9, pentraxin-3, and glomerular filtration associations with calculated CVD risk scores. Combined testing of pentraxin-3, galectin-3, MMP9, and glomerular filtration rate can discriminate among renal patients with high and low risk of coronary events [32]. The MMP3 level higher than 9.3 ng/mL had a lower survival rate. MMP3 baseline level in patients with a history of CAD is a potential predictor for cardiovascular outcomes [16].

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Correlation study

The correlation study in Table 2 showed various correlation coefficients that, in general, are produced by the effect of vitamin D or MMP3 and its inhibitor TIMP1 and their effect on the inflammation and overall health status of HD patients. There was a positive correlation between glomerular filtration rate and MMP3 activity in diabetic patients. Thus MMP3 may have a role in the pathogenesis of diabetic nephropathy progressions toward macroalbuminuria, and therefore, MMP3 activity may be used in evaluating albuminuria status [3]. The correlation analysis

MMP3 correlated significantly with biological parameters showed that 250 with uric acid [16]. A previous study showed a negative correlation between the 251 eGFR and MMP2, MMP3, and TIMP2 and a positive correlation between creatinine 252 and MMP3 levels, indicating the role of MMPs and TIMP2 in renal dysfunction. 253 The serum level of urea is correlated with MMP3 [23]. Calcium signaling is critical 254 for the proteolytic activity of MMP3 [17], two putative Ca²⁺ binding sites were found 255 in the catalytic domain of MMP3 and several other members of the MMP gene 256 family. These putative Ca²⁺ binding sites are postulated to play an important role in 257 stabilizing active MMP3 and other members of the MMPs gene family by protecting 258 them against autolysis [19]. 259

Previously, inflammatory response and MMP genes were modulated by the dropin and spexin that protect against inflammation and CKD [58]. MMP3 serum levels increase in parallel with the elevated circulating levels of IL-6. Serum MMP3 may be a useful predictor of chronic inflammation and osteoarticular disorders in dialysis-related amyloidosis patients [20]. Studies have shown that MMP2, MMP9, and TIMP1 and -2 also play an important role in the pathogenesis of renal damage [15]. A negative correlation between the eGFR and MMP2, MMP3, and TIMP2 and a positive correlation between creatinine and MMP3 levels indicate the role of MMPs and TIMP2 in renal dysfunction [23]. MMP3 is associated with inflammation, and most inflammatory disorders are associated with changes in MMP3 [25,48, 52. 59]. Inorganic phosphate (Pi) significantly increased MMP3 protein as a signaling molecule [43]. It appears that serum levels of MMP3 reflect positively rheumatoid arthritis disease activity, joint and bone injury, and radiological erosion and predict disease outcome and drug responsiveness [25]. Also, MMP3 is associated with calcium levels, and serum MMP3 levels may be used as an indicator for structural damage, such as erosions in the early stages of the disease, and to monitor disease activity [1, 49]. The data indicated measurable differences in the expression of MMPs within the dialysis

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patient population. Because dialysis can be associated with local and systemic inflammation, increased levels of MMP3 in the hemodialysis group may reflect gene stimulation induced by inflammatory cytokines and should be considered a marker of chronic, local inflammation [37]. MMP3 significantly and positively correlated with serum creatinine [41]. The mean expression of MMP2, MMP9, TIMP1, ADAMTS-1, and FSP-1 was significantly higher in the fibrotic kidney compared with the normal kidney [56].

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The NN analysis

The other important findings of the present study are the results of NN studies in Table 3. The measured parameters have a moderate sensitivity with excellent specificity for the prediction of HD+CRP versus HD-CRP. Figure 1 shows the top four predicting variables for predicting a high risk of inflammation in HD patients, which are MMP3 followed by creatinine, duration of disease, and total calcium. While NN#2 showed a great sensitivity of the model in predicting HD patients from the control group with the usual biomarkers of HD (urea and creatinine). However, MMP3 and vitamin D also act as possible predictive variables. Various metabolites may generate or be absorbed due to elevated serum urea levels, which probably lead to malnutrition, inflammation, and uremic toxicity [12]. TIMP1 is expressed in human glomeruli and is upregulated in glomerulosclerosis [9]. In clinical studies, patients with diabetic kidney diseases have been shown to have abnormalities in MMP/TIMP modulation. In patients with DKD, increasing glomerular lesions have been associated with reductions in serum TIMP1 and TIMP2 levels and increases in serum and urine TIMP1 levels [35, 42]. The induction of the decrease in serum MMP9 and MMP3 levels is one of the possible mechanisms responsible for the decrease in urea levels [50]. There was a significant positive correlation between the total score of kidney injury molecule 1 (KIM-1) expression and kidney function parameters for AKI, including serum creatinine and blood urea.

In addition, strong positive correlations were found between the total score of KIM-1 expression and proximal tubular necrosis and MMP3 expression. The KIM-1 shedding might be stimulated by MMP3 [38].

Conclusion

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The NN model can predict the existence of inflammation in HD patients with 310 a 89.2% sensitivity and 100.0% specificity utilizing the impacts of MMP3 (100%) 311 and creatinine (87.1%). Compared to the other groups, inflammation is linked to 312 prolonged disease duration, higher MMP3 levels, lower total and ionized calcium, 313 314 and lower vitamin D levels. TIMP1 and CRP positivity are related. MMP3 and HD duration are negatively affected by vitamin D. Significant correlations between 315 MMP3 and urea, creatinine, and CRP were found. The measured values have 100% 316 sensitivity and 97.1% specificity for predicting HD. MMP3 and HD inflammation 317 are related. At the very least, via their relationship with the inflammation in HD, 318 MMP3 is connected to the pathogenesis of the disease. 319

Declaration of interest

The authors have no financial or any conflict of interest.

Funding 322

There was no specific funding for this specific study. 323

Authorships

All authors contributed significantly to the paper and approved the final version.

Acknowledgments

The authors wish to express their gratitude for the highly skilled work of the Asia Laboratory's staff in measuring the biomarkers.

ТАБЛИЦЫ

Table 1. Demographic and clinical data of healthy controls (HC) and HD patients.

Variables	HC A	HD-CRP B	HD+CRP C	F/χ^2	р
Variables	n=30	n=28	n=32	Ι / λ	Р
Age Yr.	47.27±7.177	45.93±8.959	46.83±11.390	0.159	0.853
Sex (Female/Male)	10/20	13/15	16/16	1.910	0.385
Duation of HD Yr.	-	2.743±2.751 ^C	3.293±2.684 ^B	12.238	< 0.001
BMI kg/m ²	28.353±6.241 ^{B, C}	24.717±4.272 ^A	25.092±3.687 ^A	5.085	0.008
Smoking (Yes/No)	29/1	27/1	31/1	0.009	0.995
Family history N/Y	30/0	26/2	28/4	3.903	0.142
Creatinine mg/dl	0.710(0.460-1.011) ^{B, C}	8.600(2.500-11.700) ^A	8.500(6.400-10.800) ^A	KWT	< 0.001
Urea mg/dl	26.500(23.00-35.000) ^{B, C}	151.500(65.000-178.000) ^A	156.000(146.000-183.000) ^A	KWT	< 0.001
Pi mg/dl	5.052±0.782 ^{B, C}	6.883±0.981 ^A	7.386±0.873 ^A	58.139	< 0.001
Uric acid mg/dl	4.733±0.946 ^{B, C}	5.723±1.631 ^A	$5.480\pm1.578^{\text{ A}}$	3.472	0.037
Glucose mM	5.415±0.783 ^{B, C}	5.624±0.634 ^A	6.097±1.098 ^A	4.951	0.009
Albumin g/l	43.426±6.800	43.858±6.360	46.474±7.036	1.798	0.172
Magnesium mM	0.850±0.256	0.898 ± 0.220	0.882 ± 0.224	0.328	0.721
Ionized Mg mM	0.600±0.169	0.632±0.145	0.621 ± 0.148	0.328	0.721
Calcium mM	2.246±0.171 ^B	2.224±0.167	2.141±0.185 ^A	3.321	0.046
Ionizad Ca mM	1.195±0.047 ^B	1.184 ± 0.044	1.166±0.052 ^A	3.318	0.047
Total Ca/Mg	2.952±1.156	2.568 ± 0.888	2.683±0.710	1.329	0.270
Ionized Ca/Mg	2.188±0.762	1.960±0.562	2.012±0.179	1.140	0.324
Vitamin D ng/ml	10.829(9.769-12.242) ^{B, C}	8.329(7.459-8.954) ^A	$7.772(6.957-9.097)^{A}$	KWT	< 0.001
MMP3 ng/ml	46.501(27.977-73.388) ^C	56.801(29.611-108.709) ^C	120.654(75.062-137.677) ^{A, B}	KWT	< 0.001
TIMP1 ng/ml	530.356(154.406-876.295)	723.397(174.315-1032.735)	693.449(386.984-878.771)	KWT	0.556
eGFR ml/min	108.073((91.627-120.676) ^{B, C}	7.029(4.744-10.885) ^A	6.432(5.101-10.277) ^A	KWT	< 0.001

 A,B,C : Pair-wise comparison, BMI: Body mass index, Pi: inorganic phosphate, KWT: Kruskal-Wallis test, eGFR: estimated glomerular filtration rate, MMP3: matrix metalloproteinase-3, TIMP1: tissue inhibitor of metalloproteinases-1. Results are expressed as mean \pm standard deviation for the

normally distributed variables, or median (25%-75% interquartiles) for non-normally distributed variables. Categorical variables are expressed as ratios.

Table 2. Correlation matrix of MMP3, TIMP1, and vitamin D with all parameters.

	T D) (D) (D)	TD 4D4
Parameters	Vitamin D	MMP3	TIMP1
Sex	0.175	0.005	0.003
Age	0.091	-0.099	0.092
Smoking	-0.106	0.075	0.192
Duration of HD	-0.603**	0.165	0.134
BMI	0.216*	0.021	0.011
Creatinine	-0.518**	0.238^{*}	0.072
Urea	-0.482**	0.273**	0.148
Pi	-0.552**	0.324**	0.222^{*}
Uric acid	0.018	0.161	0.168
Vitamin D	1.000	-0.221*	-0.128
Albumin	-0.012	0.154	-0.129
Magnesium	-0.027	0.022	-0.123
Ionized Mg	-0.027	0.022	-0.123
Calcium	0.215*	-0.021	0.043
Ionized Ca	0.222^{*}	-0.051	0.082
Total Ca/Mg	0.104	-0.023	0.127
Ionized Ca/Mg	0.071	-0.029	0.131
CRP	-0.507**	0.425**	0.279**
MMP3	-0.221*	1.000	0.134
TIMP1	-0.128	0.134	1.000

^{*:} p<0.05, **: p<0.01, CRP: C-reactive protein, BMI: Body mass index, Pi: inorganic phosphate, eGFR: estimated glomerular filtration rate, MMP3: matrix metalloproteinase-3, TIMP1: tissue inhibitor of metalloproteinases-1.

Table 3. Results of neural networks (NN). NN#1 was made with HD+CRP vs. HD-CRP as output variables. NN#2 was made with HD vs. healthy controls

	Models	NN#1	NN#2
	Models	HD+CRP vs. HD-CRP	HD vs. Healthy controls
Input I over	Number of units	11 parameters	11 parameters
Input Layer	Rescaling method	Normalized	Normalized
	Number of hidden layers	2	2
Hidden layers	Number of units in hidden layer 1	2	4
Hidden layers	Number of units in hidden layer 2	2	3
	Activation Function	Hyperbolic tangent	Hyperbolic tangent
	Dependent variables	HD+CRP vs. HD-CRP	HD vs. Healthy controls
Output layer	Number of units	2	2
Output layer	Activation function	Identity	Identity
	Error function	Sum of squares	Sum of squares
	Sum of squares error term	5.530	3.594
Training	% incorrect or relative error	33.3%	6.5%
	Prediction (sens, spec)	56.3%-78.6%	95.0%-92.9%
	Sum of Squares error	4.985	1.459
Tastina	%incorrect or relative error	36.4%	5.6%
Testing	Prediction)sens -spec(46.2%-88.9%	100.0%-91.7%
	AUC ROC	76.7%-76.7%	96.2%-96.9%
	%incorrect or relative error	37.5%	0%
Holdout	Prediction)sens-spec) or correlation with predicted value	33.3%-80.0%	100.0-100.0%

AUC ROC: area under Receiver Operating curve; sen-spec: sensitivity – specificity.

РИСУНКИ

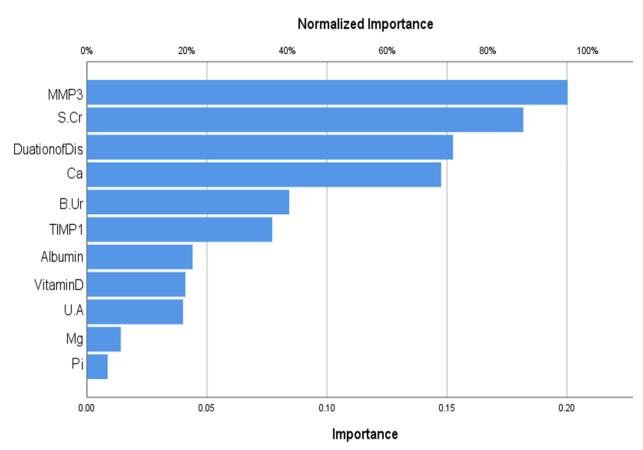


Figure 1. Results of neural network 1 (NN#1) (importance chart) with HD+CRP and HD-CRP as output variables and biomarkers as input variables.

B.ur: Blood urea, Ca: calcium, eGFR: estimated glomerular filtration rate, Mg: Magnesium, Pi: inorganic phosphate, MMP3: matrix metalloproteinase-3, S.Cr: serum creatinine, TIMP1: tissue inhibitor of metalloproteinases-1, U.A.: uric acid.

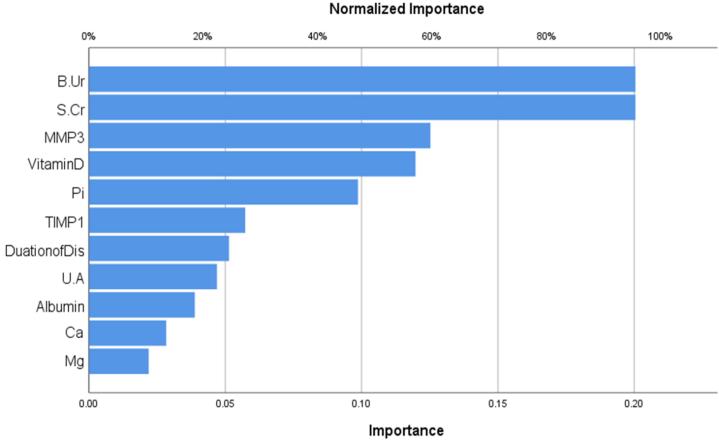


Figure 2. Results of neural network 2 (NN#2) (importance chart) with HD and healthy controls as output variables and biomarkers as input variables.

B.ur: Blood urea, Ca: calcium, eGFR: estimated glomerular filtration rate, Mg: Magnesium, Pi: inorganic phosphate, MMP3: matrix metalloproteinase-3, S.Cr: serum creatinine, TIMP1: tissue inhibitor of metalloproteinases-1, U.A.: uric acid.

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Полное название статьи:

PREDICTION OF INFLAMMATION IN HEMODIALYSIS PATIENTS USING NEURAL NETWORK ANALYSIS

НЕЙРОСЕТЕВОЙ АНАЛИЗ В ПРОГНОЗИРОВАНИИ ВОСПАЛЕНИЯ У ГЕМОДИАЛИЗНЫХ ПАЦИЕНТОВ

Сокращенное название статьи для верхнего колонтитула:

Inflammation in HD patients using NN

Воспаление у пациентов с ГД с использованием NN

Keywords: Hemodialysis patients (HD), Tissue inhibitor of metalloproteinases-1 (TIMP1), matrix metalloproteinase-1 (MMP3), vitamin D, neural network, inflammation.

Ключевые слова: пациенты, находящиеся на гемодиализе (ГД), тканевой ингибитор металлопротеиназы-1 (ТІМР1), матриксная металлопротеиназа-3 (MMP3), витамин D, нейронная сеть, воспаление.

Оригинальные статьи

Количество страниц текста -12, количество таблиц -3, количество рисунков -2.

08.08.2023.

СПИСОК ЛИТЕРАТУРЫ

Referen		
ce	Authors, title of a publication and source where it	
sequenc	was published, publisher's imprint	
e		
number		Reference's URL
1	Abdalsada, H. K., H. H. Hadi, A. F. Almulla, A. H.	https://doi.org/10.47836/pjst.31.4.27
	Najm, A. Al-Isa, and H. K. Al-Hakeim. Correlation of	
	Stromelysin-1 and Tissue Inhibitor of	
	Metalloproteinase-1 with Lipid Profile and Atherogenic	
	Indices in End-Stage Renal Disease Patients: A Neural	
	Network Study. Pertanika J. Sci. & Technol 2023,	
	August;31(4):2067-87	
2	Agarwal R. Defining end-stage renal disease in clinical	https://doi.org/10.1093/ndt/gfv289
	trials: a framework for adjudication. Nephrol Dial	
	Transplant. 2016 Jun;31(6):864-7. doi:	
	10.1093/ndt/gfv289. Epub 2015 Aug 11. PMID:	
	26264780.	
3	Amanzadeh M, Mota A, Zarghami N, Abedi-Azar S,	https://pubmed.ncbi.nlm.nih.gov/29421776/
	Abroon S, Akbarian N, Mihanfar A, Rahmati-Yamchi	
	M. Association Between Matrix Metalloproteinase-3	
	Activity and Glomerular Filtration Rate and	
	Albuminuria Status in Patients With Type 2 Diabetes	
	Mellitus. Iran J Kidney Dis. 2018 Jan;12(1):40-47.	
	PMID: 29421776.	

4	Avram MM, Mittman N, Myint MM, Fein P.	https://doi.org/10.1053/ajkd.2001.29254
	Importance of low serum intact parathyroid hormone as	
	a predictor of mortality in hemodialysis and peritoneal	
	dialysis patients: 14 years of prospective observation.	
	Am J Kidney Dis. 2001 Dec;38(6):1351-7. doi:	
	10.1053/ajkd.2001.29254. PMID: 11728974.	
5	Benjamini Y, Hochberg Y. Controlling the False	https://doi.org/10.1111/j.2517-
	Discovery Rate: A Practical and Powerful Approach to	6161.1995.tb02031.x
	Multiple Testing. J R Stat Soc Series B Stat Methodol.	
	1995;57(1):289-300. doi: 10.1111/j.2517-	
	6161.1995.tb02031.x.	
6	Block GA, Klassen PS, Lazarus JM, Ofsthun N, Lowrie	https://doi.org/10.1097/01.ASN.0000133041.276
	EG, Chertow GM. Mineral metabolism, mortality, and	<u>82.A2</u>
	morbidity in maintenance hemodialysis. J Am Soc	
	Nephrol. 2004 Aug;15(8):2208-18. doi:	
	10.1097/01.ASN.0000133041.27682.A2. PMID:	
	15284307.	
7	Bohle A, Wehrmann M, Bogenschütz O, Batz C, Müller	https://doi.org/10.1016/S0344-0338(11)80780-6
	CA, Müller GA. The pathogenesis of chronic renal	
	failure in diabetic nephropathy. Investigation of 488	
	cases of diabetic glomerulosclerosis. Pathol Res Pract.	
	1991 Mar;187(2-3):251-9. doi: 10.1016/s0344-	
	0338(11)80780-6. PMID: 2068008.	
8	Cantaluppi V, Quercia AD, Dellepiane S, Ferrario S,	https://doi.org/10.1093/ndt/gfu046
	Camussi G, Biancone L. Interaction between systemic	
	inflammation and renal tubular epithelial cells. Nephrol	

	Dial Transplant. 2014 Nov;29(11):2004-11. doi: 10.1093/ndt/gfu046. Epub 2014 Mar 2. PMID: 24589723.	
9	Carome MA, Striker LJ, Peten EP, Moore J, Yang CW, Stetler-Stevenson WG, et al. Human glomeruli express TIMP-1 mRNA and TIMP-2 protein and mRNA. Am J Physiol. 1993;264(6 Pt 2):F923-9. Epub 1993/06/01. doi: 10.1152/ajprenal.1993.264.6.F923. PubMed PMID: 8322893.	https://doi.org/10.1152/ajprenal.1993.264.6.F923
10	Chen D-Q, Cao G, Chen H, Liu D, Su W, Yu X-Y, et al. Gene and protein expressions and metabolomics exhibit activated redox signaling and wnt/β-catenin pathway are associated with metabolite dysfunction in patients with chronic kidney disease. Redox Biology. 2017;12:505-21. doi: https://doi.org/10.1016/j.redox.2017.03.017.	https://doi.org/10.1016/j.redox.2017.03.017
11	Clemmer JS, Shafi T, Obi Y. Physiological Mechanisms of Hypertension and Cardiovascular Disease in End-Stage Kidney Disease. Curr Hypertens Rep. 2022 Oct;24(10):413-424. doi: 10.1007/s11906-022-01203-7. Epub 2022 Jun 16. PMID: 35708820; PMCID: PMC10041674.	https://link.springer.com/article/10.1007/s11906- 022-01203-7
12	Crespo-Salgado J, Vehaskari VM, Stewart T, Ferris M, Zhang Q, Wang G, et al. Intestinal microbiota in pediatric patients with end stage renal disease: a Midwest Pediatric Nephrology Consortium study. Microbiome. 2016;4(1):50. Epub 2016/09/19. doi:	https://link.springer.com/article/10.1186/s40168- 016-0195-9

	10.1186/s40168-016-0195-9. PubMed PMID:	
	27640125; PubMed Central PMCID:	
	PMCPMC5027112.	
13	DeSoi CA, Umans JG. Phosphate kinetics during high-	https://doi.org/10.1681/ASN.V451214
	flux hemodialysis. J Am Soc Nephrol. 1993	
	Nov;4(5):1214-8. doi: 10.1681/ASN.V451214. PMID:	
	8305649.	
14	Eguchi T, Kubota S, Kawata K, Mukudai Y, Uehara J,	https://doi.org/10.1128/MCB.01288-07
	Ohgawara T, Ibaragi S, Sasaki A, Kuboki T, Takigawa	
	M. Novel transcription-factor-like function of human	
	matrix metalloproteinase 3 regulating the CTGF/CCN2	
	gene. Mol Cell Biol. 2008 Apr;28(7):2391-413. doi:	
	10.1128/MCB.01288-07. Epub 2008 Jan 2. PMID:	
	18172013; PMCID: PMC2268440.	
15	Gluba-Brzózka A, Michalska-Kasiczak M, Franczyk-	https://link.springer.com/article/10.1186/1476-
	Skóra B, Nocuń M, Banach M, Rysz J. Markers of	<u>511X-13-135</u>
	increased cardiovascular risk in patients with chronic	
	kidney disease. Lipids Health Dis. 2014 Aug 21;13:135.	
	doi: 10.1186/1476-511X-13-135. PMID: 25145866;	
	PMCID: PMC4246537.	
16	Guizani I, Zidi W, Zayani Y, Boudiche S, Hadj-Taieb S,	https://link.springer.com/article/10.1007/s11033-
	Sanhaji H, Zaroui A, Mechmeche R, Mourali MS, Feki	<u>019-04914-4</u>
	M, Allal-Elasmi M. Matrix metalloproteinase-3 predicts	
	clinical cardiovascular outcomes in patients with	
	coronary artery disease: a 5 years cohort study. Mol Biol	

	Rep. 2019 Oct;46(5):4699-4707. doi: 10.1007/s11033- 019-04914-4. Epub 2019 Jun 19. PMID: 31218540.	
17	Hadi T, Boytard L, Silvestro M, Alebrahim D, Jacob S, Feinstein J, et al. Macrophage-derived netrin-1 promotes abdominal aortic aneurysm formation by activating MMP3 in vascular smooth muscle cells. Nature communications. 2018;9(1):5022. Epub 2018/11/28. doi: 10.1038/s41467-018-07495-1. PubMed PMID: 30479344; PubMed Central PMCID: PMCPMC6258757.	https://www.nature.com/articles/s41467-018- 07495-1
18	Hendriks FK, Kooman JP, van Loon LJC. Dietary protein interventions to improve nutritional status in end-stage renal disease patients undergoing hemodialysis. Curr Opin Clin Nutr Metab Care. 2021 Jan;24(1):79-87. doi: 10.1097/MCO.000000000000000703. PMID: 33060457; PMCID: PMC7752218.	https://doi: 10.1097/MCO.000000000000000000000000000000000000
19	Housley TJ, Baumann AP, Braun ID, Davis G, Seperack PK, Wilhelm SM. Recombinant Chinese hamster ovary cell matrix metalloprotease-3 (MMP-3, stromelysin-1). Role of calcium in promatrix metalloprotease-3 (pro-MMP-3, prostromelysin-1) activation and thermostability of the low mass catalytic domain of MMP-3. The Journal of biological chemistry. 1993;268(6):4481-7. Epub 1993/02/25. PubMed PMID: 8440730.	https://doi.org/10.1016/S0021-9258(18)53634-6

20	Ishizaki M, Matsunaga T, Adachi K, Miyashita E. Serum matrix metalloproteinase-3 in hemodialysis patients with dialysis-related amyloidosis. Hemodial Int. 2004;8(3):219-25. Epub 2004/07/01. doi: 10.1111/j.1492-7535.2004.01099.x. PubMed PMID:	https://doi.org/10.1111/j.1492- 7535.2004.01099.x
	19379421.	
21	Kanda H, Hirasaki Y, Iida T, Kanao-Kanda M, Toyama Y, Chiba T, et al. Perioperative Management of Patients With End-Stage Renal Disease. Journal of Cardiothoracic and Vascular Anesthesia. 2017;31(6):2251-67. doi: https://doi.org/10.1053/j.jvca.2017.04.019.	https://doi.org/10.1053/j.jvca.2017.04.019
22	Khokha R, Murthy A, Weiss A. Metalloproteinases and their natural inhibitors in inflammation and immunity. Nat Rev Immunol. 2013 Sep;13(9):649-65. doi: 10.1038/nri3499. PMID: 23969736.	https://www.nature.com/articles/nri3499
23	Kobusiak-Prokopowicz M, Kaaz K, Marciniak D, Karolko B, Mysiak A. Relationships between Circulating Matrix Metalloproteinases, Tissue Inhibitor TIMP-2, and Renal Function in Patients with Myocarditis. Kidney Blood Press Res. 2021;46(6):749-757. doi: 10.1159/000519594. Epub 2021 Nov 19. PMID: 34801997.	https://doi.org/10.1159/000519594
24	Kostov K, Blazhev A. Changes in Serum Levels of Matrix Metalloproteinase-1 and Tissue Inhibitor of	https://doi.org/10.3390/bioengineering9030119

	Metalloproteinases-1 in Patients with Essential	
	Hypertension. Bioengineering (Basel). 2022 Mar	
	15;9(3):119. doi: 10.3390/bioengineering9030119.	
	PMID: 35324807; PMCID: PMC8945798.	
25	Lerner A, Neidhöfer S, Reuter S, Matthias T. MMP3 is	https://doi.org/10.1016/j.berh.2019.01.006
	a reliable marker for disease activity, radiological	
	monitoring, disease outcome predictability, and	
	therapeutic response in rheumatoid arthritis. Best Pract	
	Res Clin Rheumatol. 2018 Aug;32(4):550-562. doi:	
	10.1016/j.berh.2019.01.006. Epub 2019 Feb 14. PMID:	
	31174824.	
26	Levey AS, Coresh J, Greene T, Marsh J, Stevens LA,	https://cir.nii.ac.jp/crid/1570009750977916672
	Kusek JW, et al. Expressing the Modification of Diet in	
	Renal Disease Study equation for estimating glomerular	
	filtration rate with standardized serum creatinine values.	
	Clin Chem. 2007;53(4):766-72. Epub 2007/03/03. doi:	
	10.1373/clinchem.2006.077180. PubMed PMID:	
	17332152.	
27	Lim AI, Chan LY, Lai KN, Tang SC, Chow CW, Lam	https://doi.org/10.1016/j.biocel.2012.03.015
	MF, Leung JC. Distinct role of matrix	
	metalloproteinase-3 in kidney injury molecule-1	
	shedding by kidney proximal tubular epithelial cells. Int	
	J Biochem Cell Biol. 2012 Jun;44(6):1040-50. doi:	
	10.1016/j.biocel.2012.03.015. Epub 2012 Mar 30.	
	PMID: 22484054.	

20		1 //1 / // // // // 1 // // // // // //
28	Luczyszyn SM, de Souza CM, Braosi AP, Dirschnabel	https://doi.org/10.1016/j.archoralbio.2012.01.013
	AJ, Claudino M, Repeke CE, Faucz FR, Garlet GP,	
	Pecoits-Filho R, Trevilatto PC. Analysis of the	
	association of an MMP1 promoter polymorphism and	
	transcript levels with chronic periodontitis and end-stage	
	renal disease in a Brazilian population. Arch Oral Biol.	
	2012 Jul;57(7):954-63. doi:	
	10.1016/j.archoralbio.2012.01.013. Epub 2012 Mar 7.	
	PMID: 22401717.	
29	Lynch CC, Matrisian LM. Matrix metalloproteinases in	https://doi.org/10.1046/j.1432-
	tumor-host cell communication. Differentiation. 2002	0436.2002.700909.x
	Dec;70(9-10):561-73. doi: 10.1046/j.1432-	
	0436.2002.700909.x. PMID: 12492497.	
30	Matulka M, Konopka A, Mroczko B, Pryczynicz A,	https://doi.org/10.1155/2019/3136792
	Kemona A, Groblewska M, Sieskiewicz A, Olszewska	
	E. Expression and Concentration of Matrix	
	Metalloproteinase 9 and Tissue Inhibitor of Matrix	
	Metalloproteinases 1 in Laryngeal Squamous Cell	
	Carcinoma. Dis Markers. 2019 Apr 18;2019:3136792.	
	doi: 10.1155/2019/3136792. PMID: 31143300; PMCID:	
	PMC6501248.	
31	Melamed ML, Eustace JA, Plantinga L, Jaar BG, Fink	https://doi.org/10.1038/sj.ki.5001542
	NE, Coresh J, Klag MJ, Powe NR. Changes in serum	<u>-</u>
	calcium, phosphate, and PTH and the risk of death in	
	incident dialysis patients: a longitudinal study. Kidney	

	Int. 2006 Jul;70(2):351-7. doi: 10.1038/sj.ki.5001542.	
	Epub 2006 May 31. PMID: 16738536.	
32	Miljković M, Stefanović A, Bogavac-Stanojević N,	https://doi.org/10.20471/acc.2017.56.04.14
	Simić-Ogrizović S, Dumić J, Černe D, Jelić-Ivanović Z,	
	Kotur-Stevuljević J. Association of Pentraxin-3,	
	Galectin-3 and Matrix Metalloproteinase-9/Timp-1 with	
	Cardiovascular Risk in Renal Disease Patients. Acta	
	Clin Croat. 2017 Dec;56(4):673-680. doi:	
	10.20471/acc.2017.56.04.14. PMID: 29590722.	
33	Modi ZJ, Lu Y, Ji N, Kapke A, Selewski DT, Dietrich	https://doi:10.1001/jamacardio.2019.0375
	X, Abbott K, Nallamothu BK, Schaubel DE, Saran R,	
	Gipson DS. Risk of Cardiovascular Disease and	
	Mortality in Young Adults With End-stage Renal	
	Disease: An Analysis of the US Renal Data System.	
	JAMA Cardiol. 2019 Apr 1;4(4):353-362. doi:	
	10.1001/jamacardio.2019.0375. PMID: 30892557;	
	PMCID: PMC6484951.	
34	Mora-Gutierrez JM, Fernandez-Seara MA, Slon Roblero	https://doi.org/10.1093/ndt/gfy104.SP453
	MF, Gonzalez O, Escalada FJ, Soler MJ, et al. SP453	
	matrix metalloproteinase-10 and tissue inhibitor of	
	metalloproteinase-1 (timp-1) as early predictors of	
	nephropathy in patients with type 2 diabetes mellitus.	
	Nephrology Dialysis Transplantation.	
	2018;33(suppl_1):i500-i.	
35	Mora-Gutiérrez JM, Rodríguez JA, Fernández-Seara	https://www.nature.com/articles/s41598-019-
	MA, Orbe J, Escalada FJ, Soler MJ, Slon Roblero MF,	<u>56856-3</u>

	Riera M, Páramo JA, Garcia-Fernandez N. MMP-10 is	
	Increased in Early Stage Diabetic Kidney Disease and	
	can be Reduced by Renin-Angiotensin System	
	Blockade. Sci Rep. 2020 Jan 8;10(1):26. doi:	
	10.1038/s41598-019-56856-3. PMID: 31913319;	
	PMCID: PMC6949265.	
36	Nguyen-Khoa T, Massy ZA, De Bandt JP, Kebede M,	https://doi.org/10.1093/ndt/16.2.335
	Salama L, Lambrey G, et al. Oxidative stress and	
	haemodialysis: role of inflammation and duration of	
	dialysis treatment. Nephrol Dial Transplant.	
	2001;16(2):335-40. Epub 2001/02/07. doi:	
	10.1093/ndt/16.2.335. PubMed PMID: 11158409.	
37	Preston GA, Barrett CV, Alcorta DA, Hogan SL,	https://doi.org/10.1159/000065464
	Dinwiddie L, Jennette JC, et al. Serum matrix	
	metalloproteinases MMP-2 and MMP-3 levels in	
	dialysis patients vary independently of CRP and IL-6	
	levels. Nephron. 2002;92(4):817-23. Epub 2002/10/26.	
	doi: 10.1159/000065464. PubMed PMID: 12399626.	
38	Punsawad C, Viriyavejakul P. Increased expression of	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC
	kidney injury molecule-1 and matrix metalloproteinase-	<u>6965263/</u>
	3 in severe Plasmodium falciparum malaria with acute	
	kidney injury. Int J Clin Exp Pathol. 2017;10(7):7856-	
	64. Epub 2017/07/01. PubMed PMID: 31966633;	
	PubMed Central PMCID: PMCPMC6965263.	
39	Rashpa RS, Mahajan VK, Kumar P, Mehta KS,	https://doi.org/10.4103/idoj.IDOJ_160_17
	Chauhan PS, Rawat R, Sharma V. Mucocutaneous	

	Manifestations in Patients with Chronic Kidney Disease: A Cross-sectional Study. Indian Dermatol Online J. 2018 Jan-Feb;9(1):20-26. doi: 10.4103/idoj.IDOJ_160_17. PMID: 29441293; PMCID: PMC5803936.	
40	Ralay Ranaivo H, Hodge JN, Choi N, Wainwright MS. Albumin induces upregulation of matrix metalloproteinase-9 in astrocytes via MAPK and reactive oxygen species-dependent pathways. J Neuroinflammation. 2012 Apr 16;9:68. doi: 10.1186/1742-2094-9-68. PMID: 22507553; PMCID: PMC3419618.	https://doi.org/10.1186/1742-2094-9-68
41	Rymarz A, Mosakowska M, Niemczyk S. The significance of metalloproteinase 3 (MMP-3), chemokine CXC ligand 13 (CXCL-13) and complement component C5a in different stages of ANCA associated vasculitis. Scientific reports. 2021;11(1):5132. Epub 2021/03/06. doi: 10.1038/s41598-021-84662-3. PubMed PMID: 33664330; PubMed Central PMCID: PMCPMC7933137.	https://www.nature.com/articles/s41598-021-84662-3 https://doi.org/10.1038/s41598-021-84662-3
42	Rysz J, Banach M, Stolarek RA, Pasnik J, Cialkowska-Rysz A, Koktysz R, et al. Serum matrix metalloproteinases MMP-2 and MMP-9 and metalloproteinase tissue inhibitors TIMP-1 and TIMP-2 in diabetic nephropathy. J Nephrol. 2007;20(4):444-52. Epub 2007/09/20. PubMed PMID: 17879211.	https://pubmed.ncbi.nlm.nih.gov/17879211/

43	Sabbagh Y. Phosphate as a sensor and signaling	https://doi.org/10.5414/cn107322
	molecule. Clin Nephrol. 2013 Jan;79(1):57-65. doi:	
	10.5414/CN107322. PMID: 23006338.	
44	Sakaguchi Y, Hamano T, Isaka Y. Magnesium in	https://doi.org/10.1159/000485700
	Hemodialysis Patients: A New Understanding of the	
	Old Problem. Contributions to nephrology.	
	2018;196:58-63. Epub 2018/07/25. doi:	
	10.1159/000485700. PubMed PMID: 30041205.	
45	Saran R, Robinson B, Abbott KC, Agodoa LY, Albertus	https://doi.org/10.1053/j.ajkd.2016.12.004.
	P, Ayanian J, et al. US Renal Data System 2016 Annual	
	Data Report: Epidemiology of Kidney Disease in the	
	United States. Am J Kidney Dis. 2017 Mar;69(3 Suppl	
	1):A7-A8. doi: 10.1053/j.ajkd.2016.12.004. Erratum in:	
	Am J Kidney Dis. 2017 May;69(5):712. PMID:	
	28236831; PMCID: PMC6605045.	
46	Sarnak MJ. Cardiovascular complications in chronic	https://doi.org/10.1016/S0272-6386(03)00372-X
	kidney disease. Am J Kidney Dis. 2003 Jun;41(5	
	Suppl):11-7. doi: 10.1016/s0272-6386(03)00372-x.	
	PMID: 12776309.	
47	Shi S, Su M, Shen G, Hu Y, Yi F, Zeng Z, Zhu P, Yang	https://doi.org/10.1002/jmv.26235
	G, Zhou H, Li Q, Xie X. Matrix metalloproteinase 3 as a	
	valuable marker for patients with COVID-19. J Med	
	Virol. 2021 Jan;93(1):528-532. doi: 10.1002/jmv.26235.	
	Epub 2020 Jul 11. PMID: 32603484; PMCID:	
	PMC7362036.	

48	Siloşi I, Boldeanu MV, Mogoantă SŞ, Ghiluşi M,	https://pubmed.ncbi.nlm.nih.gov/25611261/
	Cojocaru M, Biciuşcă V, Cojocaru IM, Avrămescu CS,	
	Gheonea DI, Siloşi CA, Turculeanu A. Matrix	
	metalloproteinases (MMP-3 and MMP-9) implication in	
	the pathogenesis of inflammatory bowel disease (IBD).	
	Rom J Morphol Embryol. 2014;55(4):1317-24. PMID:	
	25611261.	
49	Tuncer T, Kaya A, Gulkesen A, Kal GA, Kaman D,	https://doi.org/10.17219/acem/94065
	Akgol G. Matrix metalloproteinase-3 levels in relation	
	to disease activity and radiological progression in	
	rheumatoid arthritis. Adv Clin Exp Med. 2019	
	May;28(5):665-670. doi: 10.17219/acem/94065. PMID:	
	30740946.	
50	Vaidya H, Giri S, Jain M, Goyal R. Decrease in serum	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC
	matrix metalloproteinase-9 and matrix	<u>3383361/</u>
	metalloproteinase-3 levels in Zucker fa/fa obese rats	
	after treatment with swertiamarin. Exp Clin Cardiol.	
	2012;17(1):12-6. Epub 2012/12/04. PubMed PMID:	
	23204894; PubMed Central PMCID:	
	PMCPMC3383361.	
51	Wan CY, Li L, Liu LS, Jiang CM, Zhang HZ, Wang JX.	https://doi.org/10.1016/j.joen.2021.04.005
	Expression of Matrix Metalloproteinases and Tissue	
	Inhibitor of Matrix Metalloproteinases during Apical	
	Periodontitis Development. J Endod. 2021	
	Jul;47(7):1118-1125. doi: 10.1016/j.joen.2021.04.005.	
	Epub 2021 Apr 23. PMID: 33895237.	

52	Wanchaitanawong W, Tantiworawit A, Piriyakhuntorn	https://doi.org/10.1080/16078454.2020.1856513
	P, Rattanathammethee T, Hantrakool S, Chai-	
	Adisaksopha C, Rattarittamrong E, Norasetthada L,	
	Niprapan P, Fanhchaksai K, Charoenkwan P. The	
	association between pre-transfusion hemoglobin levels	
	and thalassemia complications. Hematology. 2021	
	Dec;26(1):1-8. doi: 10.1080/16078454.2020.1856513.	
	PMID: 33357151.	
53	Warner RB, Najy AJ, Jung YS, Fridman R, Kim S, Kim	https://www.nature.com/articles/s41598-020-
	HC. Establishment of Structure-Function Relationship	<u>58964-x</u>
	of Tissue Inhibitor of Metalloproteinase-1 for Its	
	Interaction with CD63: Implication for Cancer Therapy.	
	Sci Rep. 2020 Feb 7;10(1):2099. doi: 10.1038/s41598-	
	020-58964-x. PMID: 32034211; PMCID:	
	PMC7005868.	
54	Warner RL, Bhagavathula N, Nerusu KC, Lateef H,	https://doi.org/10.1016/j.yexmp.2004.01.003
	Younkin E, Johnson KJ, Varani J. Matrix	
	metalloproteinases in acute inflammation: induction of	
	MMP-3 and MMP-9 in fibroblasts and epithelial cells	
	following exposure to pro-inflammatory mediators in	
	vitro. Exp Mol Pathol. 2004 Jun;76(3):189-95. doi:	
	10.1016/j.yexmp.2004.01.003. PMID: 15126100.	
55	Wu CF, Hou JS, Wang CH, Lin YL, Lai YH, Kuo CH,	https://doi.org/10.3390/ijerph17041230
	et al. Serum Sclerostin But Not DKK-1 Correlated with	
	Central Arterial Stiffness in End Stage Renal Disease	
	Patients. International journal of environmental research	

	1 11: 1 11 2020 17(1) F 1 2020 02 1 :	
	and public health. 2020;17(4). Epub 2020/02/23. doi:	
	10.3390/ijerph17041230. PubMed PMID: 32075016;	
	PubMed Central PMCID: PMCPMC7068530.	
56	Yang B, Vohra PK, Janardhanan R, Misra KD, Misra S.	https://doi.org/10.1016/j.jvir.2011.08.026
	Expression of profibrotic genes in a murine remnant	
	kidney model. Journal of vascular and interventional	
	radiology: JVIR. 2011;22(12):1765-72.e1. Epub	
	2011/10/28. doi: 10.1016/j.jvir.2011.08.026. PubMed	
	PMID: 22030458; PubMed Central PMCID:	
	PMCPMC3224153.	
57	Yatsyshyn R, Salyzhyn T. The Role of Tissue Inhibitor	https://www.thepharmajournal.com/archives/?yea
	of Matrix Metalloproteinase-1 in Cardiac and Blood	r=2016&vol=5&issue=10&ArticleId=885
	Vessels Remodeling and in Potential for Survival in	
	Case of Chronic Heart Failure of Various Origins.	
	Pharma Innov. 2016;5(10, Part B):85.	
58	Yazgan B, Avci F, Memi G, Tastekin E. Inflammatory	https://doi.org/10.1177/15353702211012417
	response and matrix metalloproteinases in chronic	
	kidney failure: Modulation by adropin and spexin. Exp	
	Biol Med (Maywood). 2021;246(17):1917-27. Epub	
	2021/05/25. doi: 10.1177/15353702211012417.	
	PubMed PMID: 34024143; PubMed Central PMCID:	
	PMCPMC8424640.	
59	Zhu QQ, Li TT, Chen R, Pan HF, Tao JH, Li XP, Ye	https://doi.org/10.3109/03009741003742789
	DQ. Elevated serum levels of MMP-2, MMP-3, and	
	MMP-13 in Chinese patients with systemic lupus	

erythematosus. Scand J Rheumatol. 2010;39(5):439-41.	
doi: 10.3109/03009741003742789. PMID: 20684736.	