

**HETEROGENEITY OF VIRULENCE FACTORS AMONG
PORPHYROMONAS GINGIVALIS CLINICAL ISOLATES FROM
PATIENTS WITH CHRONIC GENERALIZED PERIODONTITIS**

Koroleva I. V. ^{a, b},

Mikhailova E. S. ^a,

Privalova K. A. ^c,

Ermolaeva L. A. ^a,

Tumanova S. A. ^a,

Suvorov A. N. ^{a, b}

^a St. Petersburg State University, St. Petersburg, Russian Federation.

^b Institute of Experimental Medicine, St. Petersburg, Russian Federation.

^c Pavlov University, St. Petersburg, Russian Federation.

**ГЕТЕРОГЕННОСТЬ ФАКТОРОВ ВИРУЛЕНТНОСТИ СРЕДИ
КЛИНИЧЕСКИХ ИЗОЛЯТОВ *PORPHYROMONAS GINGIVALIS*,
ВЫДЕЛЕННЫХ ОТ ПАЦИЕНТОВ С ХРОНИЧЕСКИМ
ГЕНЕРАЛИЗОВАННЫМ ПАРОДОНТИТОМ**

Королева И. В. ^{1,2},

Михайлова Е. С. ²,

Привалова К. А. ³,

Ермолаева Л. А. ¹,

Туманова С. А. ¹,

Суворов А. Н. ^{1,2}

¹ ФГБОУ ВО Санкт-Петербургский государственный университет, Санкт-Петербург, Россия.

² ФГБНУ Институт экспериментальной медицины, Санкт-Петербург, Россия.

³ ФГБОУ ВО Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова МЗ РФ.

Abstract

The development of chronic generalized periodontitis occurs due to a combination of a several causes, among which the leading role is assigned to periodontal pathogens, which include *P. gingivalis*. Among *P. gingivalis* virulence factors, the polysaccharide capsule, fimbria proteins, cysteine proteases, and hemagglutinins are of special importance. The study was aimed to investigate the prevalence of specific virulence genes and identify a virulent genotype among *P. gingivalis* isolates found in patients with severe chronic generalized periodontitis (CGP). 41 patients (27 women and 14 men, average age 43.9 ± 1.5 years) were examined, of which main and control group consisted of 22 patients with severe CGP and 19 patients without inflammatory periodontal diseases, respectively. The PCR data allow to consider type II fimbria (FimA II), arginine-dependent type A protease (RghA) and lysine-dependent protease (Kgh) as specific markers for the detection of more virulent *P. gingivalis* strains. It was found that in St. Petersburg, the following *P. gingivalis* genotypes predominate among patients with severe CGP: *fimA II:kg:rghA*, *fimA II:kgh* and *fimA II:rghA*. In addition, it has been demonstrated that virulent genotypes are detected to a small extent in *P. gingivalis* isolates from healthy control group patients. The identification of *P. gingivalis* strains with a more prominent pathogenic potential and the detection of their virulent genotypes is of great practical importance, in the future allowing to develop advanced effective methods for disease prevention to be used in a personalized medicine strategy. The results obtained are also of high importance due to the recorded variability in the circulation of *P. gingivalis* strain genotypes in various worldwide regions.

Keywords: chronic generalized periodontitis, heterogeneity, virulent genotype, *P. gingivalis*, fimbriae, cysteine proteases.

Резюме

Развитие хронического генерализованного пародонтита обусловлено сочетанием целого ряда причин, среди которых ведущая роль отводится пародонтопатогенам, к которым относится *P. gingivalis*. Среди факторов вирулентности *P. gingivalis* особенно выделяют полисахаридную капсулу, белки фимбрий, цистеиновые протеазы, гемагглютинины. Цель исследования - изучение распространенности специфических генов вирулентности и определение вирулентного генотипа среди изолятов *P. gingivalis*, выявляемых у пациентов с хроническим генерализованным пародонтитом тяжелой степени (ХГП ТС). Было проведено обследование 41 пациента (27 женщин и 14 мужчин, средний возраст $43,9 \pm 1,5$ лет), из которых основную группу составили 22 пациента с ХГП ТС и контрольную группу составили 19 пациентов без воспалительных заболеваний пародонта. Результаты, полученные с использованием ПЦР, позволяют рассматривать фимбрии II типа (FimA II), аргинин-зависимую протеазу A типа (RghA) и лизин-зависимую протеазу (Kgh) в качестве специфических маркеров для обнаружения более патогенных штаммов *P. gingivalis*. Установлено, что в Санкт-Петербурге среди пациентов с ХГП ТС преобладают следующие генотипы *P. gingivalis*: *fimA II:kgh:rghA*, *fimA II:kgh* и *fimA II:rghA*. Кроме того, было продемонстрировано, что вирулентные генотипы выявляются в незначительной степени в изолятах *P. gingivalis* от здоровых пациентов контрольной группы. Идентификация штаммов *P. gingivalis* с более выраженным патогенным потенциалом и обнаружение их вирулентных генотипов имеет важное практическое значение, позволяя в будущем разработать современные эффективные методы профилактики заболевания и использовать их в стратегии персонализированной медицины. Полученные результаты также представляют высокую значимость в связи зарегистрированной вариабельностью циркуляции генотипов штаммов *P. gingivalis* в различных регионах мира.

Ключевые слова: хронический генерализованный пародонтит, гетерогенность, вирулентный генотип, *P. gingivalis*, фимбрии, цистеиновые протеазы.

1 Introduction

2 Damaging effect of dental plaque plays a leading role in the development of
3 inflammatory periodontal diseases. The today's science views dental plaque as a
4 biofilm consisting of a structured bacterial community and their metabolic products
5 [1, 17]. During the formation of dental plaque, the composition of microbiota takes
6 a trend to change from the dominance of aerobic and facultative anaerobic forms to
7 obligate anaerobic gram-negative rods and spiral-shaped bacteria [2, 15].

8 According to the "keystone pathogen hypothesis" theory *P. gingivalis* is
9 considered to be a key periodontal pathogen [9, 10]. Well described the ability of *P.*
10 *gingivalis* to have an impact on the innate immune system of the host organism
11 resulting in dysbiotic changes in the composition of microbiota and, therefore, in an
12 inflammatory response of periodontal tissues [11, 20]. Thus, *P. gingivalis* is
13 rightfully considered a "keystone pathogen" agent and, even at low concentrations,
14 it is capable of causing chronic periodontitis [9, 10, 12, 21]. Since *P. gingivalis*
15 belongs to obligate anaerobes, the main habitat of this periodontal pathogen is the
16 periodontal pocket. *P. gingivalis* is a secondary plaque colonizer and often forms
17 colonies with *S. gordonii* and *P. intermedia* [12]. In addition, as Socransky
18 "ecological theory" states, *P. gingivalis* forms a complex with *T. denticola* and *T.*
19 *forsythia* [8]. This combination of microorganisms is detected in the most severe
20 stages of chronic periodontitis. This "red complex" aggressively affects bone tissue
21 and gum mucosa, forming deep periodontal pockets and causing severe destruction
22 of jaw bone tissue [24, 28].

23 In recent years, it has become obvious that in the case of inflammatory
24 periodontal diseases, there are factors that enhance some mechanisms of disease
25 development making its course more severe. Differences in the course and disease
26 development rate are due to the diversity of the composition and degree of
27 periodontopathogen strains virulence [5, 23]. Scientists' views
28 about the influence of genetic diversity of *P. gingivalis* strains on the development
29 and course of inflammatory periodontal diseases are contradictory. It is believed that

30 *P. gingivalis* strains differ in the degree of virulence in patients with different
31 periodontitis currents, affecting the clinical course of the disease. Experimental and
32 clinical studies have revealed differences in the pathogenic potential of *P. gingivalis*
33 strains, dividing them into “invasive” and “non-invasive” strains [7, 21, 22]. On the
34 other hand, there is an idea that the course of periodontitis is determined not by the
35 diversity of strains of periodontal pathogens, but by the individual characteristics of
36 the host organism reactivity [7]. Moreover, a number of studies have shown the
37 availability of identical strains of *P. gingivalis* in patients with intact periodontium
38 and with inflammatory periodontal diseases [26].

39 *P. gingivalis* is known to express many virulence factors: fimbriae,
40 lipopolysaccharides, arginine- and lysine-dependent proteases, hemagglutinins and
41 capsular polysaccharide, which genes are very diverse [3, 16, 27]. Fimbriae of *P.*
42 *gingivalis* are classified into six genotypes - I, Ib, II, III, IV, V, that was
43 demonstrated by a variation in the nucleotide sequence of the *fimA* gene encoding
44 FimA (fimbryonic subunits) [6]. *P. gingivalis* with type I fimbriae has been shown
45 to be associated with patients without inflammatory periodontal diseases [4]. On the
46 other hand, there is an evidence that fimbriae of *P. gingivalis* with genotype II have
47 greater adhesive ability and, therefore, greater virulence compared to other
48 genotypes [6]. Pathogenic heterogeneity of *P. gingivalis* strains with type II fimbriae
49 has been demonstrated, that determines the proteolytic and invasive activity of this
50 periodontopathogen [14]. At the same time, it is believed that there are no significant
51 differences between *P. gingivalis* strains with different genotypes of fimbriae
52 regarding virulence [26].

53 The issue of genetic heterogeneity of *P. gingivalis* and its virulence factors is
54 being actively studied, and the scientific views about the role of those factors in the
55 formation of a virulent genotype are contradictory. Therefore, the further need to
56 examine the genetic heterogeneity of *P. gingivalis* in patients with inflammatory
57 periodontal diseases makes this research relevant. To study genetic heterogeneity of
58 fimbriae and cysteine proteases of *P. gingivalis* strains in patients with chronic

59 generalized periodontitis, as well as deriving characteristics of the virulent genotype
60 of *P. gingivalis* is very perspective. In addition, it is of interest to examine the
61 prevalence of the virulent genotype among *P. gingivalis* strains isolated from
62 patients without inflammatory periodontal diseases.

63 The purpose of the research: to study the prevalence of specific virulence
64 genes and identify the virulent genotype among *P. gingivalis* strains detected in
65 patients with severe chronic generalized periodontitis.

66 **2 Materials and methods**

67 A research was conducted on 41 patients (27 women and 14 men) aged 36
68 to 50 years (average age was 43.9 ± 1.5 years). The patients were divided into two
69 groups. The first group included 22 patients with severe chronic generalized
70 periodontitis (CGPS). The control group consisted of 19 patients without
71 inflammatory periodontal diseases (IPD). To form a control group, 69 patients
72 without IPD were preliminarily examined and those patients in whom *P. gingivalis*
73 was identified were selected.

74 In the course of research, a clinical examination of patients was performed,
75 including collection of anamnesis, complaints, assessment of dental status, index
76 assessment of the periodontal tissue condition (OHI-S index, Green, Vermillion,
77 1964; Silness-Loe index, Silness, Loe, 1964; CPITN index, WHO, 1978, Ainamo
78 et al., 1982; PMA index, Parma S., 1960; BOP index, Ainamo, Bau, 1975), as well
79 as microbiological examination of material from the periodontal sulcus or
80 periodontal pockets of the examined patients using PCR diagnostics. X-ray
81 examination included assessment of data obtained from GALILEOS cone-beam
82 computed tomography (Sirona, Germany).

83 Material was collected from periodontal pockets in patients with CGPS (main
84 group) and from the periodontal sulcus in healthy patients (control group) using
85 sterile paper absorbents Absorbent Paper Points from Euronda, size No. 25. Paper
86 absorbents were placed in the gingival sulcus or periodontal pockets for 7-10
87 seconds, after which they were immediately transferred into sterile sealed Eppendorf

88 tubes and stored at -50°C . A special cooling device was used to transport the material
89 to maintain storage conditions.

90 To isolate DNA, we used the Express-DNA-Bio kit in accordance with the
91 instructions. The resulting DNA samples were stored at -20°C until polymerase
92 chain reaction (PCR) was performed.

93 Primer 3 and OLIGO 4.0 computer programs was used to perform the design,
94 analysis of oligonucleotide primers and identification of the primer melting
95 temperature (Table 1). Primers to identify *fimA* I and IV genes were used from the
96 publication of Takashi Yoshino et al. [25].

97 The data obtained in the course of the research were processed on a personal
98 computer using the software system “Statistica for Windows” (v. 7.0). To visualize
99 the results of the study, diagrams were constructed in Microsoft Excel.

100 3 Results

101 All patients of the main group complained of bleeding gums during brushing,
102 tooth mobility, swelling and inflammation of the gums (Fig. 1). In 59.1% cases,
103 patients in the main group complained of food getting between the teeth, as well as
104 halitosis. Patients with CGPS in 72.73% cases marked the displacement of teeth
105 resulting from a functional secondary traumatic occlusion and in 68.18% cases -
106 itching and gum area burning caused by the inflammatory edema of periodontal
107 tissue. 2 patients in the control group had complaints only about food getting
108 between the teeth owing to lack of a contact point in the area of several tooth pairs
109 in the maxilla and mandible.

110 Hyperemia of the marginal and attached gums, and exudation from
111 periodontal pockets has been revealed in all patients of the main group. Gingival
112 recession has been found in all patients with CGPS, the average value amounted to
113 1.96 ± 0.07 mm. Lesions of the furcation in the area of the molars of the maxilla and
114 mandible were detected in 15 patients (68.2% of cases). Tooth mobility has been
115 revealed in all patients of the main group, most often grade 1-2.

116 The values of the OHI-S hygiene index in patients with CGPS in the main
117 group were 4.77 ± 0.12 , in the control group – 0.44 ± 0.04 (Fig. 2).

118 The obtained values of the OHI-S index indicate poor oral hygiene in patients
119 of the main group that is typical for patients with CGPS who lack the correct skills
120 to practice individual oral hygiene. Statistically significant differences have been
121 found in the Silness-Loe index between patients of the main and control groups
122 indicating a cause-and-effect correlation between the mineralized supra- and
123 subgingival dental plaque and inflammation in the periodontal tissues ($p < 0.001$).

124 The PMA index and BOP bleeding index rates in patients of the main group
125 indicate inflammatory phenomena in periodontal tissues. A distinct inflammatory-
126 destructive process in periodontal tissues in patients of the main group was
127 confirmed by the rates of the periodontal index. The rates of the CPITN index in
128 patients with CGPS of the main group reached 3.97 ± 0.23 . High rates of the CPITN
129 index for patients in the main group indicate the need for complex treatment,
130 including surgical treatment of periodontal diseases.

131 The results obtained during cone-beam computed tomography in patients of
132 the main group correspond to the clinical picture and the diagnosis of CGPS. In all
133 patients of the main group, destruction of the compact lamina of the alveolar bone
134 was revealed along its entire length, and bone pockets were identified in the area of
135 15.2 ± 3.7 teeth. The amount of bone tissue destruction “more than $\frac{1}{2}$ the length of
136 the root” was identified in all patients with CGPS.

137 The contents of the periodontal pockets in patients of the main group and the
138 subgingival sulcus in patients in the control group have been examined to reveal *P.*
139 *gingivalis* using PCR screening (Fig. 3).

140 In the main group, the periodontopathogen *P. gingivalis* has been detected
141 in all patients (100% cases). To form a control group, 69 patients without
142 inflammatory periodontal diseases (inflammatory periodontal diseases) were
143 previously examined, among whom *P. gingivalis* was detected in 21 patients (30.4%
144 of cases).

145 22 *P. gingivalis* isolates of the main group and 19 isolates of the control
146 group were examined by PCR to reveal genes encoding fimbriae types I, II, and IV
147 (Fig. 4).

148 The results indicate the predominance of the *fimA* II genotype (40.9% of
149 cases) among the *P. gingivalis* isolates of the main group. The *fimA* I genotype is
150 detected quite often as well, in 22.7% cases, while the prevalence of the *fimA* IV
151 genotype was insignificant (9.1% of cases). Thus, among *P. gingivalis* isolates from
152 patients with CGPS, strains carrying fimbriae of types I and II predominantly
153 circulate. Among the 19 *P. gingivalis* isolates of the control group, *fimA* II also
154 dominates (31.6% cases), the prevalence of which is only by 22.7% lower compared
155 to the prevalence rate of *fimA* II in the main group. The *fimA* I genotype in *P.*
156 *gingivalis* isolates of the control group is found quite rarely (5.3% of cases), and the
157 *fimA* IV genotype is not available at all.

158 The gene (*kgh*), encoding a lysine-dependent protease, in *P. gingivalis*
159 isolates of the main group is detected in 68.2% cases, while the gene (*rghA*),
160 encoding an arginine-dependent protease type A, is detected in *P. gingivalis* isolates
161 of all patients (100% cases) (Fig. 5).

162 When comparing the corresponding genes in control group *P. gingivalis*
163 isolates, a significant reduction in the prevalence of *kgh* and *rghA* was revealed, to
164 26.3% and 47.4% cases, respectively.

165 Table 2 shows the prevalence of various genotypes of the studied virulence
166 factors among *P. gingivalis* isolates of the main and control groups.

167 When considering three virulence factors, *P. gingivalis* isolates with genotype
168 *fimA* II:*kgh*:*rghA* (27.3% cases) and *fimA* I:*kgh*:*rghA* (18.2% cases) dominate the
169 main group. Both of those genotypes are present in almost half (45.5% cases) of *P.*
170 *gingivalis* isolates of the main group that suggests a significant contribution of *P.*
171 *gingivalis* with those genotypes in the development of chronic generalized
172 periodontitis. In *P. gingivalis* isolates of the control group, genotypes *fimA*
173 II:*kgh*:*rghA* and *fimA* I:*kgh*:*rghA* are found quite rare, in 5.3% cases. When

174 considering different genotypes from two virulence factors in *P. gingivalis* isolates
175 of the control group, genotypes *fimA* II:*rghA* (40.1% cases), *fimA* II:*kgh* (27.3%
176 cases) and *fimA* I:*rghA* (22.7% cases) predominate. In *P. gingivalis* isolates from the
177 control group, the *fimA* II:*rghA* genotype also predominate (18.2% cases). The
178 remaining genotypes listed above are found rare in *P. gingivalis* isolates from the
179 control group, in 5.3% cases.

180 4 Discussion

181 Among the virulence factors of *P. gingivalis*, the polysaccharide capsule, fimbriae
182 proteins, arginine- and lysine-dependent proteases, and hemagglutinins are
183 particularly distinguished. In the work in question, the heterogeneity of genes
184 encoding fimbriae types I, II and IV and arginine- and lysine-dependent proteases
185 was studied in clinical isolates of *P. gingivalis* obtained from healthy patients and
186 patients with CGPS. It has previously been demonstrated that *P. gingivalis* is
187 isolated from healthy patients in 10–25% cases [13]. A higher percentage of *P.*
188 *gingivalis* detection was reported in the dental plaque of healthy Japanese patients,
189 up to 36.8% cases [4]. A study of samples obtained from the subgingival sulcus of
190 69 healthy patients in St. Petersburg demonstrates a high percentage of carriage of
191 *P. gingivalis*, up to 30.4% cases. The prevalence of *P. gingivalis* in patients of the
192 main group amounted to 100% cases that fully correlates with previous studies [13,
193 18]. Among *P. gingivalis* fimbriae, six genotypes of *fimA* have been described, of
194 which genotypes II and IV have been characterized as the most common ones among
195 *P. gingivalis* isolates received from patients with IPD (inflammatory periodontal
196 disease) in Sweden and Japan [4, 25]. The remaining genotypes of fimbriae were
197 detected quite rarely [25]. In agreement with previous studies, among the three types
198 of arginine-dependent proteases, type A arginine-dependent protease (RghA) was
199 found to be dominant. Among lysine-dependent Kgh proteases, both genotypes I and
200 II occurred with approximately the same frequency [25].

201 When studying the prevalence of virulence factors in the main group, it has
202 been demonstrated that genes encoding type II fimbriae (40.9% cases) and type A

203 arginine-dependent protease (100% cases) were present in the first place by
204 frequency of occurrence. The obtained data suggest a direct correlation between
205 CGPS and *P. gingivalis* isolates carrying the *rghA* gene ($r=1.0$). The lysine-
206 dependent protease gene *kgh* is detected in 68.2% of cases. In *P. gingivalis* isolates
207 from healthy patients, type II fimbriae and type A arginine-dependent protease were
208 present in 31.6% and 47.4% of cases, respectively. Lysine-dependent protease Kgh
209 is detected in 26.3% of cases. The presented results suggest that *P. gingivalis* strains
210 producing fimbriae FimA II, as well as Kgh and RghA, have a more pronounced
211 pathogenic potential. Thus, one can consider those virulence factors as markers for
212 detecting more pathogenic strains of *P. gingivalis*.

213 Comparing data on the prevalence of *P. gingivalis* virulence factors in the
214 St. Petersburg region with previously published data for Sweden, *it's safe to say* that
215 type II fimbriae are most often found in *P. gingivalis* isolates from patients with IPD.
216 However, it is worth noting that *P. gingivalis* isolates with type II fimbriae are
217 recorded more often in Swedish patients (71% cases) compared to patients from St.
218 Petersburg (40.9% cases). Differences in *P. gingivalis* isolates from St. Petersburg
219 are also revealed when identifying type I and IV fimbriae. In *P. gingivalis* isolates
220 from Swedish patients, the second most common finding was *P. gingivalis* with type
221 IV fimbriae (16.1% cases). In the St. Petersburg region, *P. gingivalis* with type IV
222 fimbriae from patients with IPD are rarely found (9.1% cases); moreover, *P.*
223 *gingivalis* isolates with type IV fimbriae from healthy patients are not recorded at
224 all. On the other hand, in St. Petersburg, isolates of *P. gingivalis* with type I fimbriae
225 are more often detected from patients with IPD, in 22.7% cases. The relatively
226 frequent detection of *P. gingivalis* isolates with type I fimbriae (19.6% cases) from
227 Brazilian patients with IPD has been previously published [19]. In comparison, type
228 I fimbriae was rarely detected in *P. gingivalis* isolates from Swedish patients (4.8%
229 of cases). In addition, all *P. gingivalis* isolates from St. Petersburg patients with IPD
230 carry the gene for arginine-dependent protease type A, while in *P. gingivalis* isolates
231 from Swedish patients with IPD the gene for the mentioned protease is found less

232 frequently, in 75.8% cases . In summary, a study of clinical isolates of *P. gingivalis*
233 from patients with IPD demonstrates significant regional differences in the
234 prevalence of individual *P. gingivalis* virulence factors. The findings highlight the
235 significance of such studies for specific regions.

236 When analyzing genotypic variants for the studied virulence factors of *P.*
237 *gingivalis* in the main group, *fimA II:kgh:rghA* (27.3% cases), *fimA II:kgh* (27.3%)
238 and *fimA II:rghA* (40.1 % cases) are most common. Those results allow taking into
239 consideration the above-mentioned gene combinations as virulent genotypes of *P.*
240 *gingivalis*. It is important to mention that virulent genotypes were found in *P.*
241 *gingivalis* isolates from healthy control patients (*fimA II:kgh:rghA* and *fimA II:kgh* -
242 in 5.3% cases; *fimA II:rghA* - in 18.2% cases).

243 Identification of *P. gingivalis* strains with more distinct pathogenic potential
244 and detection of their virulent genotypes is of great practical importance. Using
245 obtained clinical and microbiological data, an in-depth research of the etiology of
246 IPD, development and implementation of up-to-date efficient methods to prevent the
247 disease are possible. In addition, the detection of virulent genotype strains among
248 control group patients allows predicting that some healthy patients have an increased
249 susceptibility to developing periodontitis. Keeping that knowledge in mind the
250 dentist will be able to prevent *inflammatory periodontal diseases* in advance,
251 develop individual oral hygiene for the patient and give appropriate
252 recommendations.

253 **Gratitude**

254 The authors express their gratitude to the staff of the Federal State Budgetary
255 Institution "Institute of Experimental Medicine" for the opportunity to perform this
256 scientific research.

ТАБЛИЦЫ

Table 1. Oligonucleotide primers.

	Name	5'→3'	T ⁰ _{ann.}	DNA fragments size (bp)
1	Gin1	GTATATGCTCGACGAGGTGGAA	57,0	334
2	Gin2	ATTGTCCAGGGTAACTTCTTCG		
3	Fim III	TGTTGCAGACAATAATCCTAC	51,0	250
4	Fim II2	CGATTACCAAGTAGCATTCTGA		
5	Kgp1	TCCAATTCTGACCACATCTCAA	56,0	397
6	Kgp2	AGCTTCCCGATAGTAATGAGCA		
7	RgpA1	AATCCCGGAACAACAACACTTT	56,0	331
8	RgpA2	TGAAGTTGGATGCATCGTTACC		

Table 2. Distribution of fimbria types I, II and IV (*fimA* I, II, IV), lysine-dependent protease (*kgh*) and arginine-dependent protease type A (*rghA*) genotypes.

Genotypes	Prevalence (%)	
	Main group	Control group
<i>fimA I:kgh:rghA</i>	18,2	5,3
<i>fimA II:kgh:rghA</i>	27,3	5,3
<i>fimA IV:kgh:rghA</i>	9,1	0
<i>fimA I:kgh</i>	18,2	5,3
<i>fimA I:rghA</i>	22,7	5,3
<i>fimA II:kgh</i>	27,3	5,3
<i>fimA II: rghA</i>	40,1	18,2
<i>fimA IV:kgh</i>	9,1	0
<i>fimA IV:rghA</i>	9,1	0

РИСУНКИ

Figure 1. Complaints from patients in the main and control groups.

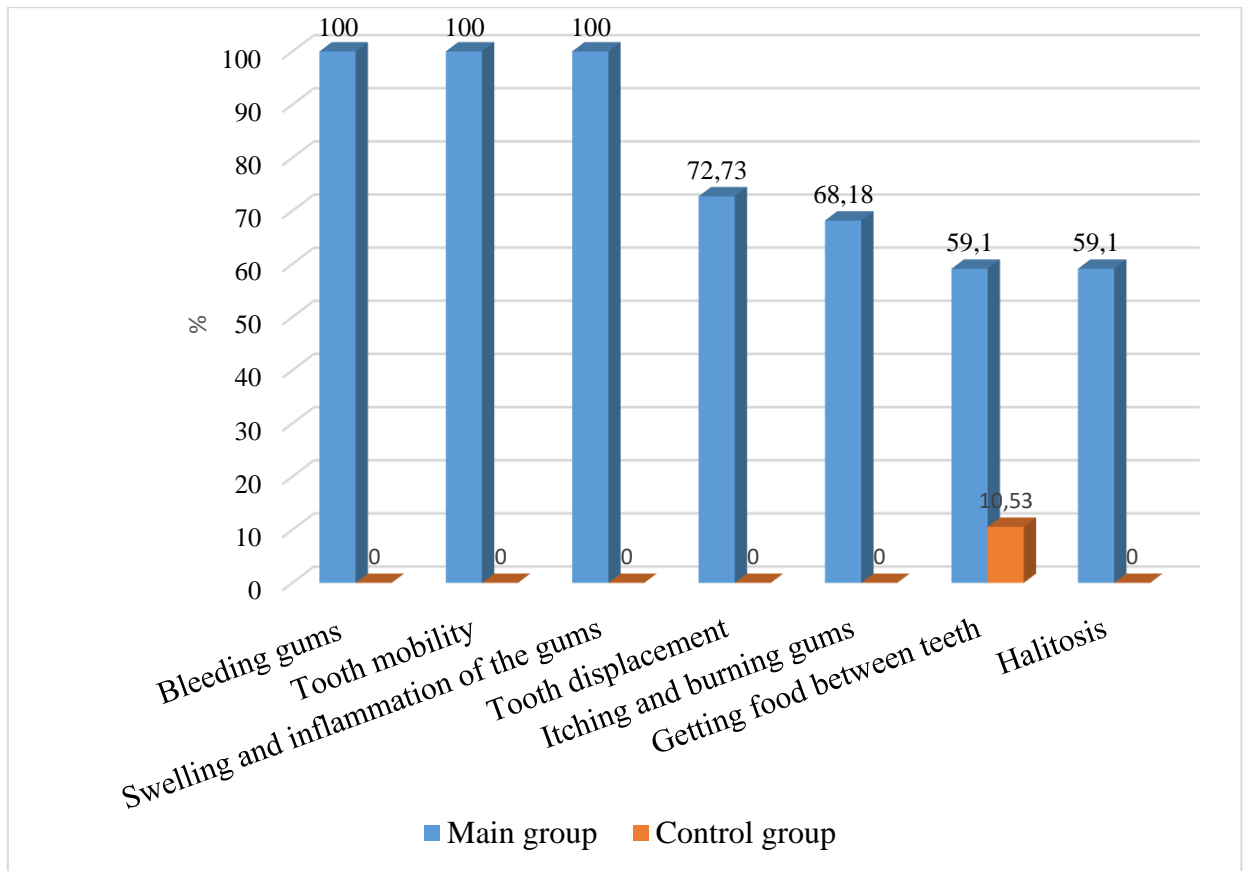


Figure 2. OHI-S, CPITN, Silness-Loe, PMA (%) и BOP (%) index rates in patients of the main and control groups.

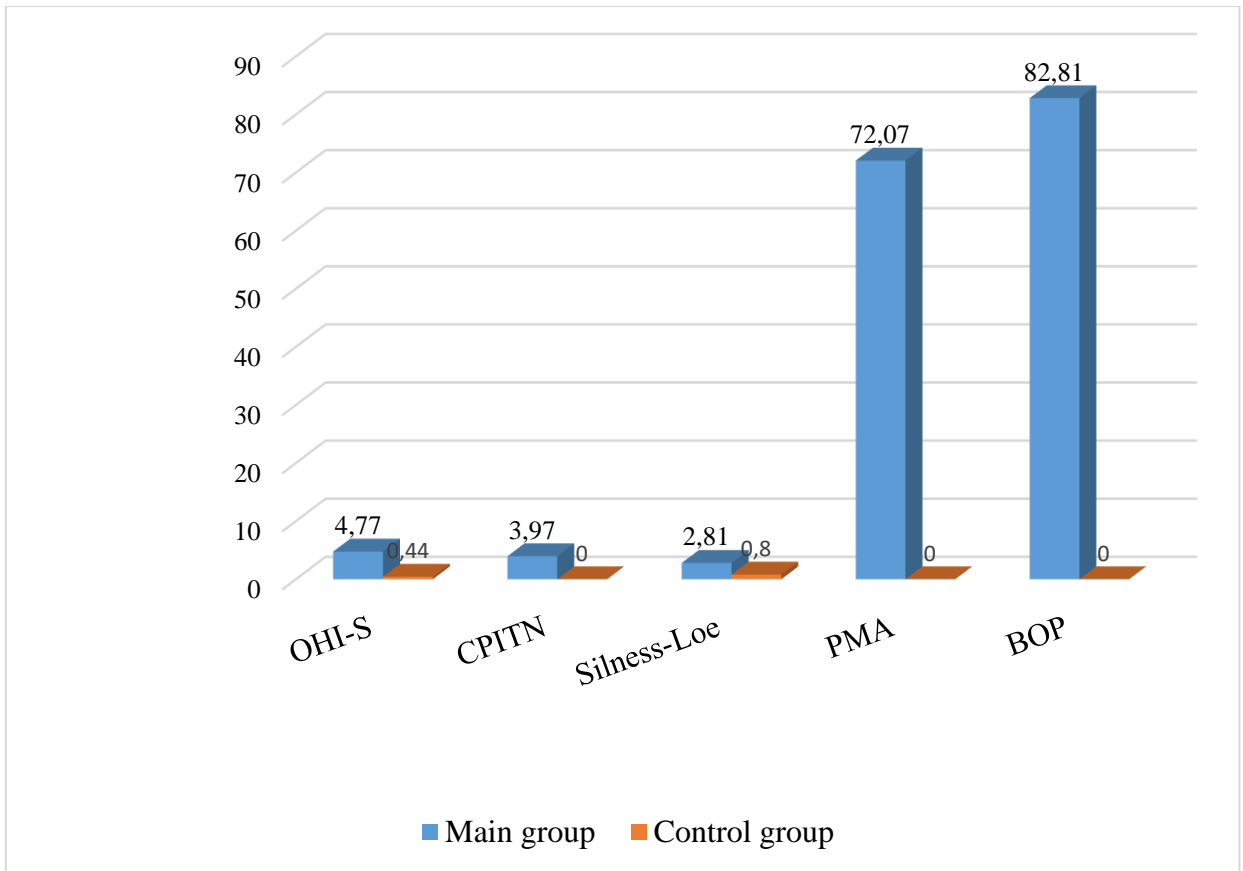


Figure 3. Frequency of *P. gingivalis* occurrence in periodontal pockets/gingival sulcus in patients of the main and control groups.

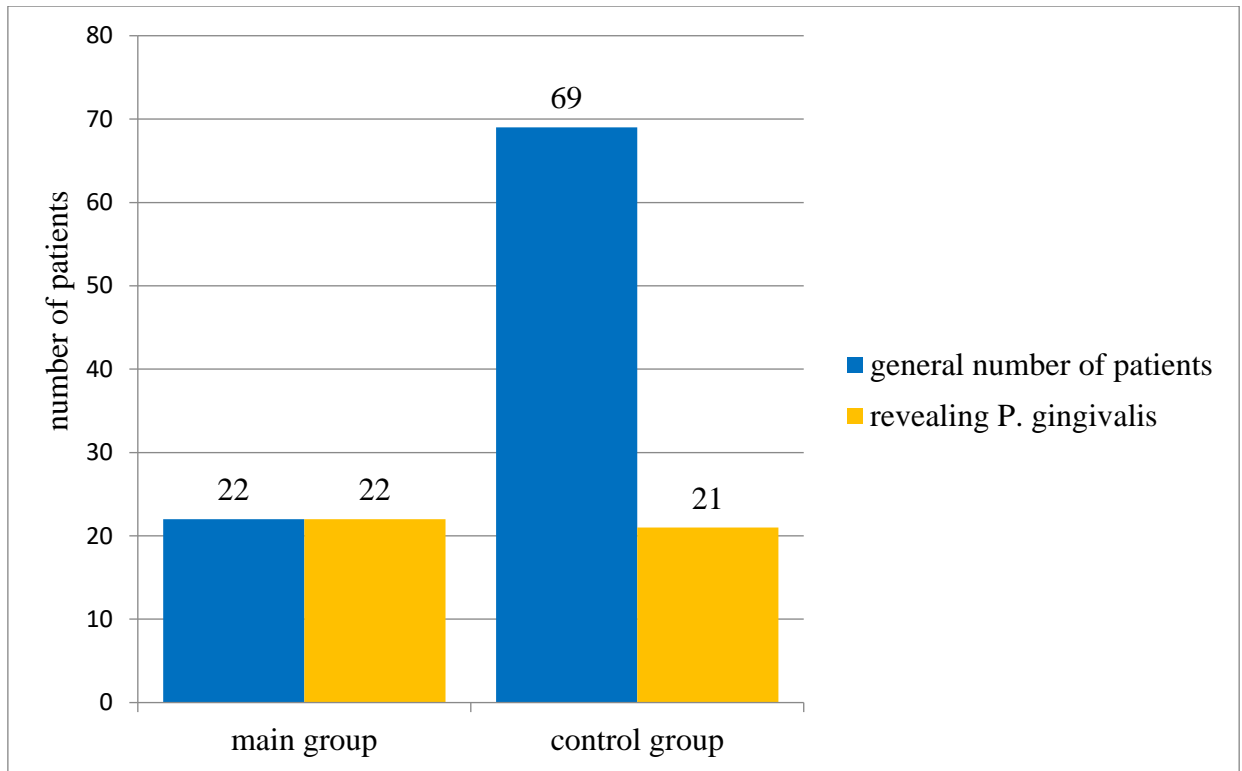


Figure 4. Prevalence of fimbriae types I, II and IV genes (fimA I, fimA II and fimA IV) among P. gingivalis isolates in patients of the main and control groups.

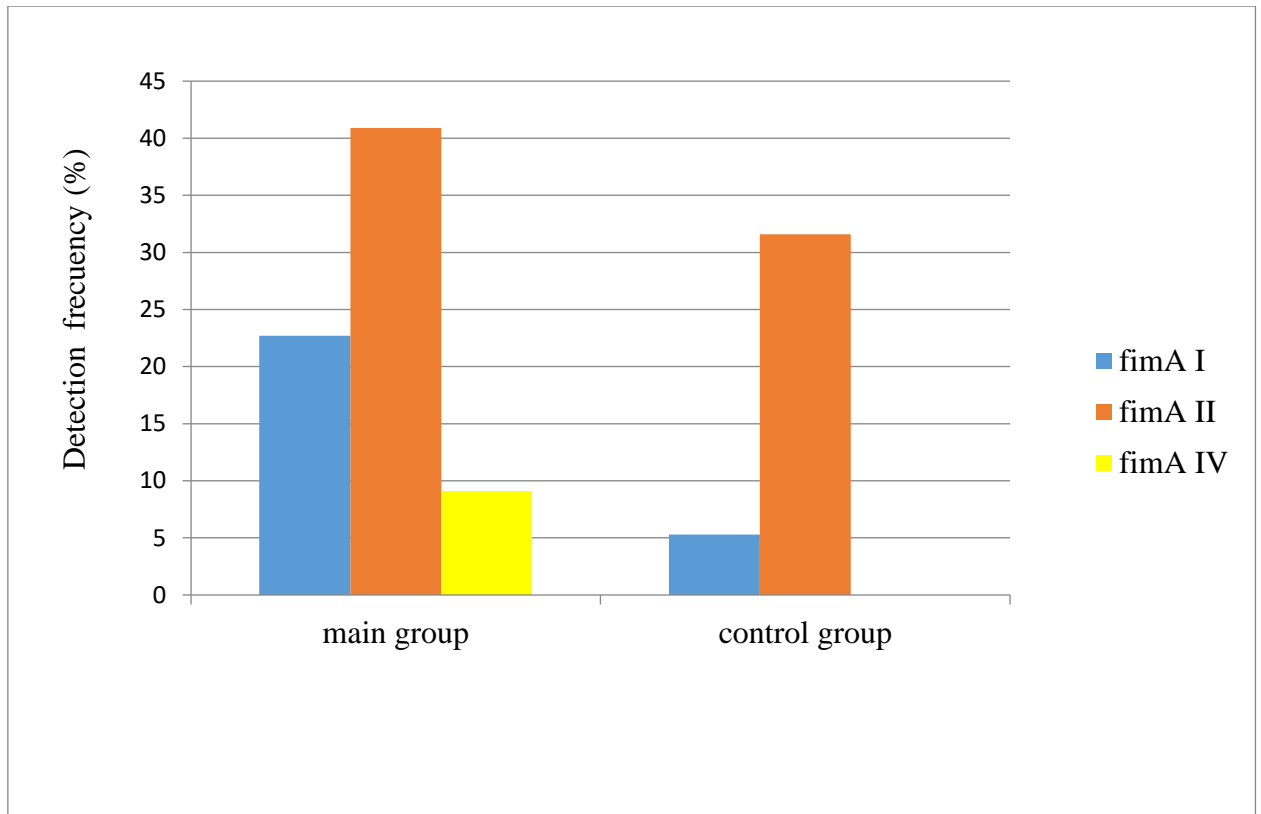
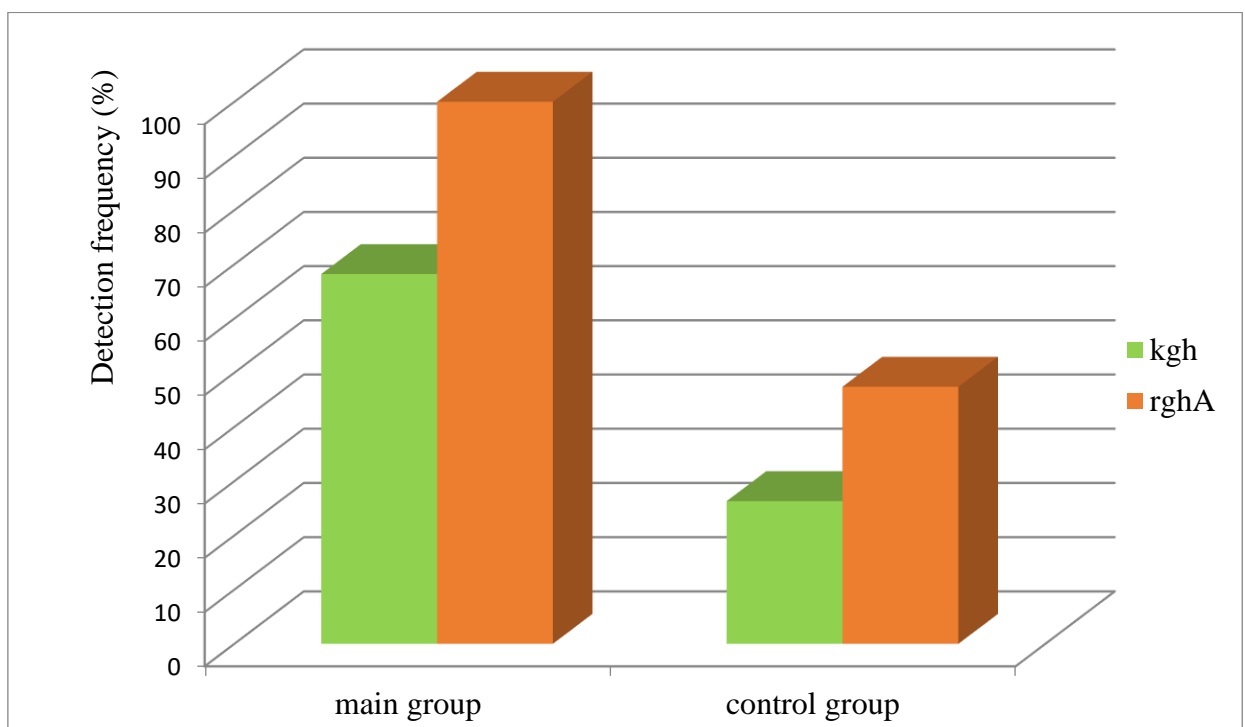


Figure 5. Prevalence of fimbriae types I, II and IV genes (fimA I, fimA II and fimA IV) among P. gingivalis isolates in patients of the main and control groups.



ТИТУЛЬНЫЙ ЛИСТ_МЕТАДААННЫЕ

Блок 1. Информация об авторе ответственном за переписку

Михайлова Е. С., д.м.н., доцент, доцент Кафедры терапевтической стоматологии Медицинского института ФГБОУ ВО «Санкт-Петербургский государственный университет», Санкт-Петербург, Россия;

адрес: 199034, Россия, Санкт-Петербург, Университетская набережная, д. 7-9, Санкт-Петербургский государственный университет;

телефон: 8(812)326-03-26;

e-mail: e.michailova@spbu.ru

Mikhailova E. S., DMS, associate Professor, Department of Therapeutic Dentistry, St. Petersburg State University, St. Petersburg, Russian Federation;

address: 197376, Russian Federation, St. Petersburg, *Universitetskaya embankment*, 7/9, St. Petersburg State University;

telephone: 8(812)326-03-26;

e-mail: e.michailova@spbu.ru

Блок 2. Информация об авторах

Koroleva I.V., PhD, Associate Professor, Department of Fundamental Problems of Medicine and Medical Technologies, St. Petersburg State University, St. Petersburg, Russian Federation;

Королева И.В., к.б.н., доцент Кафедры фундаментальных проблем медицины и медицинских технологий Медицинского института ФГБОУ ВО СПбГУ, Санкт-Петербург, Россия; старший научный сотрудник Отдела молекулярной микробиологии ФГБНУ «Институт экспериментальной медицины», Санкт-Петербург, Россия;

Senior Researcher, Department of Molecular Microbiology, Institute of Experimental Medicine, St. Petersburg, Russian Federation;

Privalova K.A., resident of Saint Petersburg State Medical University, St. Petersburg, Russian Federation;

Привалова К.А., ординатор Кафедры челюстно-лицевой хирургии и стоматологии хирургической ФГБОУ ВО «Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова» МЗ РФ, Санкт-Петербург, Россия;

Ermolaeva L.A., DMS, Professor, Head of the Therapeutic Dentistry Department, St. Petersburg State University, St. Petersburg, Russian Federation;

Ермолаева Л.А., д.м.н., профессор, зав. Кафедрой терапевтической стоматологии Медицинского института ФГБОУ ВО «Санкт-Петербургский государственный университет», Санкт-Петербург, Россия;

Tumavova S.A., PhD, associate Professor, Department of Therapeutic Dentistry, St. Petersburg State University, St. Petersburg, Russian Federation;

Туманова С.А., к.м.н., доцент, доцент Кафедры терапевтической стоматологии Медицинского института ФГБОУ ВО «Санкт-Петербургский государственный университет», Санкт-Петербург, Россия;

Suvorov A.N., DMS, Corresponding Member of the Russian Academy of Sciences, Head of the Fundamental Problems of Medicine and Medical Technologies Department, St. Petersburg State University, St. Petersburg, Russian Federation; Head of the Molecular Microbiology Department, Institute of Experimental Medicine, St. Petersburg, Russian Federation;

Суворов А.Н., д.м.н., член-корр. РАН, зав. Кафедрой фундаментальных проблем медицины и медицинских технологий Медицинского института ФГБОУ ВО СПбГУ, Санкт-Петербург, Россия; зав. Отделом молекулярной микробиологии ФГБНУ «Институт экспериментальной медицины», Санкт-Петербург, Россия;

Блок 3. Метаданные статьи

HETEROGENEITY OF VIRULENCE FACTORS AMONG *PORPHYROMONAS GINGIVALIS* CLINICAL ISOLATES FROM PATIENTS WITH CHRONIC GENERALIZED PERIODONTITIS

ГЕТЕРОГЕННОСТЬ ФАКТОРОВ ВИРУЛЕНТНОСТИ СРЕДИ КЛИНИЧЕСКИХ ИЗОЛЯТОВ *PORPHYROMONAS GINGIVALIS*, ВЫДЕЛЕННЫХ ОТ ПАЦИЕНТОВ С ХРОНИЧЕСКИМ ГЕНЕРАЛИЗОВАННЫМ ПАРОДОНТИТОМ

Сокращенное название статьи для верхнего колонтитула:

HETEROGENEITY OF *P. GINGIVALIS*

ГЕТЕРОГЕННОСТЬ *P. GINGIVALIS*

Keywords: chronic generalized periodontitis, heterogeneity, virulent genotype, *P. gingivalis*, fimbriae, cysteine proteases.

Ключевые слова: хронический генерализованный пародонтит, гетерогенность, вирулентный генотип, *P. gingivalis*, фимбрии, цистеиновые протеазы.

Оригинальные статьи.

Количество страниц текста – 9,

количество таблиц – 2,

количество рисунков – 5.

26.09.2024

СПИСОК ЛИТЕРАТУРЫ

1	Фукс Е. И., Карева Ю.А., Гализина О.А., Таболина Е.С. Современные аспекты этиологии и патогенеза заболеваний пародонта // Рос. мед.-биол. вестн. им. акад. И.П. Павлова. 2013. № 3. С. 153–159.	Fuchs E. I., Kareva Yu.A., Galizina O.A., Tabolina E.S. Modern aspects of the etiology and pathogenesis of periodontal diseases. <i>Russian Medical Biol. Bulletin named after academician I.P. Pavlov.</i> , 2013, no. 3, pp. 15-39.	https://cyberleninka.ru/article/n/sovremennye-aspekty-etologii-i-patogeneza-zabolevaniy-parodonta
2	Царев В. Н., Е. Н. Николаева, Е. В. Ипполитов. Пародонтопатогенные бактерии – основной фактор возникновения и развития пародонтита // Журн.	Tsarev V. N., Nikolaeva E. N., Ippolitov E. V. Periodontopathogenic bacteria – the main factor in the occurrence and development of periodontitis. <i>Journal. Microbiol.</i> , 2017, no. 5, pp. 101-102.	https://doi.org/10.36233/0372-9311-2017-5-101-112

	микробиол. 2017. № 5. С. 101–112.		
3	Ally N., Whisstock J.C., Sieprawska-Lupa M., Potempa J., Le Bonniec B.F., Travis J., Pike R.N. Characterization of the specificity of arginine-specific gingipains from Porphyromonas gingivalis reveals active site differences between different forms of the enzymes. <i>Biochemistry</i> , 2003, vol. 42, no. 40, pp. 11693-11700.		doi: 10.1021/bi0349726.
4	Amano A., Kuboniwa M., Nakagawa I., Akiyama S., Morisaki I., Hamada S. Prevalence of specific genotypes		doi: 10.1177/00220345000790090501.

	of <i>Porphyromonas gingivalis</i> <i>fimA</i> and periodontal health status. <i>J. Dent. Res.</i> , 2000, vol. 79, no. 9, pp. 1664-1668.		
5	Bostanci N., Belibasakis G.N. <i>Porphyromonas gingivalis</i> : an invasive and evasive opportunistic oral pathogen. <i>FEMS Microbiol. Lett.</i> , 2012, vol. 333, no. 1, pp. 1-9.		doi: 10.1111/j.1574-6968.2012.02579.x.
6	Enersen M., Nakano K., Amano A. <i>Porphyromonas gingivalis</i> fimbriae. <i>J. Oral. Microbiol.</i> , 2013, vol. 5, 10.3402/jom.v5i0.20265.		doi:10.3402/jom.v5i0.
7	Evans R.T., Klausen B., Ramamurthy N.S., Golub L.M.,		doi: 10.1016/0003-9969(92)90115-o.

	Sfintescu C., Genco R.J. Periodontopathic potential of two strains of Porphyromonas gingivalis in gnotobiotic rats. <i>Arch. Oral. Biol.</i> , 1992, vol. 37, no. 10, pp. 813-819		
8	Haffajee A.D., Socransky S.S., Patel M.R., Song X. Microbial complexes in supragingival plaque. <i>Oral. Microbiol. Immunol.</i> , 2008, vol. 23, no. 3, pp. 196-205.		doi: 10.1111/j.1399-302X.2007.00411.x.
9	Hajishengallis G., Darveau R.P., Curtis M.A. The keystone-pathogen hypothesis. <i>Nat. Rev. Microbiol.</i> , 2012, vol. 10, no. 10, pp. 717-725		doi: 10.1038/nrmicro2873.

10	Hajishengallis G., Lamont R.J. Beyond the red complex and into more complexity: the polymicrobial synergy and dysbiosis (PSD) model of periodontal disease etiology. <i>Mol. Oral Microbiol.</i> , 2012, vol. 27, no. 6, pp. 409–419.		doi: 10.1111/j.2041-1014.2012.00663.x.
11	<u>Hajishengallis G., I Diaz P.</u> <i>Porphyromonas gingivalis</i> : Immune subversion activities and role in periodontal dysbiosis. <i>Curr. Oral Health. Rep.</i> , 2020, vol. 7, no. 1, pp. 12-21.		doi: 10.1007/s40496-020-00249-3.
12	How K.Y., Song K.P., Chan K.G. <i>Porphyromonas gingivalis</i> : An Overview of Periodontopathic	-	doi: 10.3389/fmicb.2016.00053

	Pathogen below the Gum Line. <i>Front. Microbiol.</i> , 2016, vol. 7, p. 53.		
13	Igboin C.O., Griffen A.L., Leys E.J. Porphyromonas gingivalis strain diversity. <i>J. Clin. Microbiol.</i> , 2009, vol. 47, no. 10, pp. 3073-3081.		doi: 10.1128/JCM.00569-09.
14	Inaba H., Nakano K., Kato T., Nomura R., Kawai S., Kuboniwa M., Ishihara K., Ooshima T., Amano A. Heterogenic virulence and related factors among clinical isolates of Porphyromonas gingivalis with type II fimbriae. <i>Oral Microbiol. Immunol.</i> , 2008, vol. 23, no. 1, pp. 29-35.		doi: 10.1111/j.1399-302X.2007.00386.x.

15	Kuboniwa M., Lamont R.J. Subgingival biofilm formation. <i>Periodontol. 2000, 2010, vol. 52, no. 1, pp. 38-52.</i>		doi: 10.1111/j.1600-0757.2009.00311.x.
16	Laine M.L., van Winkelhoff A.J. Virulence of six capsular serotypes of Porphyromonas gingivalis in a mouse model. <i>Oral. Microbiol. Immunol., 1998, vol. 13, no. 5, pp. 322-532.</i>		doi: 10.1111/j.1399-302x.1998.tb00714.x.
17	Marsh F.D. Dental plaque as a biofilm and a microbial community - implications for health and disease. <i>BMC Oral Health., 2006, vol. 6 (Suppl 1), p. S14.</i>		doi: 10.1186/1472-6831-6-S1-S14.

18	Mikhaylova E.S., Lashchenov P.V., Koroleva I.V. Clinical and microbiological assessment of periodontal tissues in patients with type 2 diabetes mellitus. <i>J. of Intern. Pharm. Research.</i> , 2019, vol. 11, no. 4, pp. 831-840.		http://www.ijprjournals.com
19	Missailidis C.G., Umeda J.E., Ota-Tsuzuki C., Anzai D., Mayer M.P. Distribution of <i>fimA</i> genotypes of <i>Porphyromonas gingivalis</i> in subjects with various periodontal conditions. <i>Oral Microbiol. Immunol.</i> 2004, vol. 19, pp. 224–229.		doi: 10.1111/j.1399-302X.2004.00140.x.
20	Mysak J., Podzimek S., Sommerova P., Lyuya-Mi. Y.,	-	doi: 10.1155/2014/476068.

	Bartova J., Janatova T., Prochazkova J., Duskova J. Porphyromonas gingivalis: major periodontopathic pathogen overview. <i>J. Immunol. Res.</i> , 2014, vol. 2014, p. 476068.		
21	Olsen I., Lambris J.D., Hajishengallis G. Porphyromonas gingivalis disturbs host-commensal homeostasis by changing complement function. <i>J. Oral Microbiol.</i> , 2017, vol. 9, no. 1, p. 1340085.		doi: 10.1080/20002297.2017.1340085.
22	Ozmeriç N., Preus N.R., Olsen I. Genetic diversity of Porphyromonas gingivalis and its	-	doi: 10.1080/000163500429190.

	possible importance to pathogenicity. <i>Acta Odontol. Scand.</i> , 2000, vol. 58, no. 4, pp. 83-87.		
23	Rodrigues R.S., Silveira V.R., Rego R.O. Analysis of Porphyromonas gingivalis fimA genotypes in severe periodontitis patients. <i>Braz. Oral Res.</i> , 2020, vol. 34, p. e090.		doi: 10.1590/1807-3107bor-2020.vol34.0090.
24	Socransky S.S., Haffajee A.D., Cugini M.A., Smith C., Kent R.L. Jr. Microbial complexes in subgingival plaque. <i>J. Clin. Periodontol.</i> , 1998, vol. 25, no. 2, pp. 134-144.		doi: 10.1111/j.1600-051x.1998.tb02419.x.

25	Yoshino T., Laine M.L., van Winkelhoff A.J., Dahl G. Genotype variation and capsular serotypes of <i>Porphyromonas gingivalis</i> from chronic periodontitis and periodontal abscesses. <i>FEMS Microbiol. Lett.</i> , 2007, vol. 270, no. 1, pp. 75–81.	-	DOI: 10.1111/j.1574 6968.2007.00651.x
26	Umeda J.E., Missailidis C., Longo P.L., Anzai D., Wikström M., Mayer M.P. Adhesion and invasion to epithelial cells by fimA genotypes of <i>Porphyromonas gingivalis</i> . <i>Oral Microbiol. Immunol.</i> , 2006, vol. 21, no. 6, pp. 415-419.		doi: 10.1111/j.1399-302X.2006.00312.x.

27	Wang P.L., Ohura K. Porphyromonas gingivalis lipopolysaccharide signaling in gingival fibroblasts-CD14 and Toll-like receptors. <i>Crit. Rev. Oral Biol. Med.</i> , 2002, vol. 13, no. 2, pp. 132-142.		doi: 10.1177/154411130201300204.
28	Xu W., Zhou W., Wang H., Liang S. Roles of Porphyromonas gingivalis and its virulence factors in periodontitis. <i>Adv. Protein. Chem. Struct. Biol.</i> , 2020, vol. 120, pp. 45-84.	-	doi:10.1016/bs.apcsb.2019.12.001.