

**FIRST REPORT OF CLASS 1 AND CLASS 2 INTEGRONS IN
QUINOLONES RESISTANT *KLEBSIELLA PNEUMONIAE* ISOLATES
FROM NAJAF, IRAQ**

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Abstract

Background: Quinolone-resistant *Klebsiella pneumoniae* is considered a serious global threat. However, little is known regarding their multidrug resistance (MDR) and the role of integron classes in this phenotype. The present study was conducted to investigate the antimicrobial susceptibility and prevalence of class 1 and 2 integrons in quinolone resistance clinical isolates of *K. pneumoniae* from Najaf, Iraq patients.

Methods: A total of 109 *K. pneumoniae* were isolated, the quinolone-resistance isolates were selected for antibiotic resistance test as well as presence of class I and II integron assessed by PCR.

Results: Of the 109 *K. pneumoniae* isolates tested, 74 (67.8%) were shown resistant to quinolone and selected for further studies. Among the clinical specimens, a total of 40 (54%) and 30 (40.5%) of quinolones resistant isolates, 47 (63.5%) isolates were MDR, while 23 (31%) were considered as XDR, and PDR isolates were identified in 4 (5.4%) isolates resistant to all agents in all antimicrobial categories tested. Among the 74 quinolone-resistant *K. pneumoniae* isolates the class 1 integron was found in 41(55.4%), whereas it was detected in only 10 (28.5%) of quinolone susceptible *K. pneumoniae* isolates.

Conclusion: quinolone resistance becomes one of leading concern in global public health. Findings of this study clearly and obviously show that resistance to this antibiotic agent is associated with the presence of class 1 integrons suggesting that integron may assist forward the spread of quinolone-resistance in Najaf. A serious threat to human health may associate with quinolone-resistant bacteria among worldwide.

Keywords:

1 *Klebsiella pneumoniae* is an important opportunistic pathogen associated with
2 nosocomial infections and one of the leading causes of many diseases, and become
3 public health concern especially when characterized as multidrug resistance. due
4 to its responsibility of treatments failure^[1]. Transfer of antibiotic resistance genes
5 between different species of bacteria is associated with mobile DNA elements such
6 as transposons and plasmids. recently, a significant part of resistance genes occur
7 in mobile genetic elements of Gram-negative bacilli, have been detected in DNA
8 elements called integrons^[2]. Integrons encode the antibiotic resistance genes
9 through site-specific recombination and are capable of capturing, integrating and
10 mobilizing gene cassettes^[3]. Various species of Gram negative bacteria that
11 conducted from hospital environments can carry integrons^[4]. The definition of
12 integron based on their respective integrase (*intI*) genes, which located in the 5'
13 conserved segment (5'CS)^[3]. This mobile genetic element also contains a 3'
14 conserved segment that carries the ethidium bromide and quaternary ammonium
15 resistance gene (*qacEΔ1*) and sulfonamide resistance gene (*sulI*), which confer
16 resistance to ethidium bromide and quaternary ammonium compounds and to
17 sulfonamide, respectively transfer^[4]. The nucleotides sequence of integron gene
18 revealed five class this mobile genetic element. Integron class 1 and 2 are
19 frequently detected in clinical isolates of *Enterobacteriaceae*, including *K.*
20 *pneumoniae*^[4]. This study was conducted to investigate the antimicrobial
21 susceptibility and prevalence of class 1 and 2 integrons in quinolone resistance of
22 *K. pneumoniae* isolates from Najaf, Iraq patients.

23 **Materials and Methods**

24 **Bacterial Isolates**

25 A total of 1590 clinical specimens, including urine, burn wound seminal
26 fluid, wound abscesses and sputum were collected from two teaching hospitals in
27 Najaf (Al-Sader Medical City, Al-Hakeem General Hospital, and Al-Zahra

28 Maternity and Children) from December 2012 till June 2013. Among of these 109,
29 non-duplicated *K. pneumoniae* were collected. The isolates were collected from
30 urine (n=59) followed by burn wound (n=46), seminal fluid (n=2), wound
31 abscesses (n=1) and sputum (n=1).

32 **Detection of Quinolones Resistant Phenotype:**

33 All *K. pneumoniae* isolates were classified as quinolones resistant according
34 to susceptibility or resistant to nalidixic acid and ciprofloxacin antibiotics
35 (Cypress, Belgium), and confirmed by MIC Strip (Liofilchem, Italy). According to
36 the CLSI breakpoint criteria, the MIC standard for ciprofloxacin resistance was \geq
37 4 $\mu\text{g/mL}$, and $> 32 \mu\text{g/mL}$ for nalidixic acid, according to the CLSI breakpoint
38 criteria.

39 **Antibiotic Susceptibility Phenotype:**

40 Antibiotics disks (Cypress, Belgium) were used to test the susceptibility of
41 quinolones resistant *K. pneumoniae* isolates, using the Kirby-Bauer method
42 according to CLSI guidelines ^[5]: Amoxicillin (25 μg), piperacillin (25 μg),
43 amoxicillin-clavulanic acid (30 μg), ampicillin-sulbactam (20 μg), piperacillin-
44 tazobactam (10 μg), ticarcillin-clavulanic acid (85 μg), cefotaxime (30 μg),
45 ceftazidime (30 μg), ceftriaxone (30 μg), cefepime (30 μg), ceftazidime (30 μg),
46 aztreonam (30 μg), imipenem (10 μg), meropenem (10 μg), nalidixic acid (30 μg),
47 ciprofloxacin (5 μg), gatifloxacin (5 μg), levofloxacin (5 μg), lomefloxacin (10
48 μg), moxifloxacin (5 μg), norfloxacin (10 μg), ofloxacin (5 μg), amikacin (30 μg),
49 tobramycin (10 μg), gentamicin (10 μg), kanamycin (30 μg), netilmicin (30 μg),
50 chloramphenicol (30 μg), sulfamethoxazole (50 μg), trimethoprim (5 μg). The
51 ATCC standard strain *E. coli* (ATCC 25922) was used as a positive control.

52 **Detection of class 1 and 2 integrons by PCR**

53 The *K. pneumoniae* DNA was extracted as described previously by Cheng
54 and Jiang^[6], after which the DNA samples were used as a source of templates for
55 the polymerase chain reaction (PCR) amplification. The *intI1* and *intI2* genes were
56 amplified by PCR using primers obtained from bioneer (Daejeon, South Korea)
57 listed in table 1. The PCR amplifications were performed in a 20 µl reaction
58 mixture including 10 µl KAPA TaqReadyMix mixture (Kapa Biosystems,
59 Massachusetts, US), 25 pmol of each primer of the single gene, 5 µl of genomic
60 DNA, and nuclease-free water to complete the volume. The PCR amplification
61 was performed in Tprofessional thermal cycler (Biometra, Germany) in the
62 following sequence: 5 min at 94°C, followed by 35 and 30 cycles for integron class
63 1 and class 2 respectively of 30 s each at 94°C (30 s min at 55°C for integron class
64 1 and 1 min at 62°C for integron class 2), 1 min at 72°C, and a final extension step
65 at 72°C for 10 min.

66 Using gel electrophoresis (Biometra, Germany), the PCR products were
67 separated in 1.5% agarose gels with ethidium bromide stained and visualized with
68 gel documentation system (Biometra, Germany).

69 **Statistical Analysis:**

70 Chi-square(χ^2) and Fisher's exact test were used to determine the relation
71 between the presence of integrons and antibiotic resistance in SPSS software
72 (SPSS 16, USA). A P value of < 0.05 was considered as statistically significant.

73 **Results:**

74 Of the 109 *K. pneumoniae* isolates tested, 74 (67.8%) were shown resistant
75 to quinolone was chosen for further studies. Among the clinical specimens, a total
76 of 40 (54%) and 30 (40.5%) of quinolones resistant isolates were obtained from
77 urine and burn wound respectively, while the residual isolates were recovered from
78 seminal fluid 2 (2.7%), wound abscesses and sputum 1 (1.3% each) (table 2).

79 The 74 *K. pneumoniae* isolate had shown resistance to nalidixic acid and/or
80 ciprofloxacin antibiotics were involved in this study. Among the 31 detected
81 antibiotics, the highest drug resistance rate were ampicillin and amoxicillin (100%)
82 and similarly for amoxicillin-clavulanic acid, while, the lowermost drug resistance
83 rate were 36.4% for imipenem and gatifloxacin, 37.8% Meropenem, Amikacin
84 and 39.1% for Chloramphenicol (table 3).

85 According to Magiorakos *et al.*^[8], of the 74 QRKP isolates, 47 (63.5%)
86 isolates were MDR, while 23 (31%) were considered as XDR, that susceptible to
87 two or fewer antimicrobial categories and PDR isolates were identified in 4 (5.4%)
88 isolates that resistant to all agents Known (Table 4).

89 The present study indicate that integrons are widespread in *K. pneumoniae*
90 isolates. Among the 109 isolates, 51 (46.7%) were observed to have integrons,
91 whereas, no class 2 integrons were detected. Among the 74 QRKP isolates the
92 class 1 integron present in 41(55.4%0), while present in only 10(28.5%) of
93 quinolone susceptible *K. pneumoniae* isolates, and were found strongly associated
94 with QRKP isolates ($p=0.008$). class 1 integron was presented in all XDR and PDR
95 and less existent in MDR isolates (Table 5).

96 Discussion

97 The emerging quinolone-resistant clinical isolates and multidrug-resistant *K.*
98 *pneumoniae* strains have been increased among clinical isolates in worldwide and
99 become a serious therapeutic challenge. Integron has been recognized as one of the
100 major sources of genes that responsible for antimicrobial resistance and is
101 suspected to be a source of resistance genes to *Enterobacteriaceae*. Our study
102 analyzed 109 *K. pneumonaie* clinical isolates obtained from hospitals in Najaf,
103 Iraq, comprehensively for possession of integrase genes in quinolone and
104 multidrug resistance isolate, and the association between them.

105 The results of this study indicated that 74 (67.8%) of *K. pneumoniae* isolates
106 had displayed quinolone resistance, these results were higher than studies from
107 other countries ^[9,10]. Among these 74 quinolone resistance isolates, high resistance
108 to penicillins and generations of cephalosporins were observed, being in the range
109 of (97-100%) and (55.4-89.1%) respectively, which is quite high. The present
110 results also revealed the resistant pattern to fluoroquinolone a range of (36-87%),
111 in addition to other antibiotic drugs, which is relatively high. Similar results were
112 revealed previously in United States ^[11].

113 The present study demonstrated an unexpectedly high rate of MDR isolates
114 in present antibiotic resistance profile that represents 63.5% in addition 31% where
115 XDR, and extraordinary 5.4% where PDR. Maybe the increased use of antibiotics
116 during recent years in an uncontrolled way could be the cause of this. Continued
117 used of certain antibiotics also supports the selection of certain resistance elements
118 and promotes the perseverance of MDR bacteria ^[12]. Similar result was established
119 in previously in Iran ^[2].

120 Till now, there are no published studies estimated the presence of integrons
121 (class 1 and 2) in QRKP isolates from the Najaf, Iraq. Therefore, the present study
122 first reported the class 1 integron were associated with 41(55.4%0) of QRKP. A
123 study from Iran ^[13] reported integron present in 66.6% of *K. pneumoniae* isolates.
124 Similar finding previously reported revealed this association of integron and
125 QRKP isolates. This association may be due to increasing the rate of mutation of
126 the bacterial cell, and/or the presence of resistance genes on integrons that
127 responsible for decreased the permeability of membrane or improved efflux
128 pump^[14]. Coexistence of quinolone resistance with the presence of integron is an
129 important public health problem and requests for incessant surveillance,
130 monitoring, and adjustment of the antibiotic use policies.

131 In conclusion, quinolone resistance becomes the one of leading concern in
132 global public health. Findings of this study clearly obviously show that resistance
133 to this antibiotics is associated with the existence of class 1 integrons suggests that
134 integron may be assisting forward the spread of quinolone-resistant in Najaf. A
135 serious threat to human health may associate with quinolone resistance bacteria
136 among worldwide. Additional studies of integrons are required to understand the
137 mechanisms of possessing of MDR genes in quinolone resistance clinical isolates.

138 **Conflict of interest**

139 None to declare.

140

TABLES

Table 1: Primers Used to Detect Integron Type I and Type II in this Study

Target	Primer name	Primer sequence (5'-3')	References
<i>intI1</i>	Int1F	CAGTGGACATAAGCCTGTTC	7
	Int1R	CCCGAGGCATAGACTGTA	
<i>intI2</i>	intI2F	CACGGATATGCGACAAAAAGGT	7
	intI2R	GTAGCAAACGAGTGACGAAATG	

Table 2: Frequency of quinolone resistant *K. pneumoniae* isolates in clinical specimen.

Clinical specimen	Total (%) of <i>K. pneumoniae</i> isolates	No (%) of isolates had quinolone resistant
Urine (n= 1340)	59 (4.4)	40 (67.8)
Burn wound (n= 225)	46 (20.4)	30 (65.2)
Sputum (n= 7)	1 (14.3)	1 (100)
Wound abscess (n= 7)	1 (14.3)	1 (100)
Seminal fluid (n= 11)	2 (18.2)	2 (100)
Total (n= 1590)	109 (6.9)	74 (67.8)

Table 3: Antibiotic susceptibility pattern expressed by quinolone resistance *K. pneumoniae* isolates (n= 74)

Antibiotic	No. (%) of isolates showed:	
	Resistance	Susceptible
Ampicillin	74 (100)	0(0)
Amoxicillin	74 (100)	0(0)
Piperacillin	72(97.3)	2(2.7)
Amoxicillin-clavulanic acid	74 (100)	0(0)
Ampicillin-sulbactam	70 (94.6)	4 (5.4)
Piperacillin-tazobactam	57(77)	17 (22.9)
Ticarcillin-clavulanic acid	65(87.8)	9 (12.2)
Cefotaxime	66(89.1)	8 (10.8)
Ceftazidime	64(86.4)	10 (13.5)
Ceftriaxone	64(86.4)	10 (13.5)
Cefepime	55(74.3)	19 (25.7)
Cefoxitin	41(55.4)	33 (44.6)
Aztreonam	63(85.1)	11 (14.8)
Imipenem	27(36.4)	47 (63.5)
Meropenem	28(37.8)	46 (62.2)
Nalidixic acid	66(89.1)	9 (12.2)
Ciprofloxacin	65(87.8)	10 (13.5)
Gatifloxacin	27(36.4)	47 (63.5)
Levofloxacin	33(44.5)	41 (55.4)
Lomefloxacin	64(86.4)	10 (13.5)
Moxifloxacin	60(81)	15 (20.2)
Norfloxacin	39(52.7)	35 (47.3)
Ofloxacin	39(52.70)	35 (47.3)
Amikacin	28(37.8)	46 (62.1)
Gentamicin	43(58.1)	31 (41.9)
Kanamycin	44(59.4)	30 (40.5)
Netilmicin	30(40.5)	44 (59.4)
Tobromycin	57(77)	17 (22.9)
Chloramphenicol	29(39.1)	45 (60.8)

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Sulfamethoxazole	64(86.4)	10 (13.5)
Trimethoprim	66 (89.2)	8 (10.8)

Table 4: Multiple antibiotic resistance phenotypes of 74 quinolone resistant *K. pneumoniae* isolates

Resistance Pattern No.(%)	No. of antibiotic categories (n=11)	No. of resistance isolates (%)	Isolate code No.
MDR 47(63.5)	3	5(6.7)	Kp 10, 21, 31, 35, 65
	4	1(1.3)	Kp 33
	5	2(2.7)	Kp 6, 32
	6	4(5.4)	Kp 9, 50, 97, 98
	7	12(16.2)	Kp 4, 14, 20, 36, 47, 68, 91, 94, 105, 117, 118, 123
	8	14(18.9)	Kp 3, 7, 46, 62, 48, 72, 76, 74, 73, 83, 85, 108, 115, 110
	9	9(12.1)	Kp 22, 49, 67, 77, 102, 111, 114, 120, 130
XDR 23(31)	10	12(16.2)	Kp 1, 15, 45, 55, 56, 57, 58, 60, 66, 93, 95, 128
	11	11(14.8)	Kp 12, 18, 23, 37, 38, 69, 84, 87, 100, 103, 113
PDR 4(5.4)	11	4(5.4)	Kp 63, 64, 104, 92

Table 5: Frequency of class 1 integron gene in 74 quinolone resistance *K. pneumoniae* isolates

Type of Resistance	Quinolone resistant			P Value
	intI +	intI -	Total	
MDR	14	33	47	0.012
XDR	23	0	23	0.00
PDR	4	0	4	0.00
	41	33	74	

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Conflict of interest

- All authors have no conflict of interest

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